

Review **From Brain to Muscle: The Role of Muscle Tissue in Neurodegenerative Disorders**

Elisa Duranti and Chiara Villa [*](https://orcid.org/0000-0003-2730-5974)

School of Medicine and Surgery, University of Milano-Bicocca, 20900 Monza, Italy; e.duranti@campus.unimib.it ***** Correspondence: chiara.villa@unimib.it; Tel.: +39-02-6448-8138

Simple Summary: Neurodegenerative diseases (NDs), like amyotrophic lateral sclerosis (ALS), Alzheimer's disease (AD), and Parkinson's disease (PD), mainly impact the central nervous system, resulting in neuronal death and motor/cognitive impairments. However, recent studies have also shown a significant role of muscle tissue in the pathogenesis of these disorders, suggesting that pathological processes may co-exist in both the brain and muscle tissue. Understanding muscle involvement in NDs can provide novel therapeutic targets for developing more effective treatments for patients affected by NDs.

Abstract: Neurodegenerative diseases (NDs), like amyotrophic lateral sclerosis (ALS), Alzheimer's disease (AD), and Parkinson's disease (PD), primarily affect the central nervous system, leading to progressive neuronal loss and motor and cognitive dysfunction. However, recent studies have revealed that muscle tissue also plays a significant role in these diseases. ALS is characterized by severe muscle wasting as a result of motor neuron degeneration, as well as alterations in gene expression, protein aggregation, and oxidative stress. Muscle atrophy and mitochondrial dysfunction are also observed in AD, which may exacerbate cognitive decline due to systemic metabolic dysregulation. PD patients exhibit muscle fiber atrophy, altered muscle composition, and α -synuclein aggregation within muscle cells, contributing to motor symptoms and disease progression. Systemic inflammation and impaired protein degradation pathways are common among these disorders, highlighting muscle tissue as a key player in disease progression. Understanding these muscle-related changes offers potential therapeutic avenues, such as targeting mitochondrial function, reducing inflammation, and promoting muscle regeneration with exercise and pharmacological interventions. This review emphasizes the importance of considering an integrative approach to neurodegenerative disease research, considering both central and peripheral pathological mechanisms, in order to develop more effective treatments and improve patient outcomes.

Keywords: muscle; neurodegenerative diseases; therapy

1. Introduction

Neurodegenerative diseases (NDs) encompass a group of progressive disorders characterized by the deterioration and death of nerve cells, leading to a decline in cognitive and/or motor functions [\[1](#page-15-0)[,2\]](#page-15-1). Major NDs include amyotrophic lateral sclerosis (ALS), which affects motor neurons, leading to muscle weakness and progressive paralysis [\[3,](#page-15-2)[4\]](#page-15-3); Alzheimer's disease (AD), characterized by progressive memory loss and cognitive decline [\[5](#page-15-4)[,6\]](#page-15-5); and Parkinson's disease (PD), known for motor symptoms such as tremors and muscle stiffness due to the loss of dopaminergic neurons [\[7\]](#page-15-6). Despite their highly different pathophysiology and symptomatology, these diseases share several common features. A fundamental characteristic is neuronal degeneration, which manifests as the progressive loss of neurons in specific areas of the brain or spinal cord. This process affects a variety of neurons, such as those controlling movement (in ALS and PD) or those involved in memory

Citation: Duranti, E.; Villa, C. From Brain to Muscle: The Role of Muscle Tissue in Neurodegenerative Disorders. *Biology* **2024**, *13*, 719. [https://doi.org/10.3390/](https://doi.org/10.3390/biology13090719) [biology13090719](https://doi.org/10.3390/biology13090719)

Academic Editor: Hyun Jin Choi

Received: 7 August 2024 Revised: 2 September 2024 Accepted: 11 September 2024 Published: 12 September 2024

Copyright: © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license [\(https://](https://creativecommons.org/licenses/by/4.0/) [creativecommons.org/licenses/by/](https://creativecommons.org/licenses/by/4.0/) $4.0/$).

and cognitive functions (in AD) [\[8](#page-15-7)[,9\]](#page-15-8). Another common hallmark is the presence of abnormal protein aggregates within or around nerve cells. For instance, ALS in characterized by accumulations of TDP-43 and SOD-1 protein, AD by the formation of amyloid-β (Aβ) plaques and tau tangles, and PD by Lewy bodies containing α -synuclein (α -syn). These aggregates can disrupt cellular function and trigger inflammatory responses that further contribute to neuronal degeneration $[3,10-13]$ $[3,10-13]$ $[3,10-13]$. However, there is currently no definitive cure that can completely reverse the course of NDs, and the available therapeutic options are mostly focused on managing symptoms and slowing disease progression [\[14,](#page-15-11)[15\]](#page-15-12).

In NDs, muscle tissue also plays a crucial role, as the degeneration of motor neurons and alterations in neural signals lead to significant muscle function loss. In ALS, AD, and PD, muscles are directly or indirectly affected, contributing to the debilitating symptoms characteristic of these diseases [\[16,](#page-15-13)[17\]](#page-15-14). In ALS, the progressive loss of motor neurons in the brain and spinal cord causes muscle weakness and atrophy, compromising patients' ability to move and perform daily activities [\[4,](#page-15-3)[18\]](#page-15-15). Though primarily associated with cognitive deficits, even in AD, sarcopenia, or the age-related loss of muscle mass, can occur, exacerbating immobility and increasing the risk of falls [\[19,](#page-15-16)[20\]](#page-16-0). Also, in PD, a decrease in dopamine levels leads to muscle rigidity, tremors, and bradykinesia, which restrict mobility and balance [\[21,](#page-16-1)[22\]](#page-16-2). Therefore, the importance of muscle tissue in NDs lies not only in its direct impact on patients' quality of life but also in its potential as a therapeutic target. Interventions aimed at maintaining or improving muscle function, such as physical exercise, physiotherapy, and appropriate diet, can help slow symptom progression and enhance patients' independence [\[23\]](#page-16-3). Moreover, pharmacological treatments that reduce oxidative stress and inflammation in muscles offer new promise for improving muscle conditions and the quality of life in individuals with NDs [\[4,](#page-15-3)[24,](#page-16-4)[25\]](#page-16-5).

This review aims to explore muscle involvement in NDs, such as ALS, AD, and PD. It focuses primarily on muscle atrophy, mitochondrial dysfunction, protein aggregation, and inflammation, examining their involvement in disease progression and symptomatology. Understanding these muscle-related changes will help in the identification of novel pharmacological targets to improve muscle health and patient outcomes.

2. An Overview of Muscle Tissue: Structure and Function

Skeletal muscle is among the most dynamic and adaptable tissues in the human body. In humans, skeletal muscle accounts for around 40% of total body weight, contains 50–75% of the body's proteins, and is responsible for 30–50% of the total body protein turnover. Muscle is composed of 75% water and 20% proteins, with the remaining 5% consisting of inorganic salts, minerals, lipids, and carbohydrates [\[26\]](#page-16-6). Muscle mass is generally regulated by the balance between protein synthesis and degradation, which is governed by factors such as nutritional intake, hormonal levels, physical activity, exercise, and the presence of injury or disease. Muscle protein types, including structural, contractile, and regulatory proteins, are of great scientific interest due to their significant role in mobility, exercise capacity, overall function, and health [\[27](#page-16-7)[–29\]](#page-16-8).

Skeletal muscle is essential for numerous bodily functions. Mechanically, its primary function is to convert chemical energy into mechanical energy, generating force and power while preserving posture and producing movements that allow for activity, participation in social and occupational settings, health maintenance, and functional independence [\[26\]](#page-16-6). Skeletal muscle contributes to basal energy metabolism, stores key substrates such as amino acids and carbohydrates, produces heat to regulate core temperature, and consumes the majority of oxygen and fuel used during physical activity and exercise [\[30,](#page-16-9)[31\]](#page-16-10). Notably, skeletal muscle acts as a storage for amino acids required by other tissues, such as the skin, brain, and heart, for the synthesis of organ-specific proteins. Additionally, the release of amino acids from muscle helps to keep blood glucose levels stable during periods of fasting [\[32\]](#page-16-11). Reduced muscle mass impairs the body's ability to respond to stress and chronic sickness, making it detrimental to disease prevention and health maintenance.

Skeletal muscle is made up of multinucleated cells called myofibers, which are gen-Skeletal muscle is made up of multinucleated cells called myofibers, which are generated during development when myoblasts fuse together [\[33,](#page-16-12)[34\]](#page-16-13). When muscle tissue erated during development when myoblasts fuse together [33,34]. When muscle tissue is is damaged, a complex response is triggered, leading to tissue regeneration [\[35\]](#page-16-14). This damaged, a complex response is triggered, leading to tissue regeneration [35]. This regenregenerative process is primarily driven by satellite cells (SCs), which respond to signals from the environment. SCs replenish myogenic progenitor cells and differentiate into new myofibers to repair muscle damage following injury [\[33,](#page-16-12)[34\]](#page-16-13). Myofibers, the functional components of skeletal muscle, are characterized by a multinucleated structure (Figure [1\)](#page-2-0). The fusion of myoblasts into myotubes, which form these fibers, is regulated by specific transcription factors, such as MyoD, Myf5, myogenin, and MRF4, collectively known as scription factors, such as MyoD, Myf5, myogenin, and MRF4, collectively known as myo-myogenic regulatory factors (MRFs) [\[34](#page-16-13)[,36\]](#page-16-15). These factors coordinate the gene expression necessary for muscle differentiation and myofiber formation [\[37\]](#page-16-16). Skeletal muscle is primarily composed of myofibers, connective tissue, blood vessels, and nerves. Myofibers are organized into muscle fascicles, wrapped by the perimysium, and each individual myofiber is surrounded by the endomysium $[38]$. Myofibrils are cylindrical structures that extend the length of a muscle fiber. Myofibrils are composed of sarcomeres, repetitive units that represent the functional unit of muscle contraction. Sarcomeres are made up of actin and myosin filaments, whose interaction is fundamental to the muscle contraction process [[39\]](#page-16-18). The regeneration of skeletal muscle is mainly mediated by SCs, stem cells located between The regeneration of skeletal muscle is mainly mediated by SCs, stem cells located between the basal membrane and the sarcolemma of myofibers. In response to muscle damage, SCs the basal membrane and the sarcolemma of myofibers. In response to muscle damage, SCs activate and proliferate, generating myoblasts that differentiate and fuse to repair or form activate and proliferate, generating myoblasts that differentiate and fuse to repair or form new myofibers [34]. new myofibers [\[34\]](#page-16-13).

Figure 1. A schematic representation of skeletal muscle structure. The image was created with the Figure 1. A schematic representation of skeletal muscle structure. The image was created with the
use of Servier Medical Art modified templates, licensed under a Creative Common Attribution 3.0 License (https://smart.servier.com, accessed on 28 August 2024). Unported License [\(https://smart.servier.com,](https://smart.servier.com) accessed on 28 August 2024).

3. The Role of Muscle Pathology in ALS: Mechanisms and Treatment Strategies 3. The Role of Muscle Pathology in ALS: Mechanisms and Treatment Strategies

3.1. Overview of Pathogenic Mechanisms in ALS 3.1. Overview of Pathogenic Mechanisms in ALS

ALS is a fatal ND characterized by the progressive degeneration of both upper and ALS is a fatal ND characterized by the progressive degeneration of both upper and lower motor neurons, which contain cytoplasmic inclusions [\[34,](#page-16-13)[40\]](#page-16-19). Upper motor neuron deterioration results in spasticity and hyperexcitability, while the lower motor neuron loss deterioration results in spasticity and hyperexcitability, while the lower motor neuron loss leads to weakness, fasciculations, and, ultimately, muscle atrophy, followed by progressive
diagonal control of the diagonal control of the diagonal control of the diagonal control of the diagonal contr paralysis [\[3,](#page-15-2)[41\]](#page-16-20). Early symptoms include muscle cramping and stiffness, progressing to paralysis [3,41]. Early symptoms include muscle cramping and stiffness, progressing to muscle weakness that affects the arms and legs [\[34,](#page-16-13)[42\]](#page-16-21). Patients often experience slurred
masch and difficulties with sharing an availability [42]. Ultimately death a gwn as a may ble speech and difficulties with chewing or swallowing [43]. Ultimately, death occurs as a of respiratory failure and pneumonia complications within approximately 3–5 years after or respiratory failure and predimerial complications within approximately 3–5 years after symptom onset [\[3\]](#page-15-2). While most ALS cases (~90–95%) are sporadic (sALS) with no known etiology, about 5–10% of cases involve familial gene mutations following a Mendelian etiology, about 5–10% of cases involve familial gene mutations following a Mendelian speech and difficulties with chewing or swallowing [\[43\]](#page-16-22). Ultimately, death occurs as a result inheritance pattern, known as familial ALS (fALS) [\[3](#page-15-2)[,44\]](#page-16-23).

