ACS APPLIED MATERIALS & INTERFACES

www.acsami.org

Research Article

SINGLET OXYGEN

SENSITIZATION

non-radiative ENERGY TRANSFER

¹ Energy Partitioning in Multicomponent Nanoscintillators for ² Enhanced Localized Radiotherapy

³ Valeria Secchi, Francesca Cova, Irene Villa, Vladimir Babin, Martin Nikl, Marcello Campione, ⁴ and Angelo Monguzzi^{*}



⁷ sensitizers (PSS) for cytotoxic reactive oxygen species (ROS) have been proposed 8 in the last decade as coadjuvant agents for radiotherapy of cancer. They have been 9 designed to make scintillation-activated sensitizers for ROS production in an 10 aqueous environment under exposure to ionizing radiations. However, a detailed 11 understanding of the global energy partitioning process occurring during the 12 scintillation is still missing, in particular regarding the role of the non-radiative 13 energy transfer between the nanoscintillator and the conjugated moieties which is 14 usually considered crucial for the activation of PSs and therefore pivotal to

15 enhance the therapeutic effect. We investigate this mechanism in a series of PS-functionalized scintillating nanotubes where the non-16 radiative energy transfer yield has been tuned by control of the intermolecular distance between the nanotube and the conjugated 17 system. The obtained results indicate that non-radiative energy transfer has a negligible effect on the ROS sensitization efficiency, 18 thus opening the way to the development of different architectures for breakthrough radiotherapy coadjuvants to be tested in clinics.

19 KEYWORDS: radiotherapy, scintillators, energy transfer, singlet oxygen, nanomaterials

1. INTRODUCTION

20 Looking at recent research, it is clear that nanotechnology can 21 play an important role in the biomedical science thanks to the 22 successful development and use of nanoparticles for 23 theranostics, diagnostics, monitoring of specific injured tissues 24 or organs, and for the improvement in some traditional 25 therapeutic treatments.^{$1-3^{-1}$} This is mainly due to the 26 advantages of nanomaterials with respect to bulk systems, 27 such as the facile surface functionalization, the composition 28 versatility, and their tailorable optical and magnetic properties, 29 which allow them to respond to the specific demands of the 30 targeted application and use. Consequently, a huge family of 31 nanomaterials, such as metallic and semiconductor nano-32 particles, metal/lanthanide oxides, and organic and hybrid 33 systems, has been developed to be used in advanced diagnostic 34 and imaging techniques, drug delivery strategies, or innovative 35 therapeutic approaches against cancer and other deadly 36 diseases,⁴⁻⁷ as demonstrated by the increasing number of 37 nanosystems approved by the Food and Drug Administration 38 agency.

For example, we can observe an increasingly larger use of 40 radioluminescent nanoparticles, i.e., nanoscintillators, able to 41 absorb and convert the ionizing radiation (*x*- or γ -rays) into a 42 large number of UV–visible photons, which are exploitable to 43 boost the efficacy of diagnostic techniques for preclinical 44 mapping, intraoperative imaging, radiation dosimetry, and, 45 importantly, as efficient coadjutants in oncological therapies.^{8–11} The search for innovative therapies to surpass state- 46 of-the-art treatments is indeed still highly persistent. The 47 standard cancer treatment options, represented by chemo- 48 therapy, radiotherapy, and surgery, are still associated with 49 systemic side effects, disease recurrence, and drug/radio 50 resistance of malignant cells. Among them, the radiotherapy 51 exploits the effect of the ionizing radiation that directly 52 damages the cellular DNA or indirectly forms cytotoxic 53 reactive oxygen species (ROS), such as hydroxyl radicals and 54 singlet oxygen (SO), upon interaction with the intracellular 55 aqueous environment.^{7,12} However, radiotherapy is strongly 56 limited by the maximum radiation dose that can be given to a 57 tumor mass without incurring significant injuries to the 58 adjacent tissues or organs.¹³ Modern approaches envisage the 59 use of patient-specific dose-delivery plans or short radiation 60 pulses to limit collateral effects,^{14,15} but these strategies does 61 not solve the problem of absolute lack of selectivity of the 62 ionizing radiation for the sick tissues. In this regard, the 63 photodynamic therapy (PDT) has been proposed as an 64 alternative to radiotherapy due to its high selectivity and 65

- ray,

- rays

Received: January 18, 2023 Accepted: April 28, 2023



Α

www.acsami.org



Figure 1. (a) Synthesis of multicomponent scintillating nanotubes (NTs) for PDT-enhanced radiotherapy achieved by the use of an SO $({}^{1}O_{2}^{*})$ PS. (b) Photophysics of the sensitization of SO production under exposure to ionizing radiation. The free electrons and holes generated by interaction between the ionizing radiation and the NT recombine directly on the NT and on the PS. The latter is promoted to its excited-state singlet (S_n*) or triplet (T_n*) with a ratio of 1:3. The energy stored in the NT can be therefore transferred by non-radiative energy transfer (ET_{nr}) producing additional PS molecules in the S₁* state. The PS molecules in the S₁* state can subsequently experience intersystem crossing (ISC) that further populates the T₁* state. From PS in the triplet state, the energy is transferred by a second non-radiative energy transfer step to molecular oxygen, which is promoted to its excited singlet state ${}^{1}O_{2}^{*}$. (c) Sketch of ET_{nr} active and ET_{nr} inactive multicomponent scintillating NTs realized by incrementing the intermolecular distance between the NT and the PS molecules.

66 minimal invasiveness.¹⁶ PDT exploits indeed specific photo-67 sensitizer (PS) moieties which are selectively activated only by 68 light in the presence of molecular oxygen in order to produce 69 ROS.

The PDT has been utilized in the clinic for treatments of 71 different cancer types, but despite the excellent results 72 obtained, its clinical use is actually hindered by the shallow 73 tissue penetration of light, especially in the visible spectral 74 window where most of the best PSs absorb the electromagnetic 75 radiation.^{17,18}

An important step forward to overcome both radiotherapy 76 and PDT drawbacks was made in 2006, with the introduction 77 of the concept of energy transducers to transform the energy 78 deposited by X-rays or γ -rays into optical-range lumines-79 cence.¹⁹ The core of this PDT-enhanced radiotherapy is the 80 81 use of luminescent dense nanoscintillators that can interact 82 efficiently with the ionizing radiation achieving also a photon 83 down-conversion into the visible range to activate the PSs, by 84 both radiative and non-radiative energy transfer processes 85 (Figure 1a).²⁰⁻²² The presence of these nanoscintillators 86 allows (i) the promotion of localized energy deposition in the 87 tissue of interest and (ii) the activation of the PDT effect in deep tissues 2^{23-25} by means of a complex energy partitioning 88 89 scheme. To date, diverse classes of inorganic dense nano-90 scintillators have been combined with organic PSs,²⁶ and they 91 have been investigated both in vitro and in vivo.^{12,27} The 92 excellent results obtained demonstrate clearly that this 93 approach results in a synergistic therapeutic effect of 94 radiotherapy and PDT,²⁸⁻³⁰ thanks to the enhanced 95 sensitization of ROS production given by the presence of PS

f1

systems.²⁰ Nevertheless, a complete understanding of the ⁹⁶ energy partitioning that occurs in the scintillation process ⁹⁷ among the dense nanoscintillators, the PDT agent, and the ⁹⁸ biological environment is still lacking. Consequently, the ⁹⁹ general guidelines for the design of optimized nanomaterials to ¹⁰⁰ be tested in a clinical environment are still eagerly required. ¹⁰¹

Here, we studied the role of a non-radiative energy transfer 102 (ET_{nr}) process between the nanoscintillator and the PDT 103 agents in the global energy partitioning mechanism. Parallel to 104 the passive sensitized activation, triggered by the presence of 105 the dense nanoscintillator that enhances the localized release of 106 the ionizing radiation energy (Figure 1b), ET_{nr} is indeed $_{107}$ usually considered a crucial activation pathway for PDT 108 sensitizers,^{12,31} but no direct proof has been given yet.²⁸ 109 Considering that the optimization of ET_{nr} imposes several 110 severe restrictions on the material composition, architecture, 111 and electronic properties in order to couple effectively a 112 nanoscintillator to a PS system, it is therefore crucial to 113 understand its effective role in the global sensitization process 114 for the design of optimized radiotherapy coadjutants. In 115 particular, we investigate a series of scintillating nanotubes 116 (NTs) functionalized with a model conjugated PS for singlet 117 oxygen (SO). The ET_{nr} rate and yield have been finely tuned 118 by controlling the intermolecular distance between the NT and 119 the chemically coupled PS molecules. The results obtained 120 suggest that ET_{nr} has a minor role in the SO sensitization $_{121}$ process, thus opening the way to the development of different 122 architectures for highly effective radiotherapy coadjuvants to 123 be tested in clinics. 124



Figure 2. (a) Transmission electron microscopy (TEM) image of scintillating chrysotile NTs and their size distribution (inset). (b) On the left, the PL (dashed-dotted line, exc. 250 nm) and radioluminescence (RL, solid line) spectrum of the NT under soft X-ray exposure (dashed-dotted line). On the right, absorption (dashed line) and PL (solid line) spectra of the conjugated chromophore Rhodamine Red C_2 maleimide selected as the model SO PS. (c) Attenuated reflectance FT-IR spectra of NTs and the multicomponent nanoscintillator series obtained by tuning the PS-to-NT intermolecular distance from 5 Å (NT-5*) to 46 Å (NT-46*). The asterisks mark the sample where Rhodamine Red C_2 maleimide is substituted with rhodamine B. Shaded areas mark the characteristic IR mode of the NT (gray at around 1000 and 4700 cm⁻¹) and of the PS (orange, 2300–2500 cm⁻¹). (d) PL of multicomponent nanoscintillators as a function of the NT-to-PS intermolecular distance under UV excitation at 250 nm. (e) PL intensity decay in time recorded at 430 nm under pulsed excitation at 250 nm of NTs and the functionalized NT sample series. The inset is a digital picture of the NT-20 sample under daylight.

2. RESULTS AND DISCUSSION

125 As detailed in the Experimental Methods section, the PS-126 functionalized nanoscintillators have been realized by coupling biocompatible chrysotile NTs to the conjugated chromophore 127 Rhodamine Red C2 maleimide by means of several 128 129 heterobifunctional bridges of different lengths (Figure 1a, see 130 Supporting Information file, Table S1, Supporting Figure S1). 131 The NTs have been synthesized in aqueous solution under 132 hydrothermal conditions in the presence of Mg and Si 133 precursors. We obtained pure chrysotile NTs (Figure S2) of 134 diameter 50 nm and average length 100 nm (Figure 2a) with a 135 blue scintillation and photoluminescence (PL) peaked at 430 $_{136}$ nm (Figure 2b). The external surface of the NTs is brucitic, ³² 137 showing a positive ζ -potential which allows the coulombic interaction with anionic species such as the carboxyl functional 138 group at one end of the bridge ligand series employed (Figure 139 1a). The PS system has been selected because of (i) a suitable 140 energetic resonance between its ground state absorption and 141 the NT scintillation emission (Figure 2b), which allows the 142 occurrence of non-radiative ET_{nr} by both the Dexter and 143 örster mechanisms between the NT and the PS molecules,³³ 144 and (ii) the presence of the maleimide functionality. The latter 145 146 is a crucial point because this functionality allows us to exploit 147 the thiol-maleimide click reaction with the -SH functional group at one end of the NT surface ligand to anchor the PS 148 (Figure 1a), therefore controlling their compositon.^{34,35} So, 149 150 although the resonance with the NT emission is not ideal, the 151 employed PS is the ideal system to perform the designed 152 experiments. In such a way, by varying the length of the 153 connecting ligand, we can tune the rate and yield of ET_{nr} by 154 increasing the intermolecular distance between the NT and PS 155 from 17 to 37 Å (Table S1). The different samples are labeled

as NT-x, where x is the distance between the NT and the PS $_{156}$ expressed in angstroms. It is worth noting that the organic 157 ligand employed is not rigid; so, the considered intermolecular 158 distances are nominal values taken as the reference. To have 159 also very short or very large intermolecular distances of 5 and 160 46 Å, we used as the PS the conjugated chromophore 161 rhodamine B (Figure S1) that possesses the right anionic 162 functionality to be directly anchored on the NT surface (NT- 163 5*)^{36,37} or placed quite far by using polyethylene glycol as the 164 connecting ligand (NT-46*). Considering the typical non- 165 radiative interaction radii and the poor luminescence yield of 166 NTs,³³ in sample NT-5*, the ET_{nr} yield ϕ_{ET}^{nr} should be 167 maximized while minimized in the sample NT-46*. In the 168 latter case, given the limited energetic resonance between NT 169 emission and PS absorption, the contribution of ET_{nr} to the 170 SO sensitization can be for sure neglected and therefore 171 completely decoupled from the other mechanisms involved 172 (Figure 1b). 173

The successful functionalization of the NT surfaces with the 174 heterobifunctional chains and fluorescent PS molecules has 175 been confirmed by means of vibrational and optical spectros- 176 copy experiments. Figure 2c reports the infrared spectra of the 177 bare NTs and the NT-*x* sample series. In all spectra, we can 178 observe the main chrysotile vibrational peaks located at around 179 3700 cm⁻¹ (MgOH stretching) and in the region around 1000 180 cm⁻¹ (Si-O-Mg, Si-O-Si, and Si-O stretching).³⁷ 181 Furthermore, the spectra of the samples from NT-17 to NT-182 37 show clearly the peaks related to the Rhodamine Red C₂ 183 maleimide or to the rhodamine B functionalities (the C=C 184 stretching vibrations at 1628 and 1542 cm⁻¹, the N-C 185 bending at 1291 cm⁻¹, and the C-H stretching in the region 186 around 3000 cm⁻¹).^{38,39} The average number of PS molecules 187