The pathogenic mechanisms underlying ALS are complex, involving a combination of protein dysfunction, oxidative stress, mitochondrial dysfunction, excitotoxicity, neuroinflammation, alterations in axonal transport, and genetic factors. One of the main pathogenic processes of ALS is the formation of cytoplasmic inclusions within motor neurons. These inclusions are often composed of misfolded proteins, such as superoxide dismutase 1 (SOD1) and transactive response DNA-binding protein 43 kDa (TDP-43). The dysfunction of these proteins can cause endoplasmic reticulum (ER) stress and the activation of the unfolded protein response, ultimately leading to cell death [\[3,](#page-15-2)[45\]](#page-16-24). Oxidative stress is another significant factor in the pathophysiology of ALS. Motor neurons are particularly vulnerable to oxidative damage due to their high metabolic activity, which results in the production of reactive oxygen species (ROS). The mutation of SOD1, an antioxidant enzyme, impairs the cells' ability to neutralize ROS, leading to cellular damage and neuronal death [\[46,](#page-16-25)[47\]](#page-16-26). On the other hand, mutant SOD1 can cause a global variation in structure, which may result in a gain of toxicity, exacerbating the disease [\[48\]](#page-16-27). Mitochondrial dysfunction plays a critical role in ALS pathogenesis. The mitochondria of affected motor neurons exhibit structural and functional abnormalities, reducing ATP generation and increasing ROS production. This energetic dysfunction promotes neuronal degeneration and disease progression [\[49](#page-16-28)[,50\]](#page-16-29). Excitotoxicity, primarily mediated by glutamate, is another pathogenic mechanism of ALS. Affected motor neurons show a diminished capacity to remove synaptic glutamate due to the decreased expression of glutamate transporters such as EAAT2. The accumulation of extracellular glutamate causes excessive activation of N-methyl-Daspartate (NMDA) receptors, leading to increased calcium influx into cells and triggering apoptotic pathways [\[51](#page-16-30)[,52\]](#page-17-0). Neuroinflammation is widely recognized as a significant factor in ALS. Activated microglia and astrocytes release pro-inflammatory cytokines, such as TNF- α and IL-1 β , which contribute to neuronal degeneration. Additionally, the infiltration of peripheral immune cells into the central nervous system (CNS) may exacerbate the inflammatory response, accelerating disease progression. Furthermore, accumulated protein aggregates in the brain also trigger a pro-inflammatory response through microglial activation, contributing to a neurotoxic environment. This chronic inflammatory process not only exacerbates neurodegeneration but also can further promote the accumulation of misfolded proteins, creating a vicious cycle that accelerates neuronal damage [\[53\]](#page-17-1). Alterations in axonal transport are also implicated in ALS pathogenesis [\[54\]](#page-17-2). Motor neurons rely significantly on axonal transport to distribute organelles, proteins, and other essential molecules. Mutations in genes involved in axonal transport, such as *DCTN1* and *TUBA4A,* can impair this process, leading to axonal degeneration and motor neuron death [\[55\]](#page-17-3).

3.2. Involvement of Muscle Tissue in ALS Pathology

Muscle atrophy in ALS is caused primarily by the loss of neural input from degenerating motor neurons. This process begins with the degeneration of lower motor neurons, which affects neuromuscular junctions (NMJs), highly specialized synapses that ensure efficient communication between motor neurons and muscle fibers. Without these impulses, muscle fibers lose their ability to contract effectively, leading to a reduction in muscle mass and strength [\[56\]](#page-17-4). This process is characterized by a significant decrease in muscle fiber size, known as atrophy, and eventually results in muscle fiber loss [\[4,](#page-15-3)[57\]](#page-17-5).

Multiple cellular mechanisms contribute to muscle atrophy in ALS. One of the key pathways involved is the ubiquitin–proteasome system (UPS), which is responsible for the degradation of damaged or misfolded proteins [\[4\]](#page-15-3). Muscle-specific E3 ubiquitin ligases, such as MuRF1 and atrogin-1/MAFbx, are upregulated in ALS, targeting muscle proteins for degradation [\[58\]](#page-17-6). This increased proteolytic activity accelerates muscle wasting [\[56](#page-17-4)[,59\]](#page-17-7). The UPS is a tightly regulated system that labels defective proteins with ubiquitin molecules, marking them for destruction by the proteasome, a large protease complex. In ALS, the dysregulation of this system leads to excessive protein degradation, contributing to the rapid loss of muscle mass [\[4\]](#page-15-3). Additionally, the autophagy–lysosome pathway (ALP), another crucial protein degradation system, is dysregulated in ALS. Autophagy,

the process by which cells degrade and recycle their components, becomes overactive in ALS-affected muscles [\[60\]](#page-17-8). This results in the excessive breakdown of muscle proteins and organelles, further contributing to muscle atrophy [\[61](#page-17-9)[,62\]](#page-17-10). The activation of autophagy is a complex phenomenon, and its role as a cause or effect of the disease remains a topic of ongoing debate [\[63\]](#page-17-11). This activation could be a protective response aimed at clearing toxic protein aggregates and damaged cellular components that accumulate due to ALSrelated effects. However, some evidence suggests that dysregulated or excessive autophagy may contribute to neuronal damage, potentially exacerbating the disease [\[64](#page-17-12)[,65\]](#page-17-13). Given these dual perspectives, it is crucial to further investigate whether autophagy in ALS primarily serves as a compensatory mechanism or whether it plays a more direct role in disease progression. Apoptosis, or programmed cell death, is also a significant factor in ALS muscle pathology [\[66](#page-17-14)[,67\]](#page-17-15). The balance between pro-apoptotic and anti-apoptotic factors is disrupted in ALS, leading to increased muscle cell death. Elevated amounts of pro-apoptotic proteins such as Bax, along with decreased levels of anti-apoptotic proteins like Bcl-2, promote muscle fiber apoptosis [\[56](#page-17-4)[,68\]](#page-17-16). This apoptotic cascade is triggered by a variety of stress signals, including mitochondrial dysfunction and oxidative stress, leading to the systematic dismantling and removal of muscle cells [\[56](#page-17-4)[,69\]](#page-17-17).

The process of denervation and reinnervation is a dynamic feature of ALS muscle pathology. Early in the disease, surviving motor neurons attempt to compensate for lost connections by sprouting new axons that reinnervate denervated muscle fibers. This compensatory reinnervation can temporarily maintain muscle function [\[18,](#page-15-15)[70\]](#page-17-18). However, as the disease advances, the potential for reinnervation diminishes, resulting in extensive and irreversible muscle atrophy [\[70\]](#page-17-18). The initial phase of reinnervation involves the reorganization of the motor unit, but the relentless progression of motor neuron death eventually overwhelms the compensatory mechanisms, leading to the progressive loss of muscle function [\[57\]](#page-17-5). A notable aspect of ALS muscle pathology is the shift in muscle fiber types. ALS predominantly affects type II (fast-twitch) muscle fibers, which are more prone to denervation $[67,71,72]$ $[67,71,72]$ $[67,71,72]$. This initially preserves type I (slow-twitch) fibers, but they progressively undergo atrophy [\[71,](#page-17-19)[72\]](#page-17-20). The shift alters the functional properties of muscles, reducing their overall strength and endurance. Fast-twitch fibers are critical for rapid, powerful movements, whereas slow-twitch fibers are essential for endurance and sustained activities. The selective vulnerability of fast-twitch fibers in ALS disrupts the balance and coordination of muscle activity, contributing to the characteristic weakness and fatigue observed in patients [\[73](#page-17-21)[,74\]](#page-17-22). While the precise mechanisms of this selective vulnerability are not fully understood, several hypotheses have been proposed. One key factor is oxidative stress. Fast-twitch muscle fibers, which rely more heavily on anaerobic glycolysis for energy, are more prone to the accumulation of ROS. This oxidative stress can cause greater damage to these fibers compared to slow-twitch fibers, which are better equipped to handle oxidative stress due to their reliance on oxidative phosphorylation. Further support for this is provided by several studies showing that oxidative damage is particularly detrimental to fast-twitch fibers in ALS [\[56](#page-17-4)[,75](#page-17-23)[,76\]](#page-17-24). Another contributing factor is the susceptibility of fast-twitch fibers to apoptosis. These fibers seem to be more sensitive to apoptotic signals, making them more vulnerable to degeneration in ALS [\[72\]](#page-17-20). Additionally, the interaction between motor neurons and muscle fibers plays a role: fast-twitch fibers are often innervated by motor neurons that are more susceptible to degeneration in ALS. As these motor neurons deteriorate, the fast-twitch fibers they control are more likely to be affected [\[77\]](#page-17-25).

Mitochondrial dysfunction is another important element of muscle involvement in ALS [\[4\]](#page-15-3). Mitochondria in ALS-affected muscles exhibit structural abnormalities, such as swollen and fragmented cristae, which impair their function. These mitochondrial defects reduce ATP synthesis while increasing ROS production [\[78\]](#page-17-26). The resulting bioenergetic deficits and oxidative stress further damage muscle cells and contribute to the progression of muscle atrophy [\[79\]](#page-18-0) (Figure [2\)](#page-5-0). Mitochondria are the powerhouses of the cell, and their dysfunction in ALS leads to an energy crisis within muscle cells, compromising their viability and function. Moreover, the accumulation of ROS causes oxidative damage to proteins, lipids, and DNA, further exacerbating muscle cell death $[4,56]$ $[4,56]$. Inflammation and oxidative stress are significant contributors to ALS muscle damage $[80,81]$ $[80,81]$. Pro-inflammatory cytokines such as TNF-α, IL-1β, and IL-6 are elevated in ALS muscle tissue. These cytokines activate signaling pathways that exacerbate muscle atrophy and fibrosis [\[80\]](#page-18-1). Additionally, oxidative stress markers, including lipid peroxidation and protein carbonylation, are increased in ALS muscles, indicating greater oxidative damage. Chronic inflammation and oxidative stress generate a hostile environment within the muscle tissue, promoting catabolic processes while inhibiting regenerative efforts [\[56,](#page-17-4)[82\]](#page-18-3).

Muscle atrophy in ALS patients

function and cellular damage. This cascade of events results in muscle atrophy and weakness. The image was created with the use of Servier Medical Art modified templates, licensed under a Creative Common Attribution 2.0 Unnerted License (bttps://great.com/gregory.com/gregory.com/gregory.com/gregory.com/gregory.com/gregory.com/gregory.com/gregory.com/gregory.com/gregory.com/gregory.com/gregory.com/gregory.com/gregory mon Attribution 3.0 Unported License (https://smart.servier.com, accessed on 28 August 2024). Common Attribution 3.0 Unported License [\(https://smart.servier.com,](https://smart.servier.com) accessed on 28 August 2024). **Figure 2.** A schematic representation of the accumulation of ROS leading to mitochondrial dys-

and contributing to muscle weakness [\[83\]](#page-18-4). Structural changes in the NMJ include the fragmentation of postsynaptic acetylcholine receptor clusters and a reduction in presynaptic vesicles [\[4\]](#page-15-3). These alterations hinder the effective transmission of nerve signals to muscles, exacerbating muscle atrophy and weakness [\[84\]](#page-18-5). In ALS, the disruption of NMJ architecture affects neuromuscular transmission, further contributing to muscle function loss [\[85](#page-18-6)[,86\]](#page-18-7). In ALS, the integrity of the NMJ is compromised, limiting synaptic transmission

Muscle regeneration in ALS is severely impaired due to malfunctioning SCs, the resident stem cells responsible for muscle maintenance and repair [\[57](#page-17-5)[,87\]](#page-18-8). In ALS, their number is reduced, and their ability to proliferate and differentiate is compromised [\[4](#page-15-3)[,67\]](#page-17-15). This is partly due to the decreased expression of MRFs, such as MyoD and myogenin, which are essential for muscle regeneration [\[36\]](#page-16-15). As a result, the muscle's ability to repair itself in response to damage is considerably reduced [4[,57\]](#page-17-5).