Figure 3. (a) RL spectra of the multicomponent nanoscintillator series as a function of the PS-to-NT intermolecular distance, normalized to the residual PS emission intensity. (b) Scintillation pulses recorded at ca. 620 nm under pulsed X-ray excitation at 40 keV. (c) PL intensity decay at 620 nm under pulsed excitation at 250 nm. (d) Relative increment of the SO concentration as a function of the irradiation time under soft X-rays for the NT-20 sample (4.0 mg/mL, PBS). The SO increment has been monitored by recording the SO optical probe SOSG under simultaneous CW laser excitation at 473 nm (inset). (e) NT-to-PS energy transfer yield (ϕ_{ET}^{nr} , dots), relative scintillation yield ($\bar{\phi}_{scint}^{PS}$) of the PS, and SO relative sensitization ability $\bar{\phi}_{SO}$ after 600 s of exposure to soft X-rays for the multicomponent nanoscintillator series, as a function of the NT-to-PS intermolecular distance. Error bars are put as the mean standard deviation calculated on a N = 3 measurement replica.

188 (n) coupled to each NT has been evaluated by means of 189 optical absorption measurements (Table S1, Figure S1). Under 190 UV excitation at 250 nm, all the functionalized NTs show a multiband PL spectrum (Figure 2d) where a residual NT 191 emission at 430 nm can be observed, even very weak in some 192 193 cases due the occurrence of $\mathrm{ET}_{\mathrm{nr}}.$ The PS fluorescence around 194 600 nm from the Rhodamine Red C₂ maleimide and at 580 nm 195 from the rhodamine B can be clearly distinguished in samples 196 NT-17-NT-37 and samples NT-5* and NT-46*, respectively. 197 No change in the emission properties is observed after keeping 198 NT-*x* in a phosphate buffered saline (PBS) dispersion for up to 199 6 months, thus demonstrating the excellent stability of the 200 synthesized materials. The residual NT PL intensity at 430 nm 201 increases as a function of the NT-to-PS distance, thus 202 suggesting the progressive reduction of $\phi_{\rm ET}^{\rm nr}$ by separating the 203 NT from the PS. $\phi_{\rm ET}^{\rm nr}$ has been quantitatively evaluated by 204 means of time-resolved PL experiments. Figure 2e shows the 205 PL intensity decay in time of the samples monitored at 430 nm 206 as a function of the NT-to-PS intermolecular distance. As 207 expected, the emission decay accelerates by shortening the 208 intermolecular distance that increases the ET_{nr} rate, which 209 becomes competitive with the spontaneous recombination of 210 the NT excited state.³³ Both the bare NTs and the NT-x211 sample series show emission intensity decays with a multi-212 exponential behavior. The characteristic lifetime is calculated 213 as the average emission lifetime $\langle \tau_X \rangle$ (Table S1). The $\phi_{\rm ET}^{\rm nr}$ χ_{214}^{nr} value is then calculated as $\phi_{\rm ET}^{nr} = 1 - \langle \tau_X \rangle / \langle \tau_{\rm NT} \rangle$, where $\langle \tau_{\rm NT} \rangle$ 215 is the average lifetime of the bare NT emission. $\phi_{\text{ET}}^{\text{nr}}$ is reduced

from 70% down to 10% by increasing the NT-to-PS nominal 216 intermolecular distance from 5 to 46 Å (vide infra, Figure 3e). 217 f3 Figure 3a shows the RL spectra of the NT and NT-x sample 218 series powders (16 mg) recorded under steady-state excitation 219 by soft X-rays (Experimental Methods). Similar to the PL 220 spectra, also in this case, we can observe clearly the typical PS 221 luminescence with a residual blue luminescence from NTs. 222 The relative scintillation yield of the PS dyes $\overline{\phi}_{\rm scint}^{\rm PS}$ is taken as $_{223}$ the RL intensity integrated in the PS emission spectral range 224 $(I_{\rm RL}^{\rm PS})$. Notably, the PS scintillation luminescence is slightly red- 225 shifted with respect to the PL spectrum in dispersion due to 226 the enhanced self-absorption of dyes and the possible 227 formation of aggregates in the powder form. Figure 3b 228 shows the corresponding scintillation pulses recorded at the 229 dye emission wavelength by exposing the powders to a pulsed 230 X-ray source (Experimental Methods, Table S2). This 231 experiment has been performed to have a hint on the 232 luminescence properties of the materials under exposure to 233 ionizing radiation. These measurements have been performed 234 on powders because the pulsed X-ray source irradiance is too 235 weak to record reliable signals from the diluted aqueous 236 suspensions employed to generate the SO. For the samples 237 from NT-5* to NT-24, the scintillation pulse lifetime is around 238 3 ns, with no significant differences. The observed values are 239 slightly higher with respect to the corresponding PS PL decay 240 time (Figure S3), in agreement with the possible self- 241 absorption delay effect of the dye on the apparent emission 242 lifetime. On the other side, the formation of low-energy J- 243

244 aggregates is most probably responsible for the longer-emission 245 component in the scintillation of samples NT-30, NT-37, and 246 NT-46* and for the fast quenching observed in samples NT-37 247 and NT-46*.³⁶ The more marked presence of aggregates in 248 these samples agrees with the presence of long and more 249 flexible surface ligands, which allows the connected dyes to 250 interact more freely with respect to the NT functionalized with 251 shorter ligands, which keep the dyes far enough to limit 252 detrimental intermolecular interactions, especially in the 253 powder form.

On the other hand, considering that the NT will be used in 254 255 diluted aqueous dispersion, for a quantitative and reliable 256 comparison between the different samples, we have measured 257 the recombination kinetics of PS PL in the aqueous dispersion where they will be used to sensitize the SO production. Figure 258 3c reports the NT-x PL intensity decay with time recorded at 259 260 610 nm under pulsed laser excitation at 250 nm (Experimental 261 Methods). In all cases, we observe an average decay time 262 shorter from the one observed for the single chromophore in 263 diluted solution (Figure S4), again most probably due to the 264 presence of quenching J-aggregates on the NT surface, but 265 there is no evident coherent trend. In some cases, the emission 266 intensity decays as a single exponential function in a time 267 shorter than the spontaneous one (1.97 ns for Rhodamine Red 268 C₂ maleimide and 2.71 ns for rhodamine B, Figure S4), while 269 in some cases, we observe also a multi-exponential decay 270 behavior (N-17, NT-19, and NT-37). Nevertheless, independ-271 ently from its origin, the observed partial emission quenching 272 suggests that upon functionalization, the PL yield $\phi_{
m pl}^{
m PS}$ (Experimental Methods, Table S3) of the PS is reduced. 273 274 This means that the recombination properties of the PS singlet 275 excited state are modified upon binding to NTs, including the 276 intersystem crossing (ISC) rate that populates the triplet state 277 from which the SO sensitization occurs by energy transfer to 278 the ground-state molecular oxygen in solution (Figure 1b)¹⁷ 279 and the triplet state lifetime that also affects the transfer to 280 molecular oxygen. Therefore, also, the SO generation efficiency 281 can be affected. This effect has been taken into account (vide 282 infra) in order to have a reliable relative comparison of the 283 samples ϕ_{SO} .