3.3. Therapeutic Strategies for Muscle Tissue in ALS

3.3.1. Pharmacological Treatments

Therapeutic options for ALS aim to delay disease progression, manage symptoms, and improve the quality of life for patients $[42]$. In the past decade, only four drugs have been approved by the FDA for ALS treatment, with differing regulatory approvals worldwide: riluzole, edaravone, the combination therapy $AMX0035$, and tofersen $[4,88]$ $[4,88]$.

Since 1995, the only medication approved for ALS treatment is riluzole, which inhibits glutamate release. Riluzole can halt disease progression and extend the median survival time by approximately three to six months, although it might cause side effects such as liver problems and diarrhea [\[89,](#page-18-10)[90\]](#page-18-11). Despite its benefits, riluzole has not been shown to improve motor neuron function, lung function, fasciculations, or muscle strength. Its mechanism of action involves inhibiting voltage-gated sodium channels in the CNS, thereby reducing calcium influx and glutamate-induced toxicity in the motor cortex and spinal cord. Additionally, riluzole may have antioxidant properties, which aid in alleviating oxidative stress generated by various oxidizing agents [\[91,](#page-18-12)[92\]](#page-18-13). Research on this drug in the muscular field is still in its early stages, and it appears that its effects on muscles are indirect and limited.

Radicava (edaravone) was approved by the FDA in 2017 for the treatment of ALS patients, recognizing its potential therapeutic benefits, as demonstrated in several trials. Edaravone acts as a free radical scavenger, reducing lipid peroxides in a manner comparable to antioxidants like vitamin E and ascorbic acid [\[93](#page-18-14)[–95\]](#page-18-15). It targets hydroxyl radicals, peroxynitrites, and other ROS [\[96](#page-18-16)[,97\]](#page-18-17). This drug protects neuronal, glial, endothelial vascular, and muscle cells against oxidative stress [\[4\]](#page-15-3). However, the precise mechanism by which edaravone exerts its effects in ALS patients remains unclear, and its direct benefits for muscle tissue are still being investigated.

In September 2022, the FDA also approved AMX0035, which is believed to mitigate neuronal cell death by reducing ER stress and mitochondrial dysfunction [\[98\]](#page-18-18). Specifically, this drug operates through a dual mechanism to address neurodegeneration, combining two active compounds: sodium phenylbutyrate and taurursodiol. Sodium phenylbutyrate acts as a chemical chaperone, which alleviates ER stress, whereas taurursodiol, a bile acid, supports mitochondrial function and mitigates oxidative stress. By targeting these cellular stress responses, AMX0035 reduces the levels of oxidative stress and inhibits apoptosis in motor neurons, delaying the course of ALS [\[99,](#page-18-19)[100\]](#page-18-20). This multifaceted approach aims to protect motor neurons from damage and preserve their function, offering a significant advancement in the treatment of this debilitating disease [\[97\]](#page-18-17). In a later open-label extension study, patients treated with AMX0035 exhibited a significantly slower decline in physical function and a longer median overall survival compared to those given a placebo during the phase 2 CENTAUR clinical trial. Clinical trials have shown that AMX0035 can reduce the loss of physical function in ALS patients, suggesting that it might help retain muscle mass and strength [\[4,](#page-15-3)[100](#page-18-20)[–102\]](#page-18-21).

Last year, in April, tofersen received FDA approval based on the results of phase 3 VALOR clinical trial results for ALS patients with a mutation in the *SOD1* gene [\[103\]](#page-18-22). Tofersen is an antisense oligonucleotide designed to lower SOD1 concentrations in the cerebrospinal fluid (CSF) and reduce plasmatic levels of neurofilament light chain (NfL), a marker of neurodegeneration [\[4\]](#page-15-3). Although tofersen did not improve clinical endpoints and was associated with adverse effects, a phase 3 open-label extension study (ATLAS) is ongoing to evaluate its clinical benefits in patients with presymptomatic SOD1-ALS. Although its direct therapeutic effects on muscles are still being investigated, tofersen has shown the ability to reduce the levels of NfL, which could have a favorable impact on muscle health [\[4](#page-15-3)[,103–](#page-18-22)[105\]](#page-18-23).

3.3.2. Therapeutic Potential of Exercise for Muscle Preservation in ALS

Regular physical activity has considerable benefits for patients with neuromuscular disease, including ALS, since it may slow muscle degeneration and preserve NMJ integrity [\[106\]](#page-18-24). Exercise activates muscle metabolism, enhances glucose utilization, and supports muscle regeneration. It also improves antioxidant capacity, mitochondrial biogenesis, and neurogenesis [\[107,](#page-19-0)[108\]](#page-19-1). Research explores these benefits and investigates therapeutic applications cautiously, as incorrect application can lead to adverse effects. Moderateintensity training mitigates cell damage from inflammation and retains muscle mitochondrial function with aging, contrasting with high-intensity training effects [\[4](#page-15-3)[,109,](#page-19-2)[110\]](#page-19-3). In ALS models, moderate exercise improves the phenotypes of SOD1-G93A mice, with aquatic activities like swimming extending the lifespan more effectively than running. Studies suggest swimming's impact on motor units may explain this discrepancy [\[111–](#page-19-4)[113\]](#page-19-5). Exercising, particularly swimming, in ALS models has been linked to the dysregulation of the

BDNF/TrkB pathway due to muscle-contraction-induced BDNF over-secretion, which may exacerbate neurodegeneration [\[114,](#page-19-6)[115\]](#page-19-7). Human studies indicate that exercise therapy improves ALS patients' physical conditions and quality of life compared to standard therapy alone, highlighting its therapeutic promise, pending more targeted studies for optimized treatment options [\[116](#page-19-8)[,117\]](#page-19-9).

4. Exploring Muscle Pathology in Alzheimer's Disease: Current Research and Future Directions

4.1. Overview of Pathogenic Mechanisms in AD

AD is an age-related ND and the most common form of dementia among the elderly population globally, accounting for up to 80% of all diagnoses [\[118\]](#page-19-10). AD is clinically defined as irreversible and progressive neurodegeneration characterized by initial memory loss and cognitive impairments that influence speech, behavior, motor system, and visuospatial orientation, ultimately resulting in an autonomy loss that requires full-time medical care [\[119\]](#page-19-11). Brain atrophy, the extracellular deposition of senile plaques made of insoluble Aβ peptide, and the intracellular formation of neurofibrillary tangles (NFTs) formed by hyperphosphorylated twisted filaments of the microtubule-associated protein tau are the major pathological hallmarks of AD [\[120](#page-19-12)[,121\]](#page-19-13). AD pathogenesis involves not only A β and tau pathology but also microglia-mediated inflammation, oxidative stress, mitochondrial dysfunction, and synaptic damage. These pathways may be linked to cognitive decline, indicating a complex etiology [\[122\]](#page-19-14).

The majority of AD cases are sporadic with a late onset (LOAD), and they frequently affect people aged 65 or older. The two primary risk factors for AD are aging and carrying the ε4 allele of the *APOE* gene, which encodes Apolipoprotein E (ApoE) [\[123\]](#page-19-15). On the other hand, the uncommon early-onset forms of AD (EOAD) usually affect people under the age of 65 and have an autosomal dominant inheritance pattern. These forms are caused by mutations in presenilin-1, presenilin-2, and amyloid precursor protein (APP), which are encoded by the genes *PSEN1*, *PSEN2*, and *APP*, respectively [\[124\]](#page-19-16). They all contribute to APP maturation and processing, resulting in an increase in $A\beta$ synthesis or aggregation [\[125\]](#page-19-17).

4.2. The Impact of Muscle Tissue on AD Pathology

The role of muscle tissue in AD is a developing area of research that could provide new insights into the disease mechanisms and potential therapeutic interventions [\[126\]](#page-19-18). While AD research has historically focused on the brain, it is becoming increasingly clear that the disease has systemic implications, including significant effects on muscle tissue [\[127\]](#page-19-19).

Alterations in muscle tissue in patients with AD are multifactorial and can be attributed to various pathophysiological mechanisms [\[127,](#page-19-19)[128\]](#page-19-20). One of the key mechanisms is mitochondrial dysfunction, which has been extensively documented in the brains of AD patients [\[129\]](#page-19-21). This mitochondrial dysfunction is not limited to the brain but also affects skeletal muscles, leading to reduced ATP production and increased oxidative stress. Muscle cells with malfunctioning mitochondria are unable to maintain efficient energy metabolism, resulting in muscle weakness and fatigue. The inability of mitochondria to generate appropriate energy disrupts normal muscle function and contributes to the overall decline in physical capabilities observed in AD patients [\[130](#page-19-22)[–133\]](#page-19-23). Additionally, the accumulation of $A\beta$ has also been observed in skeletal muscles. This toxic peptide can interfere with muscle function through various mechanisms, including the induction of oxidative stress, ER dysfunction, and the activation of pro-apoptotic pathways [\[134,](#page-20-0)[135\]](#page-20-1). The accumulation of Aβ in muscles may directly contribute to the muscle mass loss and muscle dysfunction observed in AD patients. Therapies targeting Aβ may benefit both neurological and muscle systems due to their pathological overlap [\[134,](#page-20-0)[136](#page-20-2)[,137\]](#page-20-3).

Another critical factor is neuroinflammation, which plays a central role in the pathogenesis of AD. Neuroinflammation is characterized by the chronic activation of glial cells and the release of pro-inflammatory cytokines [\[138\]](#page-20-4), which can spread systemically and

negatively impact muscle tissue, inducing a local inflammatory state [\[139\]](#page-20-5). Chronic inflammation in muscles can lead to protein degradation and reduced protein synthesis, further contributing to the sarcopenia associated with AD. The systemic nature of inflammation highlights the importance of addressing inflammatory pathways in the treatment of AD [\[140](#page-20-6)[,141\]](#page-20-7).

Recent studies have also highlighted alterations in insulin signaling in AD patients, which can affect muscle metabolism. Insulin resistance, common in AD patients, can impair glucose uptake in muscles and alter muscle energy metabolism [\[142](#page-20-8)[–144\]](#page-20-9). This can cause lower protein synthesis and increased protein degradation in muscles, exacerbating muscle mass loss. Insulin resistance not only affects brain glucose metabolism but also has a significant impact on peripheral tissues, implying that metabolic interventions could be beneficial [\[145,](#page-20-10)[146\]](#page-20-11).

The loss of muscle mass and function in AD patients not only compromises mobility and independence but also can worsen cognitive decline [\[127](#page-19-19)[,147](#page-20-12)[–149\]](#page-20-13). Reduced physical activity due to muscle weakness can contribute to further cognitive deterioration through various mechanisms, including reduced cerebral blood flow and increased systemic inflammation. This bidirectional relationship between muscle and cognitive function suggests that preserving muscle health could have significant benefits for overall disease management [\[147,](#page-20-12)[148\]](#page-20-14).

4.3. Therapeutic Approaches for Muscle Tissue in AD

4.3.1. Pharmacological Treatments

Pharmacological interventions targeting systemic inflammation may mitigate muscle damage in AD patients. Nonsteroidal anti-inflammatory drugs (NSAIDs) and other antiinflammatory agents can potentially reduce inflammatory cytokine levels and muscle inflammation [\[150](#page-20-15)[,151\]](#page-20-16). Additionally, agents that enhance mitochondrial function, such as coenzyme Q10 and creatine, can help preserve muscle energy metabolism and prevent atrophy [\[152](#page-20-17)[,153\]](#page-20-18).

Neuromuscular electrical stimulation (NMES) is the process of causing muscles to contract by sending electrical impulses to them. This therapy can help maintain muscle mass and strength in AD patients with limited mobility [\[154\]](#page-20-19). NMES has shown promise in improving muscle function and physical performance in older adults, particularly those affected by NDs. By stimulating muscle contraction and enhancing blood flow, NMES can counteract the effects of reduced physical activity in AD patients.

Hormones such as growth hormone and insulin-like growth factor-1 (IGF-1) play crucial roles in muscle growth and repair [\[155\]](#page-20-20). Hormone supplementation has the potential to reverse muscle atrophy in AD patients. However, the safety and efficacy of such treatments require further investigation. Studies on the use of these hormones in other muscle-wasting conditions suggest potential benefits, but their specific effects on AD need to be explored in greater detail [\[155](#page-20-20)[,156\]](#page-20-21). Ongoing research is focused on understanding the molecular mechanisms linking AD pathology to muscle degeneration. Identifying new therapeutic targets within these pathways could lead to innovative treatments [\[157](#page-20-22)[–159\]](#page-20-23). Combining multiple therapeutic approaches, including exercise, nutritional support, pharmacological treatments, and NMES, may offer synergistic benefits. Personalized medicine approaches that tailor interventions to individual patient profiles hold promise for improving outcomes.

4.3.2. Therapeutic Potential of Exercise for Muscle Preservation in AD

Muscle tissue in AD patients undergoes several pathological changes, including muscle fiber atrophy, increased presence of type II fibers, mitochondrial dysfunction, and heightened oxidative stress [\[160,](#page-21-0)[161\]](#page-21-1). The degeneration of the central nervous system (CNS) disrupts neuromuscular signaling, leading to reduced muscle use and subsequent atrophy. Additionally, systemic inflammation and metabolic dysfunction associated with AD exacerbate muscle degeneration.

Regular physical exercise is one of the most effective interventions for improving muscle health in AD patients. Exercise enhances muscle strength, improves mitochondrial function, and reduces oxidative stress [\[162,](#page-21-2)[163\]](#page-21-3). Aerobic exercises, such as walking and cycling, improve cardiovascular health and muscle endurance, while resistance training increases muscle mass and strength [\[164\]](#page-21-4). Exercise also promotes the release of neurotrophic factors like brain-derived neurotrophic factor (BDNF), which supports neuronal health and function [\[165\]](#page-21-5). Tailoring exercise programs to the capabilities and limitations of AD patients is essential for maximizing benefits and minimizing risks [\[166\]](#page-21-6).

In addition to physical exercise, this condition requires a proper nutritional program [\[167\]](#page-21-7). Adequate nutrition is crucial for maintaining and repairing muscle tissue [\[168\]](#page-21-8). Protein supplementation is vital for counteracting muscle atrophy, providing the necessary building blocks for muscle repair and growth [\[169\]](#page-21-9). Antioxidants, such as vitamins E and C and omega-3 fatty acids, help reduce oxidative stress and inflammation, supporting overall muscle health [\[170,](#page-21-10)[171\]](#page-21-11). Ensuring that AD patients receive a balanced and adequate diet can significantly impact their muscle function and quality of life [\[172\]](#page-21-12).

5. Muscle Dysfunctions in PD: Implications for Motor Symptoms and Therapeutic Strategies

5.1. Overview of Pathogenic Mechanisms in PD

PD is a progressive ND primarily characterized by the degeneration of dopaminergic neurons in the substantia nigra pars compacta, a region in the midbrain [\[173,](#page-21-13)[174\]](#page-21-14). This process leads to decreased dopamine levels in the striatum, adversely affecting motor control and resulting in the characteristic symptoms of the disease, such as resting tremor, muscle rigidity, bradykinesia, and postural instability [\[22,](#page-16-2)[175\]](#page-21-15).

The pathogenesis of PD is complex and multifactorial, arising from the interplay of genetic and environmental factors. Approximately 10–15% of PD cases are familial, indicating a strong genetic component [\[176](#page-21-16)[,177\]](#page-21-17). Several genes have been associated with the disease, including *SNCA* (α-syn), *LRRK2* (leucine-rich repeat kinase 2), *PARK2* (parkin), *PARK7* (DJ-1), and *PINK1* (PTEN-induced kinase 1). Mutations in these genes can lead to cellular dysfunctions that contribute to neurodegeneration [\[178,](#page-21-18)[179\]](#page-21-19). A key pathological hallmark of PD is the presence of Lewy bodies, eosinophilic cytoplasmic inclusions primarily composed of aggregated α -syn [\[180](#page-21-20)[,181\]](#page-21-21). Alpha-syn plays a crucial role in the pathogenesis of PD. Under normal conditions, α -syn regulates synaptic function and neuronal plasticity. However, in pathological conditions, the protein can misfold and aggregate, forming toxic oligomers and insoluble fibrils that contribute to synaptic dysfunction and neuronal death [\[182,](#page-21-22)[183\]](#page-21-23).

Mitochondrial dysfunction and oxidative stress are also pivotal in PD pathogenesis [\[184](#page-21-24)[,185\]](#page-21-25). Mitochondrial dysfunction leads to the excessive production of ROS, resulting in oxidative stress, damage to cellular macromolecules, and neuronal death. Mutations in the *PINK1* and *PARK2* genes, which are involved in mitophagy, the selective removal of damaged mitochondria, can impair mitochondrial function, increasing susceptibility to neurodegeneration [\[186–](#page-21-26)[189\]](#page-22-0).

Autophagy, a cellular process essential for degrading and recycling damaged proteins and organelles, is also disrupted in PD [\[190](#page-22-1)[,191\]](#page-22-2). This impairment leads to the accumulation of misfolded proteins and cellular debris. UPS dysfunction, another crucial pathway for protein degradation, has also been implicated in PD pathogenesis [\[192](#page-22-3)[,193\]](#page-22-4). Dysfunctions in these proteostasis pathways can result in the accumulation of α -syn and other toxic proteins.

Neuroinflammation contributes significantly to the progression of PD [\[194](#page-22-5)[,195\]](#page-22-6). When neurons degenerate, microglial cells, the resident macrophages of the CNS, become activated and release pro-inflammatory cytokines, ROS, and other inflammatory molecules. This inflammatory environment can further damage neurons and contribute to disease progression. Chronic microglial activation and systemic inflammation have been correlated with the severity of PD [\[196](#page-22-7)[,197\]](#page-22-8).

Various cellular signaling pathways are altered in PD. The LRRK2 kinase pathway is of particular interest, as mutations in *LRRK2* are among the most common genetic causes of PD [\[198\]](#page-22-9). LRRK2 is involved in various cellular processes, including vesicular trafficking, autophagy, and immune signaling. Mutations in *LRRK2* can lead to its hyperactivation, causing multiple cellular dysfunctions [\[199](#page-22-10)[,200\]](#page-22-11).

5.2. The Role of Muscle Tissue on PD Pathology

The importance of peripheral tissues, particularly muscle tissue, in PD pathophysiology has gained increasing attention. Recent evidence suggests that muscle tissue not only undergoes secondary changes due to neural degeneration but also may actively contribute to disease progression and symptomatology [\[201\]](#page-22-12). Muscle tissue in PD patients reveals various structural and functional alterations [\[202\]](#page-22-13). These changes are primarily attributed to reduced neural input resulting from dopaminergic neuron degeneration and the consequent motor deficits. Histopathological analysis of muscle biopsies from PD patients has revealed several characteristic changes, including muscle fiber atrophy, type II fiber predominance, and the increased presence of mitochondrial abnormalities [\[203\]](#page-22-14). Muscle fiber atrophy, particularly in type II (fast-twitch) fibers, is commonly observed. This atrophy is likely a consequence of denervation and disuse, secondary to the impaired motor function in PD. There is also a shift toward a higher proportion of type I (slow-twitch) fibers [\[204\]](#page-22-15). This alteration in fiber-type composition may be an adaptive response to the chronic nature of PD, favoring endurance over rapid, forceful contractions. Muscle biopsies frequently reveal an increased presence of abnormal mitochondria, including swelling, disrupted cristae, and the accumulation of electron-dense material. Mitochondrial dysfunction plays a critical role in PD pathogenesis, affecting both the CNS and peripheral tissues such as muscle [\[205,](#page-22-16)[206\]](#page-22-17). Key features of mitochondrial dysfunction in PD muscle tissue include reduced respiratory chain activity, oxidative stress, and genetic mutations [\[207\]](#page-22-18). Muscle biopsies from PD patients exhibit decreased activity of mitochondrial respiratory chain complex I, which is consistent with studies of the substantia nigra [\[208](#page-22-19)[,209\]](#page-22-20). This reduction in complex I activity impairs oxidative phosphorylation and ATP synthesis. Mitochondrial dysfunction is also associated with the increased production of ROS, leading to oxidative damage to cellular components, including lipids, proteins, and DNA [\[210](#page-22-21)[,211\]](#page-22-22). Oxidative stress further exacerbates mitochondrial impairment and muscle damage. Additionally, mutations in genes such as *PINK1* and *PARK2*, which are involved in mitochondrial quality control through mitophagy, have been implicated in PD. Defects in these pathways lead to the accumulation of dysfunctional mitochondria in muscle cells, contributing to muscle degeneration [\[212,](#page-22-23)[213\]](#page-22-24).

Recent studies have identified the presence of α -syn aggregates in the muscle tissues of PD patients [\[214\]](#page-22-25). These aggregates can disrupt cellular homeostasis and contribute to muscle pathology through impaired protein degradation, cytotoxicity, and neuronal–muscular crosstalk. Alpha-syn aggregates can inhibit the UPS system and autophagy, leading to the accumulation of damaged proteins and organelles in muscle cells (Figure [3\)](#page-11-0) [\[215,](#page-22-26)[216\]](#page-23-0). In the UPS, α -syn aggregates can physically obstruct the proteasome's active sites, preventing the degradation of ubiquitinated proteins. Additionally, these aggregates can impair the function of the proteasome by interacting with its subunits, resulting in a decrease in proteasomal activity. This leads to the accumulation of undegraded and damaged proteins, which contributes to cellular stress and dysfunction [\[217\]](#page-23-1).

The presence of α -syn aggregates can also induce cytotoxic effects, including the disruption of cellular membranes, interference with synaptic function, and the triggering of apoptosis [\[218\]](#page-23-2). Alpha-syn pathology in muscle tissue may reflect the spread of pathogenic protein species from the CNS to peripheral tissues, highlighting a bidirectional relationship between neuronal and muscular degeneration in PD [\[219](#page-23-3)[,220\]](#page-23-4).

Neuroinflammation is a key feature of PD, and there is growing evidence that peripheral inflammation, including within muscle tissue, may contribute to disease progres-sion [\[221](#page-23-5)[,222\]](#page-23-6). Elevated levels of pro-inflammatory cytokines, such as TNF- α , IL-1β, and

IL-6, have been detected in the sural nerves of PD patients [\[223\]](#page-23-7). These cytokines can exacerbate muscle degeneration and contribute to the overall disease burden. Immune cells, including macrophages and T lymphocytes, have been found to infiltrate PD muscle tissue more frequently [\[223,](#page-23-7)[224\]](#page-23-8). This immune response may be triggered by the release of damage-associated molecular patterns (DAMPs) from degenerating muscle cells [\[225](#page-23-9)[,226\]](#page-23-10). Chronic systemic inflammation, as observed in PD, can have deleterious effects on muscle tissue, promoting catabolic pathways and muscle wasting [\[227\]](#page-23-11).

Figure 3. The aggregates of α-synuclein caused by alterations in protein degradation pathways promote the muscle cell alterations typically found in PD patients. The image was created with the use of Servier Medical Art modified templates, licensed under a Creative Common Attribution 3.0 ported License (https://smart.servier.com, accessed on 28 August 2024). Unported License [\(https://smart.servier.com,](https://smart.servier.com) accessed on 28 August 2024).