 $\phi_{
m SO}$ has been directly observed by the measurement of the 284 285 relative SO production efficiency under soft X-ray exposure. 286 Figure 3c reports the evolution of the SO concentration in PBS 287 dispersion of NT-20, as an example, which has been monitored 288 in situ by using the SO Sensor Green (SOSG, Experimental 289 Methods) as an optical probe.²⁰ The SOSG PL intensity is 290 proportional to the concentration of SO;40 thus, upon its 291 selective excitation, we can compare the evolution of the SO 292 concentration as a function of time (inset of Figures 3c and 293 S5). Specifically, ϕ_{so} is defined here as the relative increment ²⁹⁴ of the SO concentration and calculated as ²⁹⁵ $\phi_{SO} = 100 \times [I_{pl}^{SOSG}(t) - I_{pl}^{SOSG}(0)]/I_{pl}^{SOSG}(0)$. All the sam-296 ples in the series have been monitored under steady-state X-297 rays exposure up to 600 s which corresponds to a delivered 298 dose of approximately 260 Gy, in glass vials. All the samples 299 show a SO sensitization ability (Figure S4). In order to have a 300 reliable relative comparison, the SO sensitization efficacy is 301 finally calculated as a relative normalized SO sensitization ability $\overline{\phi}_{SO} = \frac{\phi_{SO}}{\langle n \rangle \phi_{p_1}^{PS}}$, thus taking into account the perturbation

ability $\phi_{SO} = \frac{1}{\langle n \rangle \phi_{pl}^{PS}}$ individuality into account the perturbation 303 of the PS observed upon binding that is assumed to modify its 304 ϕ_{pl}^{PS} and thus indirectly the SO ability.⁴¹

The comparative analysis among the observed $\phi_{\rm ET}^{\rm nr},~\overline{\phi}_{\rm scint}^{\rm PS},~_{305}$ and $\overline{\phi}_{\rm SO}$ as a function of surface ligand length is reported in $_{306}$ Figure 3e. It is worth noting that the absorption of the PS 307 molecules in the investigated dispersions is very low (Figure 308 S8), and the NT emission efficiency is very weak (\ll 5%) so 309 that we can exclude a priori a relevant photoexcitation of the 310 conjugated PS by direct absorption of the NT scintillation light 311 (i.e., radiative energy transfer). As discussed above, the $\phi_{
m ET}^{
m nr}$ 312 value decreases by about 1 order of magnitude by moving 313 progressively far away the PS molecules from the scintillating 314 NT, until a $\phi_{\rm ET}^{\rm nr}$ = 10% is observed in the NT-46* sample in 315 agreement with the distance-dependent behavior of the non- 316 radiative ET_{nr} rate. On the other side, both $\overline{\phi}_{\rm scint}^{\rm PS}$ and $\overline{\phi}_{\rm SO}$ show $_{317}$ a substantially constant behavior completely uncorrelated to 318 $\phi_{\rm ET}^{\rm nr}$ thus suggesting that the scintillation light output and the 319 efficiency of the SO sensitizer are independent from the system 320 architecture. Even in the best configuration with $\phi_{\rm ET}^{\rm nr}$ = 70%, no 321 enhancement is observed in the SO production. Similar results 322 are observed by using a different optical probe for the SO 323 formation (Figures S6 and S7). These findings demonstrate 324 therefore the negligible role of ET_{nr} between the nano- 325 scintillator NTs and the PS moiety in activating the SO 326 sensitization ability of multicomponent materials for PDT- 327 enhanced radiotherapy. Moreover, these results confirm 328 experimentally for the first time the output of radiation/matter 329 interaction simulations in nanoscintillators recently pro- 330 posed.⁴² According to the dimension of our nanoscintillators, 331 only a minor fraction of the energy deposited upon interaction 332 of the X-rays with the high-Z elements is stored in the particle 333 itself, while most of the energy is spread around the particle by 334 generating a swarm of secondary charges that can diffuse for 335 distance up to hundreds of nanometers. Thus, even in the best 336 case where $\phi_{\rm ET}^{\rm nr}$ equals unity, the effective boosting of the PDT 337 activity due to the ET_{nr} channel can be only negligible, while 338 the major role in the global energy partitioning process is 339 played by the direct recombination of free charges on the PSs, 340 which is locally sensitized by the presence of the dense 341 nanoscintillator. 342

3. CONCLUSIONS

In conclusion, we successfully realized a series of multi- 343 component nanoscintillators as a model system for PDT- 344 enhanced radiotherapy coadjutants. Their architecture has 345 been finely tailored in order to control the efficiency of the 346 non-radiative energy transfer process between the building 347 blocks of the multicomponent system, namely, the scintillating 348 dense nanoparticle, responsible for the localized interaction 349 with the ionizing radiation, and the attached ROS-sensitizing 350 PS species that enable the PDT. The obtained results 351 demonstrate that the non-radiative energy transfer plays a 352 marginal role in the global energy partitioning process 353 responsible for the evident synergistic effect of radiotherapy 354 and deep-tissue X-ray-activated PDT usually observed during 355 cancer treatment using these materials. This finding has 356 important consequences, by pointing out some new guideline 357 pivotal for the design and realization of optimized multi- 358 component radiotherapy coadjuvants. First, the matching 359 between the electronic transitions of the scintillator and the 360 PS is no more strictly required since the PS is mainly activated 361 by direct recombination of the free charges produced during 362 the primary and secondary interaction events in the 363

Е

366 scintillators and PSs is no more required to maximize the 367 energy transfer rate, thus again significantly relaxing the 368 constraints on the system architectures and avoiding the 369 problems originating from the need for specific control of 370 intermolecular interactions between close-packed species. 371 Third, considering that heaviest elements such as lead could 372 represent a critical issue for their poor biocompatibility, the 373 obtained results indicate that larger but still biocompatible 374 nanoparticles are required to maximize the local radio-375 sensitization effect in tumors. For example, hafnia and/or 376 zirconia nanoparticles 7,42,43 with size up to 100–200 nm can 377 be envisaged. According to the obtained results, the best 378 arrangement for the PS moiety could be, for example, a shell wrapped around the dense nanoparticle with thickness up to 379 380 100 nm, in order to harvest the most of the diffusing charge 381 energy. This design will result in a bigger multicomponent 382 system with still good cellular uptake and delivery in the 383 body^{27,44-47} and a simultaneous good interaction with the 384 ionizing radiations and optimized energy harvesting and 385 partitioning that will potentially lead to a breakthrough 386 increment of the radiotherapy effect even at low doses.