5.3. Therapeutic Interventions for Muscle Tissue in PD

5.3.1. Pharmacological Treatments with synaptic function, and the triggering of triggering of the triggering o

Pharmacological interventions targeting mitochondrial dysfunction and oxidative stress are also being investigated. Mitochondrial enhancers such as coenzyme Q10 and creatine have shown potential in improving mitochondrial function [\[228\]](#page-23-12). Coenzyme Q10 is a component of the electron transport chain and is essential for mitochondrial ATP production. While early studies suggested potential benefits, larger clinical trials revealed conflicting results. Creatine, another mitochondrial enhancer, has shown promise in preclinical models but requires further investigation in clinical trials [\[229](#page-23-13)[–231\]](#page-23-14).

Antioxidants such as N-acetylcysteine (NAC) and alpha-lipoic acid (ALA) aim to mitigate oxidative stress in muscle cells. NAC acts as a precursor to glutathione, a major cellular antioxidant, while ALA is a cofactor for mitochondrial enzymes and a potent antioxidant [232]. These compounds have shown potential in reducing oxidative damage and preserving muscle function, but larger clinical trials are needed to confirm their efficacy in PD [233]. \blacksquare

drugs such as NSAIDs and corticosteroids may help mitigate this [\[234\]](#page-23-17). NSAIDs inhibit cyclooxygenase (COX) enzymes, reducing the production of pro-inflammatory prosugalated by the contessectorus, on the other hand, suppress a broad range of in mitimetery paintwys. These agents come readed masche inhancement and degeneratery,
although their long-term use must be carefully managed due to potential side effects. Chronic inflammation contributes to muscle pathology in PD, and anti-inflammatory prostaglandins [\[235\]](#page-23-18). Corticosteroids, on the other hand, suppress a broad range of in-

Adequate protein intake is essential for muscle repair and growth. Protein suppleis a comparison transportein transport chain and is essential for mitochondrial and growth. If you'll receive ments, such as whey protein, provide a convenient way to ensure that PD patients receive α duction. While early studies suggested potential behavior contains the suggested potential trials reduced appear contains revealed α difficulty α dinox. Where sufficient protein, especially those with a reduced appetite or difficulty eating. Whey

protein contains important amino acids, particularly leucine, which is critical for muscle protein synthesis. Branched-chain amino acids (BCAAs), including leucine, isoleucine, and valine, are particularly effective in stimulating muscle protein synthesis [\[236](#page-23-19)[–239\]](#page-23-20). Leucine, in particular, activates the mTOR pathway, which is crucial for muscle growth and repair [\[240\]](#page-23-21). Supplementing with BCAAs may help prevent muscle wasting and improve muscle strength in PD patients [\[241,](#page-23-22)[242\]](#page-23-23).

5.3.2. Therapeutic Potential of Exercise for Muscle Preservation in PD

Exercise has been shown to have neuroprotective effects in PD, potentially mediated through its impact on muscle tissue. Regular physical activity can improve muscle strength, enhance mitochondrial function, and reduce inflammation. Exercise-induced muscle contraction stimulates the production of neurotrophic factors such as BDNF, which can cross the blood–brain barrier and exert protective effects on dopaminergic neurons [\[243\]](#page-24-0). Additionally, exercise promotes autophagy, which aids in the clearance of damaged proteins and organelles in muscle cells, potentially reducing the pathological burden in both muscle and neural tissues.

Aerobic exercise is particularly beneficial for cardiovascular health and muscle endurance in PD patients. It enhances mitochondrial biogenesis, increasing the number and function of mitochondria within muscle cells, which is critical given the mitochondrial dysfunction observed in PD [\[244,](#page-24-1)[245\]](#page-24-2). Aerobic activities, such as brisk walking, cycling, and swimming, also promote neurogenesis and the release of neurotrophic factors like BDNF, which support neuronal health and synaptic plasticity. Studies have demonstrated that regular aerobic exercise can improve motor function, reduce bradykinesia, and enhance the overall quality of life in PD patients [\[246](#page-24-3)[,247\]](#page-24-4).

Resistance training, which involves lifting weights or using resistance bands, can significantly increase muscle mass and strength. This type of exercise stimulates muscle protein synthesis and counteracts the sarcopenia (muscle wasting) commonly observed in PD patients [\[248\]](#page-24-5). Research shows that resistance training can improve muscle strength, enhance balance and coordination, and reduce the risk of falls, which are common in PD due to postural instability [\[249,](#page-24-6)[250\]](#page-24-7).

Flexibility and balance exercises, such as yoga and tai chi, are critical for preserving joint range of motion and enhancing proprioception. These exercises improve flexibility, balance, and coordination, reducing the risk of falls and improving functional mobility. Tai chi, in particular, has been shown to improve balance and reduce the number of falls in PD patients through its slow, controlled movements and emphasis on body awareness [\[251–](#page-24-8)[253\]](#page-24-9).

6. Muscle Degeneration across Neurodegenerative Disorders: Comparative Insights from ALS, AD, and PD

6.1. Similarities in the Effects on Muscle Tissue

6.1.1. Muscle Atrophy and Weakness

In all three NDs, weakness and muscle atrophy are common pathological features [\[254\]](#page-24-10). Muscle atrophy occurs due to a combination of factors, including denervation, reduced motor neuron input, and the systemic effects of chronic disease [\[255\]](#page-24-11). In ALS and PD, the progressive loss of motor neurons directly affects the muscle tissue, leading to atrophy and weakness [\[256\]](#page-24-12). AD patients also exhibit muscle wasting, which may be due to generalized neurodegeneration and the physical inactivity associated with cognitive decline [\[257\]](#page-24-13).

6.1.2. Mitochondrial Dysfunction

Mitochondrial dysfunction characterizes the pathophysiology of all three NDs, affecting both neural and muscle tissues [\[258\]](#page-24-14). ALS muscle pathology also involves mitochondrial dysfunction, which is caused by defects in mitochondrial dynamics and bioenergetics that are linked to muscle weakness and atrophy [\[4\]](#page-15-3). Mitochondrial abnormalities are observed in the muscle tissues of AD patients, where reduced mitochondrial function contributes to decreased energy production and increased oxidative stress [\[259\]](#page-24-15). In PD,

similar mitochondrial dysfunction is evident in both the substantia nigra neurons and muscle cells, characterized by impaired mitochondrial respiratory chain complex I activity [\[205](#page-22-16)[,260\]](#page-24-16). Across these diseases, mitochondrial dysfunction leads to reduced ATP production, increased ROS, and subsequent muscle fatigue and degeneration.

6.1.3. Protein Aggregation

Abnormal protein aggregation is a critical pathogenic feature in ALS, AD, and PD, affecting both the CNS and peripheral tissues such as muscle [\[261\]](#page-24-17). In ALS, the aggregation of proteins such as TDP-43 and SOD1 within muscle cells parallels their aggregation in motor neurons, leading to muscle pathology and dysfunction $[3,4]$ $[3,4]$. These protein aggregates interfere with normal cellular functions, including proteostasis, autophagy, and intracellular transport, further exacerbating muscle degeneration. In AD, $\rm Aβ$ and tau protein aggregates are found in both the brain and muscle tissues, contributing to cytotoxicity and muscle degeneration [\[262\]](#page-24-18). Similarly, in PD, α -syn aggregates form Lewy bodies in the brain and have been detected in muscle tissue, compromising cellular homeostasis [\[214\]](#page-22-25).

6.1.4. Inflammatory Process

Neuroinflammation plays a significant role in the progression of ALS, AD, and PD and is also evident in the muscle tissues of affected patients [\[222\]](#page-23-6). ALS pathology involves widespread inflammation, with inflammatory markers and immune cells detected in muscle biopsies from patients [\[4\]](#page-15-3). Inflammation exacerbates muscle damage through mechanisms such as increased oxidative stress, the disruption of muscle repair processes, and the release of pro-inflammatory mediators. AD is characterized by chronic inflammation in the brain, with systemic inflammation potentially impacting muscle health and function [\[263\]](#page-24-19). In PD, elevated levels of pro-inflammatory cytokines and immune cell infiltration are observed in both the brain and muscle tissues, contributing to muscle degeneration [\[264,](#page-24-20)[265\]](#page-24-21).

6.2. Specific Differences between the Diseases

6.2.1. Mechanisms of Motor Neuron Degeneration

The primary pathological mechanisms leading to muscle involvement differ significantly among ALS, AD, and PD [\[266\]](#page-24-22). ALS is directly characterized by the degeneration of both upper and lower motor neurons, leading to rapid and severe muscle atrophy and weakness as a primary feature of the disease [\[3\]](#page-15-2). AD is primarily a cognitive disorder, with muscle pathology developing as a result of overall physical decline and reduced activity rather than direct motor neuron degeneration [\[267\]](#page-24-23). In contrast, the motor symptoms of PD, including bradykinesia and rigidity, result in decreased physical activity and secondary muscle wasting [\[268\]](#page-24-24).

6.2.2. Disease Onset and Progression

The onset and progression of muscle pathology vary between ALS, AD, and PD. In ALS, muscle pathology is rapid and severe from the onset, with patients experiencing significant muscle weakness and atrophy early in the disease course [\[4](#page-15-3)[,57\]](#page-17-5). The rapid progression in ALS contrasts sharply with the more gradual muscle involvement seen in AD and PD. AD typically begins with cognitive decline, with muscle involvement occurring later in the disease course as physical function deteriorates [\[269,](#page-25-0)[270\]](#page-25-1). In PD, muscle involvement is gradual, correlating with the slow progression of motor symptoms. Patients often maintain some amount of muscle function for many years following diagnosis.

6.2.3. Therapeutic Responses

Therapeutic strategies targeting muscle pathology also differ among these diseases. ALS therapies focus on slowing disease progression and managing symptoms, with limited options for directly improving muscle health. Exercise may help maintain muscle function for a time, but the rapid progression of muscle atrophy in ALS limits the long-term benefits [\[271\]](#page-25-2). In AD, physical activity is recommended to improve general health and

cognitive function, but specific interventions targeting muscle pathology are less developed [\[272\]](#page-25-3). In PD, exercise and physical therapy are widely recognized for their benefits in improving muscle strength, flexibility, and overall motor function. Aerobic and resistance training are particularly effective in mitigating muscle atrophy and improving the quality of life [\[244](#page-24-1)[,245\]](#page-24-2).

6.2.4. Molecular Pathways

The molecular pathways involved in muscle pathology differ significantly among these disorders. ALS muscle pathology is closely linked to disruptions in RNA processing and protein homeostasis, with mutations in genes such as *SOD1*, *TARDBP*, and *FUS* playing critical roles [\[3](#page-15-2)[,273\]](#page-25-4). AD muscle pathology involves pathways related to APP metabolism and tau phosphorylation, with a lower emphasis on mitochondrial dysfunction than in PD [\[274\]](#page-25-5). Muscle degeneration in PD is characterized by mitochondrial dysfunction and oxidative stress, with specific pathways such as PINK1/Parkin-mediated mitophagy being implicated [\[212,](#page-22-23)[213\]](#page-22-24). These molecular differences underscore the distinct pathological mechanisms driving muscle involvement in each disease.

6.2.5. Role of Different Quality Control Mechanisms

In the context of NDs, both autophagy and the UPS play crucial roles in maintaining cellular homeostasis by managing protein degradation and organelle turnover. However, their functions and specificity can vary depending on the cell types involved and the specific ND in question. Autophagy, which involves the formation of autophagosomes that degrade damaged organelles and misfolded proteins, is particularly important in neurons due to their post-mitotic nature. This process helps prevent the accumulation of toxic materials that could otherwise lead to neurodegeneration [\[275\]](#page-25-6). In contrast, the UPS, which targets proteins for degradation through proteasomes, is essential for the regulation of various cellular processes, including the removal of misfolded proteins and the regulation of signaling pathways critical for neuronal function and survival [\[276\]](#page-25-7). In terms of specificity, the functions of autophagy and the UPS can be quite distinct depending on the disease context and the types of cells affected. For example, in AD, autophagy is involved in degrading Aβ plaques, while the UPS regulates tau protein levels and prevents tau aggregation [\[277](#page-25-8)[–279\]](#page-25-9). Similarly, in PD, both systems manage α-syn aggregates, but their relative contributions can vary based on disease stage and neuronal type [\[280,](#page-25-10)[281\]](#page-25-11). Moreover, while autophagy and the UPS sometimes share substrates, such as ubiquitinated proteins and protein aggregates, their roles can be complementary or divergent depending on the disease. For instance, both pathways are engaged in managing protein aggregates in AD and PD, yet they address different aspects of protein quality control and stress responses [\[279\]](#page-25-9).

In summary, although autophagy and the UPS both play essential roles in neurodegenerative diseases by handling misfolded proteins and damaged organelles, their functions can be specific to different cell types and disease contexts. They share some substrates, particularly those related to protein aggregation, but their mechanisms and impact can vary significantly depending on the type of ND and the specific cellular processes involved [\[282\]](#page-25-12).

7. Conclusions

The involvement of muscle tissue in NDs such as ALS, AD, and PD presents a complex interplay of pathogenic mechanisms that significantly impact patient health and quality of life. Across these conditions, common muscular anomalies such as atrophy, mitochondrial dysfunction, protein aggregation, and inflammation are observed, each contributing to disease progression and symptomatology. However, the underlying molecular pathways and the extent of muscle involvement vary, underlining the importance of disease-specific approaches in understanding and managing these anomalies.

Therapeutic strategies targeting muscle pathology in these diseases show promise in mitigating symptoms and enhancing patient outcomes. Pharmacological interventions aimed at improving mitochondrial function, reducing protein aggregates, and modulating inflammation offer a potential avenue for ameliorating muscle dysfunction. Additionally, physical therapies, including personalized exercise regimens and innovative modalities such as electrical stimulation, have demonstrated beneficial effects in maintaining muscle mass, improving strength, and enhancing overall motor function.

Future research must continue to elucidate the precise mechanisms driving muscle pathology in ALS, AD, and PD to refine these therapeutic approaches. Integrating pharmacological and physical therapies, along with personalized nutritional plans, could provide a comprehensive strategy to address muscle health in NDs. By improving our understanding of muscle involvement and optimizing treatment modalities, we can improve the quality of life for patients and potentially slow the progression of these debilitating conditions.

Author Contributions: E.D. and C.V. carried out the literature review, conceptualization, and writing of the manuscript. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Conflicts of Interest: The authors declare no conflicts of interest.