4. EXPERIMENTAL SECTION

387 4.1. Synthesis of Stoichiometric Chrysotile Nanotubes. 388 Chrysotile NTs were synthesized according to a previously used 389 synthetic method.²⁰ A hydrothermal reactor with a 100 cm³ 390 polypropylene vessel was used to carry out the hydrothermal reaction 391 of 1522 mg of Na₂SiO₃ and 764 mg of MgCl₂ in an aqueous solution 392 of NaOH (220 mL 0.4 M) at 250 °C with a run duration of 16 h. The 393 precipitate removed from the solution was repeatedly washed with 394 deionized water before being dried for 3 h at 110 °C.

4.2. Functionalization of Chrysotile Nanotubes with Chains 395 396 of Different Lengths. For the preparation of each sample, 100 mg 397 of NT powder was suspended in 25 mL of PBS and 30 mg of 16-398 mercaptohexadecanoic acid suspended in 25 mL of PBS or 40 mg of 399 11-mercaptoudecanoic acid suspended in 20 mL of PBS or 810 μ L of 400 8 mercaptooctanoic acid, or 600 μ L of 3 mercaptopropionic acid, or 401 120 mg of L-cysteine suspended in 25 mL of PBS , or 620 μ L of 402 thioglycolic acid were added slowly under stirring for 10 min. Samples 403 were centrifuged for 5 min at 6500 rpm. The precipitate removed 404 from the solution was repeatedly washed with deionized water before 405 being dried for 3 h at 50 °C.

4.3. Functionalization of NTs + Variable Length Chain with 406 407 Invitrogen Rhodamine Red C₂ Maleimide. 40 mg of NTs 408 functionalized with chains of different lengths was dispersed in 5 mL 409 of tris(hydroxymethyl)aminomethane (TRIS), and 4 mL of 410 Invitrogen Rhodamine Red C2 maleimide (1.4 mg in 70 mL of 411 TRIS) was added in the solution. Maleimide is a thiol-reactive probe 412 and reacts with thiol groups in a typical thiol-maleimide "click" 413 chemistry reaction to give thioether-coupled products. Samples were 414 centrifuged for 5 min at 6500 rpm. The precipitate removed from the 415 solution was repeatedly washed with deionized water and PBS before 416 being dried for 3 h at 50 °C.

4.4. Functionalization of Chrysotile Nanotubes with Rhod-417 418 amine B. 60 mg of NTs was dispersed in 15 mL of PBS, and 2 mL of 419 rhodamine B (3×10^{-5} M in PBS) was added in the solution. Samples 420 were centrifuged for 5 min at 6500 rpm. The precipitate removed 421 from the solution was repeatedly washed with deionized water and 422 PBS before being dried for 3 h at 50 °C.

4.5. Functionalization of Chrysotile Nanotubes with Rhod-423 424 amine B-PEG2k-COOH (Sigma-Aldrich). 60 mg of NTs was 425 dispersed in 15 mL of PBS, and 2 mL of rhodamine B- PEG2k-426 COOH (2 mg/7 mL PBS) was added in the solution. Samples were 427 centrifuged for 5 min at 6500 rpm. The precipitate removed from the

solution was repeatedly washed with deionized water and PBS before 428 being dried for 3 h at 50 °C.

4.6. Diffraction Experiment (XRD). Powder X-ray diffraction 430 patterns were acquired in Bragg-Brentano geometry with Cu Ka 431 radiation (analytical X'Pert Pro powder diffractometer). 432

4.7. Transmission Electron Microscopy. Transmission electron 433 microscopy (TEM) observations have been performed with a JEOL 434 JEM1220. TEM samples were prepared by dispersing a few milligrams 435 of the compounds in 2 mL of distilled water and dropping 3 μ L of 436 solution on carbon-coated copper grids. 437

4.8. Attenuated Total Reflection Fourier-Transform Infrared 438 Spectroscopy. Attenuated total reflection Fourier-transform infrared 439 spectroscopy spectra of dried samples were obtained on a Thermo 440 Scientific Nicolet iS20 FTIR spectrometer. 441

4.9. Optical Studies. Absorption spectra were recorded using a 442 Cary Lambda 900 spectrophotometer at normal incidence with 443 Suprasil quartz cuvettes with a 0.1 cm optical path length. Steady-state 444 PL and PL excitation spectra have been recorded using a xenon lamp 445 as an excitation source, together with a double monochromator 446 (Jobin-Yvon Gemini 180 with a 1200 grooves/mm grating), and 447 recorded through a nitrogen-cooled charge-coupled device (CCD) 448 detector coupled to a monochromator (Jobin-Yvon Micro HR). 449 Under cw laser excitation, signals have been recorded using a 450 nitrogen-cooled CCD coupled with a double monochromator, Triax- 451 190 (HORIBA Jobin-Yvon), with a spectral resolution of 0.5 nm. All 452 spectra have been corrected for the setup optical response. Time- 453 resolved PL spectra have been recorded using a pulsed light-emitting 454 diode (LED) at 250 nm (3.65 eV, EP-LED 340 Edinburgh 455 Instruments, a pulse width of 700 ps) or a pulsed laser at 405 nm 456 (3.06 eV, EPL-405 Edinburgh Instruments, a pulse width of 150 ps) 457 as a light source. Data were obtained with an Edinburgh Instruments 458 FLS-980 spectrophotometer, with a 5 nm bandwidth and a time 459 resolution of 0.1 ns. 460

4.10. Radioluminescence Experiments. RL measurements 461 were performed by irradiating the samples at room temperature 462 with Philips 2274 (steady-state RL spectroscopy) or a Machlett OEG 463 50 (SO production monitoring experiment) X-ray tubes, both with a 464 tungsten target, equipped with a beryllium window and operated at 20 465 kV and 20 mA. At this voltage, X-rays are generated by the 466 bremsstrahlung mechanism superimposed onto the L and M 467 transition lines of tungsten due to the impact of electrons generated 468 through the thermionic effect and accelerated onto a tungsten target. 469 No beam filtering has been applied. RL spectra have been recorded 470 using a homemade apparatus featuring a liquid nitrogen-cooled CCD 471 (Jobin-Yvon Symphony II) coupled to a monochromator (Jobin-Yvon 472 Triax 180) with a 100 grooves/mm grating as the detection system. 473 The spectra were corrected for the setup optical response. For RL 474 experiments, the NT-x powder was used to fill small aluminum 475 crucibles of 1 mm thickness to completely absorb the incident X-rays. 476 Therefore, in all samples, we have the same amount of deposited 477 energy. Therefore, $\overline{\phi}_{\rm scint}^{\rm PS}$ is directly given by the ratio of the integrated $_{478}$ intensity of the RL spectra. 479

4.11. Scintillation Experiments. Scintillation decays under 480 pulsing X-ray excitation were measured at room temperature using 481 picosecond (ps) X-ray tube N5084 (Hamamatsu Photonics, Japan) at 482 40 kV. The X-ray tube was driven by the ps light pulse from a laser 483 with a repetition rate of up to 1 MHz. The signal was detected by a 484 hybrid ps photon detector and Fluorohub unit (Horiba Scientific, 485 Japan). The setup instrumental response function full width at half- 486 maximum was about 70 ps. The scintillation decay curves were 487 detected using a high-pass filter for the range above 580 nm. The 488 emission was monitored from the same sample's surface where it was 489 excited. 490

4.12. SO Relative Concentration Measurement. The optical 491 probe SOSG has been purchased from Thermo Fisher and used as is. 492 The SOSG powder has been diluted in a 1:10 solution of dimethyl 493 sulfoxide and PBS, which has been used to disperse the NTs with a 494 concentration of 4 mg/mL. The intensity of the SOSG fluorescence, 495 which is directly proportional to the concentration of SO in the 496

ACS Applied Materials & Interfaces

497 environment, has been monitored during the X-ray exposure under 498 continuous-wavelength laser light excitation at 473 nm. The 499 integrated SOSG PL is then proportional to the amount of SO 500 produced upon irradiation. The SOSG emission intensity was 501 integrated between 500 and 530 nm, in order to avoid inclusion of 502 the emission of the PSs. The measured values have been corrected by 503 the dye quantum yield, by the relative intrinsic efficiency of SO 504 generation of the two rhodamines (Figure S2), and by the average 505 number of dyes per NT..

506 ASSOCIATED CONTENT

507 Supporting Information

508 The Supporting Information is available free of charge at so9 https://pubs.acs.org/doi/10.1021/acsami.3c00853.

Optical, scintillation, and SO sensitization experiments 510 (PDF) 511

512 **AUTHOR INFORMATION**

513 Corresponding Author

- Angelo Monguzzi Dipartimento di Scienza Dei Materiali, 514
- Università; Degli Studi Milano-Bicocca, 20125 Milano, Italy; 515
- NANOMIB, Center for Biomedical Nanomedicine, University 516
- of Milano-Bicocca, 20126 Milan, Italy; orcid.org/0000-517
- 0001-9768-4573; Email: angelo.monguzzi@unimib.it 518

519 Authors

- Valeria Secchi Dipartimento di Scienza Dei Materiali, 520
- Università; Degli Studi Milano-Bicocca, 20125 Milano, Italy; 521
- NANOMIB, Center for Biomedical Nanomedicine, University 522
- of Milano-Bicocca, 20126 Milan, Italy 523

Francesca Cova – Dipartimento di Scienza Dei Materiali, 524 Università; Degli Studi Milano-Bicocca, 20125 Milano, 525

- Italy; orcid.org/0000-0001-7367-109X 526
- Irene Villa FZU—Institute of Physics of the Czech Academy 527
- of Sciences, 16 200 Prague, Czech Republic; Dipartimento di 528
- Scienza Dei Materiali, Università; Degli Studi Milano-52.9 Bicocca, 20125 Milano, Italy; O orcid.org/0000-0002-530 6150-7847
- 531
- Vladimir Babin FZU—Institute of Physics of the Czech 532 Academy of Sciences, 16 200 Prague, Czech Republic; 533
- orcid.org/0000-0003-3072-2242 534
- Martin Nikl FZU-Institute of Physics of the Czech 535
- Academy of Sciences, 16 200 Prague, Czech Republic; 536 orcid.org/0000-0002-2378-208X 537
- Marcello Campione Department of Earth and 538
- Environmental Sciences, Università; Degli Studi Milano-539
- Bicocca, 20126 Milano, Italy; NANOMIB, Center for 540
- Biomedical Nanomedicine, University of Milano-Bicocca, 541
- 20126 Milan, Italy; orcid.org/0000-0001-5627-6186 542

543 Complete contact information is available at:

s44 https://pubs.acs.org/10.1021/acsami.3c00853

545 Notes

546 The authors declare no competing financial interest.

547 **ACKNOWLEDGMENTS**

548 This work has been supported by the Italian Ministero degli 549 Affari Esteri e della Cooperazione Internazionale (MAECI) 550 Project X-PATH 2020-H45H19000070001, by the Ministero 551 della Salute (project code RF-2016-02362263, NanoTrack-552 EXO), by the Marie Skłodowska-Curie Actions Widening 553 Fellowships (MSCA-WF) grant no. 101003405—HANSOME, 554 and by the Operational Programme Research, Development

and Education financed by European Structural and Invest- 555 ment Funds and the Czech Ministry of Education, Youth and 556 Sports-Project no. SOLID21 CZ.02.1.01/0.0/0.0/16 019/ 557 0000760. 558

REFERENCES

(1) Haque, S.; Whittaker, M. R.; McIntosh, M. P.; Pouton, C. W.; 560 Kaminskas, L. M. Disposition and safety of inhaled biodegradable 561 nanomedicines: Opportunities and challenges. Nanomedicine 2016, 562 12, 1703-1724. 563

(2) Teleanu, D. M.; Chircov, C.; Grumezescu, A. M.; Teleanu, R. I. 564 Neuronanomedicine: An up-to-date Overview. Pharmaceutics 2019, 565 11, 101. 566

(3) Liu, Y.; Zhao, G.; Xu, C.-F.; Luo, Y.-L.; Lu, Z.-D.; Wang, J. 567 Systemic Delivery of CRISPR/Cas9 with PEG-PLGA Nanoparticles 568 for Chronic Myeloid Leukemia Targeted Therapy. Biomater. Sci. 569 2018, 6, 1592-1603. 570

(4) Dong, H.; Du, S.-R.; Zheng, X.-Y.; Lyu, G.-M.; Sun, L.-D.; Li, L.- 571 D.; Zhang, P.-Z.; Zhang, C.; Yan, C.-H. Lanthanide Nanoparticles: 572 from Design toward Bioimaging and Therapy. Chem. Rev. 2015, 115, 573 10725-10815. 574

(5) Zarschler, K.; Rocks, L.; Licciardello, N.; Boselli, L.; Polo, E.; 575 Garcia, K. P.; De Cola, L.; Stephan, H.; Dawson, K. A. Ultrasmall 576 inorganic nanoparticles: State-of-the-art and perspectives for bio- 577 medical applications. Nanomedicine 2016, 12, 1663-1701. 578

(6) Li, M.; Luo, Z.; Zhao, Y. J. C. o. M. Self-assembled Hybrid 579 Nanostructures: versatile Multifunctional Nanoplatforms for Cancer 580 Diagnosis and Therapy. Chem. Mater. 2018, 30, 25-53.