References

- 1. Dugger, B.N.; Dickson, D.W. Pathology of Neurodegenerative Diseases. *Cold Spring Harb. Perspect. Biol.* **2017**, *9*, a028035. [\[CrossRef\]](https://doi.org/10.1101/cshperspect.a028035)
- 2. Villa, C.; Paudel, Y.N.; Piperi, C. New Insights into Molecular Mechanisms Underlying Neurodegenerative Disorders. *Brain Sci.* **2022**, *12*, 1190.
- 3. Duranti, E.; Villa, C. Molecular Investigations of Protein Aggregation in the Pathogenesis of Amyotrophic Lateral Sclerosis. *Int. J. Mol. Sci.* **2022**, *24*, 704. [\[CrossRef\]](https://doi.org/10.3390/ijms24010704) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36614144)
- 4. Duranti, E.; Villa, C. Muscle Involvement in Amyotrophic Lateral Sclerosis: Understanding the Pathogenesis and Advancing Therapeutics. *Biomolecules* **2023**, *13*, 1582. [\[CrossRef\]](https://doi.org/10.3390/biom13111582) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38002264)
- 5. Bastin, C.; Giacomelli, F.; Miévis, F.; Lemaire, C.; Guillaume, B.; Salmon, E. Anosognosia in Mild Cognitive Impairment: Lack of Awareness of Memory Difficulties Characterizes Prodromal Alzheimer's Disease. *Front. Psychiatry* **2021**, *12*, 631518. [\[CrossRef\]](https://doi.org/10.3389/fpsyt.2021.631518)
- 6. Scheltens, P.; De Strooper, B.; Kivipelto, M.; Holstege, H.; Chételat, G.; Teunissen, C.E.; Cummings, J.; van der Flier, W.M. Alzheimer's disease. *Lancet* **2021**, *397*, 1577–1590. [\[CrossRef\]](https://doi.org/10.1016/S0140-6736(20)32205-4)
- 7. Váradi, C. Clinical Features of Parkinson's Disease: The Evolution of Critical Symptoms. *Biology* **2020**, *9*, 103. [\[CrossRef\]](https://doi.org/10.3390/biology9050103)
- 8. Palanisamy, C.P.; Pei, J.; Alugoju, P.; Anthikapalli, N.V.A.; Jayaraman, S.; Veeraraghavan, V.P.; Gopathy, S.; Roy, J.R.; Janaki, C.S.; Thalamati, D.; et al. New strategies of neurodegenerative disease treatment with extracellular vesicles (EVs) derived from mesenchymal stem cells (MSCs). *Theranostics* **2023**, *13*, 4138–4165. [\[CrossRef\]](https://doi.org/10.7150/thno.83066) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37554286)
- 9. Gitler, A.D.; Dhillon, P.; Shorter, J. Neurodegenerative disease: Models, mechanisms, and a new hope. *Dis. Model Mech.* **2017**, *10*, 499–502. [\[CrossRef\]](https://doi.org/10.1242/dmm.030205)
- 10. Mallucci, G.R.; Klenerman, D.; Rubinsztein, D.C. Developing Therapies for Neurodegenerative Disorders: Insights from Protein Aggregation and Cellular Stress Responses. *Annu. Rev. Cell Dev. Biol.* **2020**, *36*, 165–189. [\[CrossRef\]](https://doi.org/10.1146/annurev-cellbio-040320-120625)
- 11. Schaffert, L.N.; Carter, W.G. Do Post-Translational Modifications Influence Protein Aggregation in Neurodegenerative Diseases: A Systematic Review. *Brain Sci.* **2020**, *10*, 232. [\[CrossRef\]](https://doi.org/10.3390/brainsci10040232)
- 12. Candelise, N.; Scaricamazza, S.; Salvatori, I.; Ferri, A.; Valle, C.; Manganelli, V.; Garofalo, T.; Sorice, M.; Misasi, R. Protein Aggregation Landscape in Neurodegenerative Diseases: Clinical Relevance and Future Applications. *Int. J. Mol. Sci.* **2021**, *22*, 6016. [\[CrossRef\]](https://doi.org/10.3390/ijms22116016) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34199513)
- 13. Duranti, E.; Villa, C. Insights into Dysregulated Neurological Biomarkers in Cancer. *Cancers* **2024**, *16*, 2680. [\[CrossRef\]](https://doi.org/10.3390/cancers16152680) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/39123408)
- 14. Myszczynska, M.A.; Ojamies, P.N.; Lacoste, A.M.B.; Neil, D.; Saffari, A.; Mead, R.; Hautbergue, G.M.; Holbrook, J.D.; Ferraiuolo, L. Applications of machine learning to diagnosis and treatment of neurodegenerative diseases. *Nat. Rev. Neurol.* **2020**, *16*, 440–456. [\[CrossRef\]](https://doi.org/10.1038/s41582-020-0377-8) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32669685)
- 15. Durães, F.; Pinto, M.; Sousa, E. Old Drugs as New Treatments for Neurodegenerative Diseases. *Pharmaceuticals* **2018**, *11*, 44. [\[CrossRef\]](https://doi.org/10.3390/ph11020044) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29751602)
- 16. Burtscher, J.; Millet, G.P.; Place, N.; Kayser, B.; Zanou, N. The Muscle-Brain Axis and Neurodegenerative Diseases: The Key Role of Mitochondria in Exercise-Induced Neuroprotection. *Int. J. Mol. Sci.* **2021**, *22*, 6479. [\[CrossRef\]](https://doi.org/10.3390/ijms22126479)
- 17. Bian, X.; Wang, Q.; Wang, Y.; Lou, S. The function of previously unappreciated exerkines secreted by muscle in regulation of neurodegenerative diseases. *Front. Mol. Neurosci.* **2023**, *16*, 1305208. [\[CrossRef\]](https://doi.org/10.3389/fnmol.2023.1305208)
- 18. Shefner, J.M.; Musaro, A.; Ngo, S.T.; Lunetta, C.; Steyn, F.J.; Robitaille, R.; De Carvalho, M.; Rutkove, S.; Ludolph, A.C.; Dupuis, L. Skeletal muscle in amyotrophic lateral sclerosis. *Brain* **2023**, *146*, 4425–4436. [\[CrossRef\]](https://doi.org/10.1093/brain/awad202)
- 19. Beeri, M.S.; Leugrans, S.E.; Delbono, O.; Bennett, D.A.; Buchman, A.S. Sarcopenia is associated with incident Alzheimer's dementia, mild cognitive impairment, and cognitive decline. *J. Am. Geriatr. Soc.* **2021**, *69*, 1826–1835. [\[CrossRef\]](https://doi.org/10.1111/jgs.17206)
- 20. Raleigh, S.M.; Orchard, K.J.A. Sarcopenia as a Risk Factor for Alzheimer's Disease: Genetic and Epigenetic Perspectives. *Genes* **2024**, *15*, 561. [\[CrossRef\]](https://doi.org/10.3390/genes15050561)
- 21. Halli-Tierney, A.D.; Luker, J.; Carroll, D.G. Parkinson Disease. *Am. Fam. Physician* **2020**, *102*, 679–691.
- 22. Ferreira-Sánchez, M.D.R.; Moreno-Verdú, M.; Cano-de-la-Cuerda, R. Quantitative Measurement of Rigidity in Parkinson's Disease: A Systematic Review. *Sensors* **2020**, *20*, 880. [\[CrossRef\]](https://doi.org/10.3390/s20030880) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32041374)
- 23. Waters, D.L.; Baumgartner, R.N.; Garry, P.J.; Vellas, B. Advantages of dietary, exercise-related, and therapeutic interventions to prevent and treat sarcopenia in adult patients: An update. *Clin. Interv. Aging* **2010**, *5*, 259–270. [\[CrossRef\]](https://doi.org/10.2147/CIA.S6920) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/20852673)
- 24. Ahmadian-Moghadam, H.; Sadat-Shirazi, M.S.; Zarrindast, M.R. Therapeutic potential of stem cells for treatment of neurodegenerative diseases. *Biotechnol. Lett.* **2020**, *42*, 1073–1101. [\[CrossRef\]](https://doi.org/10.1007/s10529-020-02886-1) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32342435)
- 25. Golpich, M.; Amini, E.; Mohamed, Z.; Azman Ali, R.; Mohamed Ibrahim, N.; Ahmadiani, A. Mitochondrial Dysfunction and Biogenesis in Neurodegenerative diseases: Pathogenesis and Treatment. *CNS Neurosci. Ther.* **2017**, *23*, 5–22. [\[CrossRef\]](https://doi.org/10.1111/cns.12655)
- 26. Frontera, W.R.; Ochala, J. Skeletal muscle: A brief review of structure and function. *Calcif. Tissue Int.* **2015**, *96*, 183–195. [\[CrossRef\]](https://doi.org/10.1007/s00223-014-9915-y) 27. Vainshtein, A.; Sandri, M. Signaling Pathways That Control Muscle Mass. *Int. J. Mol. Sci.* **2020**, *21*, 4759. [\[CrossRef\]](https://doi.org/10.3390/ijms21134759)
- 28. Francaux, M.; Deldicque, L. Exercise and the control of muscle mass in human. *Pflug. Arch.* **2019**, *471*, 397–411. [\[CrossRef\]](https://doi.org/10.1007/s00424-018-2217-x)
- 29. Rasmussen, B.B.; Phillips, S.M. Contractile and nutritional regulation of human muscle growth. *Exerc. Sport. Sci. Rev.* **2003**, *31*, 127–131. [\[CrossRef\]](https://doi.org/10.1097/00003677-200307000-00005)
- 30. Westerblad, H.; Bruton, J.D.; Katz, A. Skeletal muscle: Energy metabolism, fiber types, fatigue and adaptability. *Exp. Cell Res.* **2010**, *316*, 3093–3099. [\[CrossRef\]](https://doi.org/10.1016/j.yexcr.2010.05.019)
- 31. Zurlo, F.; Nemeth, P.M.; Choksi, R.M.; Sesodia, S.; Ravussin, E. Whole-body energy metabolism and skeletal muscle biochemical characteristics. *Metabolism* **1994**, *43*, 481–486. [\[CrossRef\]](https://doi.org/10.1016/0026-0495(94)90081-7)
- 32. Kamei, Y.; Hatazawa, Y.; Uchitomi, R.; Yoshimura, R.; Miura, S. Regulation of Skeletal Muscle Function by Amino Acids. *Nutrients* **2020**, *12*, 261. [\[CrossRef\]](https://doi.org/10.3390/nu12010261)
- 33. Fukada, S.I. The roles of muscle stem cells in muscle injury, atrophy and hypertrophy. *J. Biochem.* **2018**, *163*, 353–358. [\[CrossRef\]](https://doi.org/10.1093/jb/mvy019) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29394360)
- 34. Duranti, E.; Villa, C. Influence of DUX4 Expression in Facioscapulohumeral Muscular Dystrophy and Possible Treatments. *Int. J. Mol. Sci.* **2023**, *24*, 9503. [\[CrossRef\]](https://doi.org/10.3390/ijms24119503) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37298453)
- 35. Morgan, J.; Partridge, T. Skeletal muscle in health and disease. *Dis. Models Mech.* **2020**, *13*, dmm042192. [\[CrossRef\]](https://doi.org/10.1242/dmm.042192) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32066552)
- 36. Shirakawa, T.; Toyono, T.; Inoue, A.; Matsubara, T.; Kawamoto, T.; Kokabu, S. Factors Regulating or Regulated by Myogenic Regulatory Factors in Skeletal Muscle Stem Cells. *Cells* **2022**, *11*, 1493. [\[CrossRef\]](https://doi.org/10.3390/cells11091493)
- 37. Zammit, P.S. Function of the myogenic regulatory factors Myf5, MyoD, Myogenin and MRF4 in skeletal muscle, satellite cells and regenerative myogenesis. *Semin. Cell Dev. Biol.* **2017**, *72*, 19–32. [\[CrossRef\]](https://doi.org/10.1016/j.semcdb.2017.11.011)
- 38. Cretoiu, D.; Pavelescu, L.; Duica, F.; Radu, M.; Suciu, N.; Cretoiu, S.M. Myofibers. *Adv. Exp. Med. Biol.* **2018**, *1088*, 23–46. [\[CrossRef\]](https://doi.org/10.1007/978-981-13-1435-3_2)
- 39. Mukund, K.; Subramaniam, S. Skeletal muscle: A review of molecular structure and function, in health and disease. *Wiley Interdiscip. Rev. Syst. Biol. Med.* **2020**, *12*, e1462. [\[CrossRef\]](https://doi.org/10.1002/wsbm.1462)
- 40. Wang, H.; Guan, L.; Deng, M. Recent progress of the genetics of amyotrophic lateral sclerosis and challenges of gene therapy. *Front. Neurosci.* **2023**, *17*, 1170996. [\[CrossRef\]](https://doi.org/10.3389/fnins.2023.1170996)
- 41. Irwin, K.E.; Sheth, U.; Wong, P.C.; Gendron, T.F. Fluid biomarkers for amyotrophic lateral sclerosis: A review. *Mol. Neurodegener.* **2024**, *19*, 9. [\[CrossRef\]](https://doi.org/10.1186/s13024-023-00685-6)
- 42. Kubat, G.B.; Picone, P. Skeletal muscle dysfunction in amyotrophic lateral sclerosis: A mitochondrial perspective and therapeutic approaches. *Neurol. Sci.* **2024**, *45*, 4121–4131. [\[CrossRef\]](https://doi.org/10.1007/s10072-024-07508-6)
- 43. Mitchell, J.D.; Borasio, G.D. Amyotrophic lateral sclerosis. *Lancet* **2007**, *369*, 2031–2041. [\[CrossRef\]](https://doi.org/10.1016/S0140-6736(07)60944-1) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/17574095)
- 44. Renton, A.E.; Chiò, A.; Traynor, B.J. State of play in amyotrophic lateral sclerosis genetics. *Nat. Neurosci.* **2014**, *17*, 17–23. [\[CrossRef\]](https://doi.org/10.1038/nn.3584)
- 45. Prasad, A.; Bharathi, V.; Sivalingam, V.; Girdhar, A.; Patel, B.K. Molecular Mechanisms of TDP-43 Misfolding and Pathology in Amyotrophic Lateral Sclerosis. *Front. Mol. Neurosci.* **2019**, *12*, 25. [\[CrossRef\]](https://doi.org/10.3389/fnmol.2019.00025) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30837838)
- 46. Cunha-Oliveira, T.; Montezinho, L.; Mendes, C.; Firuzi, O.; Saso, L.; Oliveira, P.J.; Silva, F.S.G. Oxidative Stress in Amyotrophic Lateral Sclerosis: Pathophysiology and Opportunities for Pharmacological Intervention. *Oxid. Med. Cell. Longev.* **2020**, *2020*, 5021694. [\[CrossRef\]](https://doi.org/10.1155/2020/5021694)
- 47. Motataianu, A.; Serban, G.; Barcutean, L.; Balasa, R. Oxidative Stress in Amyotrophic Lateral Sclerosis: Synergy of Genetic and Environmental Factors. *Int. J. Mol. Sci.* **2022**, *23*, 9339. [\[CrossRef\]](https://doi.org/10.3390/ijms23169339) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36012603)
- 48. Huai, J.; Zhang, Z. Structural Properties and Interaction Partners of Familial ALS-Associated SOD1 Mutants. *Front. Neurol.* **2019**, *10*, 527. [\[CrossRef\]](https://doi.org/10.3389/fneur.2019.00527)
- 49. Cozzolino, M.; Carrì, M.T. Mitochondrial dysfunction in ALS. *Prog. Neurobiol.* **2012**, *97*, 54–66. [\[CrossRef\]](https://doi.org/10.1016/j.pneurobio.2011.06.003)
- 50. Smith, E.F.; Shaw, P.J.; De Vos, K.J. The role of mitochondria in amyotrophic lateral sclerosis. *Neurosci. Lett.* **2019**, *710*, 132933. [\[CrossRef\]](https://doi.org/10.1016/j.neulet.2017.06.052)
- 51. Van Den Bosch, L.; Van Damme, P.; Bogaert, E.; Robberecht, W. The role of excitotoxicity in the pathogenesis of amyotrophic lateral sclerosis. *Biochim. Biophys. Acta* **2006**, *1762*, 1068–1082. [\[CrossRef\]](https://doi.org/10.1016/j.bbadis.2006.05.002)
- 52. Arnold, F.J.; Putka, A.F.; Raychaudhuri, U.; Hsu, S.; Bedlack, R.S.; Bennett, C.L.; La Spada, A.R. Revisiting Glutamate Excitotoxicity in Amyotrophic Lateral Sclerosis and Age-Related Neurodegeneration. *Int. J. Mol. Sci.* **2024**, *25*, 5587. [\[CrossRef\]](https://doi.org/10.3390/ijms25115587)
- 53. Li, Q.; Haney, M.S. The role of glia in protein aggregation. *Neurobiol. Dis.* **2020**, *143*, 105015. [\[CrossRef\]](https://doi.org/10.1016/j.nbd.2020.105015) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32663608)
- 54. Liu, J.; Wang, F. Role of Neuroinflammation in Amyotrophic Lateral Sclerosis: Cellular Mechanisms and Therapeutic Implications. *Front. Immunol.* **2017**, *8*, 1005. [\[CrossRef\]](https://doi.org/10.3389/fimmu.2017.01005) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28871262)