(7) Lu, K.; He, C.; Guo, N.; Chan, C.; Ni, K.; Lan, G.; Tang, H.; 582 Pelizzari, C.; Fu, Y.-X.; Spiotto, M. T.; et al. Low-dose X-ray 583 Radiotherapy-radiodynamic Therapy via Nanoscale Metal-organic 584 Frameworks enhances Checkpoint Blockade Immunotherapy. Nat. 585 Biomed. Eng. 2018, 2, 600-610. 586

(8) Alqathami, M.; Blencowe, A.; Yeo, U. J.; Doran, S. J.; Qiao, G.; 587 Geso, M. Novel Multicompartment 3-dimensional Radiochromic 588 Radiation Dosimeters for Nanoparticle-enhanced Radiation Therapy 589 Dosimetry. Int. J. Radiat. Oncol. 2012, 84, e549-e555. 590

(9) Shaffer, T. M.; Drain, C. M.; Grimm, J. J. Optical Imaging of 591 Ionizing Radiation from Clinical Sources. J. Nucl. Med. 2016, 57, 592 1661-1666. 593

(10) Chen, X.; Song, J.; Chen, X.; Yang, H. X-ray-activated 594 Nanosystems for Theranostic Applications. Chem. Soc. Rev. 2019, 48, 595 3073-3101. 596

(11) Crapanzano, R.; Secchi, V.; Villa, I. Co-adjuvant Nanoparticles 597 for Radiotherapy Treatments of Oncological Diseases. Appl. Sci. 2021, 598 11, 7073. 599

(12) Sun, W.; Zhou, Z.; Pratx, G.; Chen, X.; Chen, H. 600 Nanoscintillator-mediated X-ray induced Photodynamic Therapy for 601 Deep-seated Tumors: from Concept to Biomedical Applications. 602 Theranostics 2020, 10, 1296-1318. 603

(13) Bulin, A.-L.; Broekgaarden, M.; Simeone, D.; Hasan, T. Low 604 dose Photodynamic Therapy harmonizes with Radiation Therapy to 605 induce Beneficial Effects on Pancreatic Heterocellular Spheroids. 606 Oncotarget 2019, 10, 2625-2643.

(14) Esposito, M.; Villaggi, E.; Bresciani, S.; Cilla, S.; Falco, M. D.; 608 Garibaldi, C.; Russo, S.; Talamonti, C.; Stasi, M.; Mancosu, P. 609 Estimating Dose Delivery Accuracy in Stereotactic Body Radiation 610 Therapy: a Review of in-vivo Measurement Methods. Radiother. 611 Oncol. 2020, 149, 158-167. 612

(15) Vozenin, M.-C.; Bourhis, J.; Durante, M. Towards Clinical 613 Translation of FLASH Radiotherapy. Nat. Rev. Clin. Oncol. 2022, 19, 614 791-803. 615

(16) Dolmans, D. E. J. G. J.; Fukumura, D.; Jain, R. K. 616 Photodynamic Therapy for Cancer. Nat. Rev. Cancer 2003, 3, 380- 617 387. 618

(17) DeRosa, M. C.; Crutchley, R. J. Photosensitized Singlet Oxygen 619 and its Applications. Coord. Chem. Rev. 2002, 233-234, 351-371. 620

559

(18) Agostinis, P.; Berg, K.; Cengel, K. A.; Foster, T. H.; Girotti, A.
W.; Gollnick, S. O.; Hahn, S. M.; Hamblin, M. R.; Juzeniene, A.;
Kessel, D.; Korbelik, M.; Moan, J.; Mroz, P.; Nowis, D.; Piette, J.;
Wilson, B. C.; Golab, J. Photodynamic Therapy of Cancer: An update. *Ca-Cancer J. Clin.* 2011, *61*, 250–281.

626 (19) Chen, W.; Zhang, J. Using Nanoparticles to Enable 627 Simultaneous Radiation and Photodynamic Therapies for Cancer 628 Treatment. J. Nanosci. Nanotechnol. **2006**, *6*, 1159–1166.

629 (20) Villa, I.; Villa, C.; Crapanzano, R.; Secchi, V.; Tawfilas, M.; 630 Trombetta, E.; Porretti, L.; Brambilla, A.; Campione, M.; Torrente, 631 Y.; Vedda, A.; Monguzzi, A. Functionalized Scintillating Nanotubes 632 for Simultaneous Radio- and Photodynamic Therapy of Cancer. ACS 633 Appl. Mater. Interfaces **2021**, *13*, 12997–13008.

634 (21) Lu, L.; Sun, M.; Wu, T.; Lu, Q.; Chen, B.; Huang, B. All-635 inorganic Perovskite Nanocrystals: Next-generation Scintillation 636 Materials for High-resolution X-ray Imaging. *Nanoscale Adv.* **2022**, 637 *4*, 680–696.

638 (22) Secchi, V.; Monguzzi, A.; Villa, I. Design Principles of Hybrid 639 Nanomaterials for Radiotherapy Enhanced by Photodynamic 640 Therapy. *Int. J. Mol. Sci.* **2022**, *23*, 8736.

641 (23) Chen, H.; Wang, G. D.; Chuang, Y.-J.; Zhen, Z.; Chen, X.; 642 Biddinger, P.; Hao, Z.; Liu, F.; Shen, B.; Pan, Z.; Xie, J. 643 Nanoscintillator-Mediated X-ray Inducible Photodynamic Therapy 644 for In Vivo Cancer Treatment. *Nano Lett.* **2015**, *15*, 2249–2256.

645 (24) Lan, G.; Ni, K.; Xu, R.; Lu, K.; Lin, Z.; Chan, C.; Lin, W. 646 Nanoscale Metal–Organic Layers for Deeply Penetrating X-ray-647 Induced Photodynamic Therapy. *Angew. Chem., Int. Ed.* **2017**, *56*, 648 12102–12106.

649 (25) Fan, W.; Tang, W.; Lau, J.; Shen, Z.; Xie, J.; Shi, J.; Chen, X. 650 Breaking the Depth Dependence by Nanotechnology-Enhanced X-651 Ray-Excited Deep Cancer Theranostics. *Adv. Mater.* **2019**, *31*, 652 1806381.

(26) Ren, X.-D.; Hao, X.-Y.; Li, H.-C.; Ke, M.-R.; Zheng, B.-Y.;
Huang, J.-D. Progress in the Development of Nanosensitizers for Xray-induced Photodynamic Therapy. *Drug Discovery Today* 2018, 23,
1791–1800.

(27) Shrestha, S.; Wu, J.; Sah, B.; Vanasse, A.; Cooper, L. N.; Ma, L.;
658 Li, G.; Zheng, H.; Chen, W.; Antosh, M. P. X-ray induced
659 Photodynamic Therapy with Copper-cysteamine Nanoparticles in
660 Mice Tumors. *Proc. Natl. Acad. Sci. U. S. A.* 2019, *116*, 16823–16828.
661 (28) He, L.; Yu, X.; Li, W. Recent Progress and Trends in X-ray662 Induced Photodynamic Therapy with Low Radiation Doses. *ACS*663 *Nano* 2022, *16*, 19691–19721.

664 (29) Ahmad, F.; Wang, X.; Jiang, Z.; Yu, X.; Liu, X.; Mao, R.; Chen,
665 X.; Li, W. Codoping Enhanced Radioluminescence of Nano666 scintillators for X-ray-Activated Synergistic Cancer Therapy and
667 Prognosis Using Metabolomics. ACS Nano 2019, 13, 10419–10433.
668 (30) Bulin, A.-L.; Broekgaarden, M.; Chaput, F.; Baisamy, V.;
669 Garrevoet, J.; Busser, B.; Brueckner, D.; Youssef, A.; Ravanat, J.-L.;
670 Dujardin, C.; Motto-Ros, V.; Lerouge, F.; Bohic, S.; Sancey, L.;
671 Elleaume, H. Radiation Dose-Enhancement Is a Potent Radio672 therapeutic Effect of Rare-Earth Composite Nanoscintillators in
673 Preclinical Models of Glioblastoma. Adv. Sci. 2020, 7, 2001675.

674 (31) Liu, Y.; Chen, W.; Wang, S.; Joly, A. G. Investigation of Water-675 soluble X-ray Luminescence Nanoparticles for Photodynamic 676 Activation. *Appl. Phys. Lett.* **2008**, *92*, 043901.

(32) Villa, C.; Campione, M.; Santiago-González, B.; Alessandrini,
F.; Erratico, S.; Zucca, I.; Bruzzone, M. G.; Forzenigo, L.; Malatesta,
P.; Mauri, M.; Trombetta, E.; Brovelli, S.; Torrente, Y.; Meinardi, F.;
Monguzzi, A. Self-Assembled pH-Sensitive Fluoromagnetic Nanotubes as Archetype System for Multimodal Imaging of Brain Cancer.
Adv. Funct. Mater. 2018, 28, 1707582.

683 (33) Pope, M.; Swenberg, C. E. *Electronic Processes in Organic* 684 *Crystals and Polymers*; Oxford University Press, 1999; Vol. 39.

(34) Nair, D. P.; Podgóroski, M.; Chatani, S.; Gong, T.; Xi, W.;
Fenolis, C. R.; Bowman, C. N. The Thiol-Michael Addition Click
Reaction: A Powerful and Widely Used Tool in Materials Chemistry. *Chem. Mater.* 2014, 26, 724–744.

(35) Northrop, B. H.; Frayne, S. H.; Choudhary, U. J. P. C. Thiol- 689 maleimide "click" Chemistry: Evaluating the Influence of Solvent, 690 Initiator, and Thiol on the Reaction Mechanism, Kinetics, and 691 Selectivity. *Polymers* **2015**, *6*, 3415–3430. 692

(36) De Luca, G.; Romeo, A.; Villari, V.; Micali, N.; Foltran, I.; 693 Foresti, E.; Lesci, I. G.; Roveri, N.; Zuccheri, T.; Scolaro, L. M. Self- 694 organizing Functional Materials via Ionic Self Assembly: Porphyrins 695 H-and J-aggregates on Synthetic Chrysotile Nanotubes. J. Am. Chem. 696 Soc. **2009**, 131, 6920–6921. 697

(37) Falini, G.; Foresti, E.; Gazzano, M.; Gualtieri, A. F.; Leoni, M.; 698 Lesci, I. G.; Roveri, N. Tubular-shaped Stoichiometric Chrysotile 699 Nanocrystals. *Chem. —Eur. J.* **2004**, *10*, 3043–3049. 700

(38) Parker, S. F. Vibrational spectroscopy of N-methylmaleimide. 701 Chem. —Eur. J. 1995, 51, 2067–2072. 702

(39) Altman, R. S.; Crofton, M. W.; Oka, T. Observation of the 703 Infrared ν 2 band (CH stretch) of Protonated Hydrogen Cyanide 704 HCNH+. J. Chem. Phys. **1984**, 80, 3911–3912. 705

(40) Kim, S.; Fujitsuka, M.; Majima, T. Photochemistry of Singlet 706 Oxygen Sensor Green. J. Phys. Chem. B **2013**, 117, 13985–13992. 707

(41) Bulin, A.-L.; Vasil'ev, A.; Belsky, A.; Amans, D.; Ledoux, G.; 708 Dujardin, C. Modelling Energy Deposition in Nanoscintillators to 709 Predict the Efficiency of the X-ray-induced Photodynamic Effect. 710 *Nanoscale* **2015**, *7*, 5744–5751. 711

(42) Procházková, L.; Pelikánová, I. T.; Mihóková, E.; Dědic, R.; 712 Čuba, V. Novel Scintillating Nanocomposite for X-ray Induced 713 Photodynamic Therapy. *Radiat. Meas.* **2019**, *121*, 13–17. 714

(43) Villa, I.; Moretti, F.; Fasoli, M.; Rossi, A.; Hattendorf, B.; 715 Dujardin, C.; Niederberger, M.; Vedda, A.; Lauria, A. The Bright X- 716 Ray Stimulated Luminescence of HfO₂ Nanocrystals Activated by Ti 717 Ions. *Adv. Opt. Mater.* **2020**, *8*, 1901348. 718

(44) Hill, R.; Healy, B.; Holloway, L.; Kuncic, Z.; Thwaites, D.; 719 Baldock, C. Advances in Kilovoltage X-ray Beam Dosimetry. *Phys.* 720 *Med. Biol.* **2014**, *59*, R183–R231. 721

(45) Hachadorian, R. L.; Bruza, P.; Jermyn, M.; Gladstone, D. J.; 722 Pogue, B. W.; Jarvis, L. A. Imaging Radiation Dose in Breast 723 Radiotherapy by X-ray CT calibration of Cherenkov light. *Nat.* 724 *Commun.* **2020**, *11*, 2298. 725

(46) Lin, J.; Wang, S.; Huang, P.; Wang, Z.; Chen, S.; Niu, G.; Li, 726 W.; He, J.; Cui, D.; Lu, G.; et al. Photosensitizer-loaded Gold Vesicles 727 with Strong Plasmonic Coupling Effect for Imaging-guided Photo-728 thermal/photodynamic Therapy. ACS Nano **2013**, 7, 5320–5329. 729

(47) Wu, M.; Guo, H.; Liu, L.; Liu, Y.; Xie, L. J. >Size-dependent 730 cellular uptake and localization profiles of silver nanoparticles<>. *Int.* 731 *J. Nanomed.* **2019**, *14*, 4247–4259. 732