- 55. Castellanos-Montiel, M.J.; Chaineau, M.; Durcan, T.M. The Neglected Genes of ALS: Cytoskeletal Dynamics Impact Synaptic Degeneration in ALS. *Front. Cell. Neurosci.* **2020**, *14*, 594975. [\[CrossRef\]](https://doi.org/10.3389/fncel.2020.594975)
- 56. Loeffler, J.P.; Picchiarelli, G.; Dupuis, L.; Gonzalez De Aguilar, J.L. The Role of Skeletal Muscle in Amyotrophic Lateral Sclerosis. *Brain Pathol.* **2016**, *26*, 227–236. [\[CrossRef\]](https://doi.org/10.1111/bpa.12350)
- 57. Pikatza-Menoio, O.; Elicegui, A.; Bengoetxea, X.; Naldaiz-Gastesi, N.; López de Munain, A.; Gerenu, G.; Gil-Bea, F.J.; Alonso-Martín, S. The Skeletal Muscle Emerges as a New Disease Target in Amyotrophic Lateral Sclerosis. *J. Pers. Med.* **2021**, *11*, 671. [\[CrossRef\]](https://doi.org/10.3390/jpm11070671) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34357138)
- 58. Biedasek, K.; Andres, J.; Mai, K.; Adams, S.; Spuler, S.; Fielitz, J.; Spranger, J. Skeletal muscle 11beta-HSD1 controls glucocorticoidinduced proteolysis and expression of E3 ubiquitin ligases atrogin-1 and MuRF-1. *PLoS ONE* **2011**, *6*, e16674. [\[CrossRef\]](https://doi.org/10.1371/journal.pone.0016674)
- 59. Al-Sarraj, S.; King, A.; Cleveland, M.; Pradat, P.F.; Corse, A.; Rothstein, J.D.; Leigh, P.N.; Abila, B.; Bates, S.; Wurthner, J.; et al. Mitochondrial abnormalities and low grade inflammation are present in the skeletal muscle of a minority of patients with amyotrophic lateral sclerosis; an observational myopathology study. *Acta Neuropathol. Commun.* **2014**, *2*, 165. [\[CrossRef\]](https://doi.org/10.1186/s40478-014-0165-z)
- 60. Galbiati, M.; Crippa, V.; Rusmini, P.; Cristofani, R.; Cicardi, M.E.; Giorgetti, E.; Onesto, E.; Messi, E.; Poletti, A. ALS-related misfolded protein management in motor neurons and muscle cells. *Neurochem. Int.* **2014**, *79*, 70–78. [\[CrossRef\]](https://doi.org/10.1016/j.neuint.2014.10.007)
- 61. Vicencio, E.; Beltrán, S.; Labrador, L.; Manque, P.; Nassif, M.; Woehlbier, U. Implications of Selective Autophagy Dysfunction for ALS Pathology. *Cells* **2020**, *9*, 381. [\[CrossRef\]](https://doi.org/10.3390/cells9020381)
- 62. Colasuonno, F.; Price, R.; Moreno, S. Upper and Lower Motor Neurons and the Skeletal Muscle: Implication for Amyotrophic Lateral Sclerosis (ALS). In *Roles of Skeletal Muscle in Organ Development—Prenatal Interdependence among Cells, Tissues, and Organs*; Kablar, B., Ed.; Springer: Cham, Switzerland, 2023; Volume 236, pp. 111–129. [\[CrossRef\]](https://doi.org/10.1007/978-3-031-38215-4_5)
- 63. Amin, A.; Perera, N.D.; Beart, P.M.; Turner, B.J.; Shabanpoor, F. Amyotrophic Lateral Sclerosis and Autophagy: Dysfunction and Therapeutic Targeting. *Cells* **2020**, *9*, 2413. [\[CrossRef\]](https://doi.org/10.3390/cells9112413) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33158177)
- 64. Nixon, R.A. The role of autophagy in neurodegenerative disease. *Nat. Med.* **2013**, *19*, 983–997. [\[CrossRef\]](https://doi.org/10.1038/nm.3232)
- 65. Fleming, A.; Bourdenx, M.; Fujimaki, M.; Karabiyik, C.; Krause, G.J.; Lopez, A.; Martín-Segura, A.; Puri, C.; Scrivo, A.; Skidmore, J.; et al. The different autophagy degradation pathways and neurodegeneration. *Neuron* **2022**, *110*, 935–966. [\[CrossRef\]](https://doi.org/10.1016/j.neuron.2022.01.017) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35134347)
- 66. Schoser, B.G.; Wehling, S.; Blottner, D. Cell death and apoptosis-related proteins in muscle biopsies of sporadic amyotrophic lateral sclerosis and polyneuropathy. *Muscle Nerve* **2001**, *24*, 1083–1089. [\[CrossRef\]](https://doi.org/10.1002/mus.1114)
- 67. Manzano, R.; Toivonen, J.M.; Moreno-Martínez, L.; de la Torre, M.; Moreno-García, L.; López-Royo, T.; Molina, N.; Zaragoza, P.; Calvo, A.C.; Osta, R. What skeletal muscle has to say in amyotrophic lateral sclerosis: Implications for therapy. *Br. J. Pharmacol.* **2021**, *178*, 1279–1297. [\[CrossRef\]](https://doi.org/10.1111/bph.15276)
- 68. Xiao, Y.; Ma, C.; Yi, J.; Wu, S.; Luo, G.; Xu, X.; Lin, P.H.; Sun, J.; Zhou, J. Suppressed autophagy flux in skeletal muscle of an amyotrophic lateral sclerosis mouse model during disease progression. *Physiol. Rep.* **2015**, *3*, e12271. [\[CrossRef\]](https://doi.org/10.14814/phy2.12271)
- 69. Chen, W.; Guo, L.; Li, M.; Wei, C.; Li, S.; Xu, R. The pathogenesis of amyotrophic lateral sclerosis: Mitochondrial dysfunction, protein misfolding and epigenetics. *Brain Res.* **2022**, *1786*, 147904. [\[CrossRef\]](https://doi.org/10.1016/j.brainres.2022.147904) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35390335)
- 70. Chan, Y.; Alix, J.J.P.; Neuwirth, C.; Barkhaus, P.E.; Castro, J.; Jenkins, T.M.; McDermott, C.J.; Shaw, P.J.; de Carvalho, M.; Nandedkar, S.; et al. Reinnervation as measured by the motor unit size index is associated with preservation of muscle strength in amyotrophic lateral sclerosis, but not all muscles reinnervate. *Muscle Nerve* **2022**, *65*, 203–210. [\[CrossRef\]](https://doi.org/10.1002/mus.27444) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34687220)
- 71. Leermakers, P.A.; Skov, M.; Riisager, A.; Nielsen, O.B.; Pedersen, T.H. Alterations in fast-twitch muscle membrane conductance regulation do not explain decreased muscle function of SOD1(G93A) rats. *Muscle Nerve* **2021**, *64*, 755–764. [\[CrossRef\]](https://doi.org/10.1002/mus.27413)
- 72. Quessada, C.; Bouscary, A.; René, F.; Valle, C.; Ferri, A.; Ngo, S.T.; Loeffler, J.P. Skeletal Muscle Metabolism: Origin or Prognostic Factor for Amyotrophic Lateral Sclerosis (ALS) Development? *Cells* **2021**, *10*, 1449. [\[CrossRef\]](https://doi.org/10.3390/cells10061449)
- 73. Anderson, G. Amyotrophic Lateral Sclerosis Pathoetiology and Pathophysiology: Roles of Astrocytes, Gut Microbiome, and Muscle Interactions via the Mitochondrial Melatonergic Pathway, with Disruption by Glyphosate-Based Herbicides. *Int. J. Mol. Sci.* **2022**, *24*, 587. [\[CrossRef\]](https://doi.org/10.3390/ijms24010587) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36614029)
- 74. Peggion, C.; Scalcon, V.; Massimino, M.L.; Nies, K.; Lopreiato, R.; Rigobello, M.P.; Bertoli, A. SOD1 in ALS: Taking Stock in Pathogenic Mechanisms and the Role of Glial and Muscle Cells. *Antioxidants* **2022**, *11*, 614. [\[CrossRef\]](https://doi.org/10.3390/antiox11040614) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35453299)
- 75. Dupuis, L.; Loeffler, J.P. Neuromuscular junction destruction during amyotrophic lateral sclerosis: Insights from transgenic models. *Curr. Opin. Pharmacol.* **2009**, *9*, 341–346. [\[CrossRef\]](https://doi.org/10.1016/j.coph.2009.03.007)
- 76. Lloyd, E.M.; Pinniger, G.J.; Murphy, R.M.; Grounds, M.D. Slow or fast: Implications of myofibre type and associated differences for manifestation of neuromuscular disorders. *Acta Physiol.* **2023**, *238*, e14012. [\[CrossRef\]](https://doi.org/10.1111/apha.14012)
- 77. Nijssen, J.; Comley, L.H.; Hedlund, E. Motor neuron vulnerability and resistance in amyotrophic lateral sclerosis. *Acta Neuropathol.* **2017**, *133*, 863–885. [\[CrossRef\]](https://doi.org/10.1007/s00401-017-1708-8) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28409282)
- 78. Venditti, P.; Di Meo, S. The Role of Reactive Oxygen Species in the Life Cycle of the Mitochondrion. *Int. J. Mol. Sci.* **2020**, *21*, 2173. [\[CrossRef\]](https://doi.org/10.3390/ijms21062173)
- 79. Pollari, E.; Goldsteins, G.; Bart, G.; Koistinaho, J.; Giniatullin, R. The role of oxidative stress in degeneration of the neuromuscular junction in amyotrophic lateral sclerosis. *Front. Cell. Neurosci.* **2014**, *8*, 131. [\[CrossRef\]](https://doi.org/10.3389/fncel.2014.00131)
- 80. Lu, C.H.; Allen, K.; Oei, F.; Leoni, E.; Kuhle, J.; Tree, T.; Fratta, P.; Sharma, N.; Sidle, K.; Howard, R.; et al. Systemic inflammatory response and neuromuscular involvement in amyotrophic lateral sclerosis. *Neurol. Neuroimmunol. Neuroinflamm.* **2016**, *3*, e244. [\[CrossRef\]](https://doi.org/10.1212/NXI.0000000000000244)
- 81. McCombe, P.A.; Henderson, R.D. The Role of immune and inflammatory mechanisms in ALS. *Curr. Mol. Med.* **2011**, *11*, 246–254. [\[CrossRef\]](https://doi.org/10.2174/156652411795243450)
- 82. Jensen, L.; Jørgensen, L.H.; Bech, R.D.; Frandsen, U.; Schrøder, H.D. Skeletal Muscle Remodelling as a Function of Disease Progression in Amyotrophic Lateral Sclerosis. *Biomed. Res. Int.* **2016**, *2016*, 5930621. [\[CrossRef\]](https://doi.org/10.1155/2016/5930621)
- 83. Liu, W.; Chakkalakal, J.V. The Composition, Development, and Regeneration of Neuromuscular Junctions. *Curr. Top. Dev. Biol.* **2018**, *126*, 99–124. [\[CrossRef\]](https://doi.org/10.1016/bs.ctdb.2017.08.005) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29305005)
- 84. Martineau, É.; Di Polo, A.; Vande Velde, C.; Robitaille, R. Dynamic neuromuscular remodeling precedes motor-unit loss in a mouse model of ALS. *eLife* **2018**, *7*, e41973. [\[CrossRef\]](https://doi.org/10.7554/eLife.41973) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30320556)
- 85. Dadon-Nachum, M.; Melamed, E.; Offen, D. The "dying-back" phenomenon of motor neurons in ALS. *J. Mol. Neurosci.* **2011**, *43*, 470–477. [\[CrossRef\]](https://doi.org/10.1007/s12031-010-9467-1) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21057983)
- 86. Lepore, E.; Casola, I.; Dobrowolny, G.; Musarò, A. Neuromuscular Junction as an Entity of Nerve-Muscle Communication. *Cells* **2019**, *8*, 906. [\[CrossRef\]](https://doi.org/10.3390/cells8080906)
- 87. Le Grand, F.; Rudnicki, M. Satellite and stem cells in muscle growth and repair. *Development* **2007**, *134*, 3953–3957. [\[CrossRef\]](https://doi.org/10.1242/dev.005934)
- 88. Lynch, K. Optimizing pharmacologic treatment for ALS to improve outcomes and quality of life. *Am. J. Manag. Care* **2023**, *29*, S112–S119. [\[CrossRef\]](https://doi.org/10.37765/ajmc.2023.89389)
- 89. Hinchcliffe, M.; Smith, A. Riluzole: Real-world evidence supports significant extension of median survival times in patients with amyotrophic lateral sclerosis. *Degener. Neurol. Neuromuscul. Dis.* **2017**, *7*, 61–70. [\[CrossRef\]](https://doi.org/10.2147/DNND.S135748)
- 90. Masrori, P.; Van Damme, P. Amyotrophic lateral sclerosis: A clinical review. *Eur. J. Neurol.* **2020**, *27*, 1918–1929. [\[CrossRef\]](https://doi.org/10.1111/ene.14393) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32526057)
- 91. Nagoshi, N.; Nakashima, H.; Fehlings, M.G. Riluzole as a neuroprotective drug for spinal cord injury: From bench to bedside. *Molecules* **2015**, *20*, 7775–7789. [\[CrossRef\]](https://doi.org/10.3390/molecules20057775)
- 92. Storch, A.; Burkhardt, K.; Ludolph, A.C.; Schwarz, J. Protective effects of riluzole on dopamine neurons: Involvement of oxidative stress and cellular energy metabolism. *J. Neurochem.* **2000**, *75*, 2259–2269. [\[CrossRef\]](https://doi.org/10.1046/j.1471-4159.2000.0752259.x)
- 93. Duranti, E.; Cordani, N.; Villa, C. Edaravone: A Novel Possible Drug for Cancer Treatment? *Int. J. Mol. Sci.* **2024**, *25*, 1633. [\[CrossRef\]](https://doi.org/10.3390/ijms25031633) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38338912)
- 94. Cha, S.J.; Kim, K. Effects of the Edaravone, a Drug Approved for the Treatment of Amyotrophic Lateral Sclerosis, on Mitochondrial Function and Neuroprotection. *Antioxidants* **2022**, *11*, 195. [\[CrossRef\]](https://doi.org/10.3390/antiox11020195) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35204078)
- 95. Morén, C.; deSouza, R.M.; Giraldo, D.M.; Uff, C. Antioxidant Therapeutic Strategies in Neurodegenerative Diseases. *Int. J. Mol. Sci.* **2022**, *23*, 9328. [\[CrossRef\]](https://doi.org/10.3390/ijms23169328) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36012599)
- 96. Yamamoto, T.; Yuki, S.; Watanabe, T.; Mitsuka, M.; Saito, K.I.; Kogure, K. Delayed neuronal death prevented by inhibition of increased hydroxyl radical formation in a transient cerebral ischemia. *Brain Res.* **1997**, *762*, 240–242. [\[CrossRef\]](https://doi.org/10.1016/S0006-8993(97)00490-3)
- 97. Tzeplaeff, L.; Wilfling, S.; Requardt, M.V.; Herdick, M. Current State and Future Directions in the Therapy of ALS. *Cells* **2023**, *12*, 1523. [\[CrossRef\]](https://doi.org/10.3390/cells12111523) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37296644)
- 98. Jiang, J.; Wang, Y.; Deng, M. New developments and opportunities in drugs being trialed for amyotrophic lateral sclerosis from 2020 to 2022. *Front. Pharmacol.* **2022**, *13*, 1054006. [\[CrossRef\]](https://doi.org/10.3389/fphar.2022.1054006)
- 99. Khalaf, K.; Tornese, P.; Cocco, A.; Albanese, A. Tauroursodeoxycholic acid: A potential therapeutic tool in neurodegenerative diseases. *Transl. Neurodegener.* **2022**, *11*, 33. [\[CrossRef\]](https://doi.org/10.1186/s40035-022-00307-z)
- 100. Johnson, S.A.; Fang, T.; De Marchi, F.; Neel, D.; Van Weehaeghe, D.; Berry, J.D.; Paganoni, S. Pharmacotherapy for Amyotrophic Lateral Sclerosis: A Review of Approved and Upcoming Agents. *Drugs* **2022**, *82*, 1367–1388. [\[CrossRef\]](https://doi.org/10.1007/s40265-022-01769-1)
- 101. Nikitin, D.; Makam, A.N.; Suh, K.; McKenna, A.; Carlson, J.J.; Richardson, M.; Rind, D.M.; Pearson, S.D. The effectiveness and value of AMX0035 and oral edaravone for amyotrophic lateral sclerosis: A summary from the Institute for Clinical and Economic Review's Midwest Comparative Effectiveness Public Advisory Council. *J. Manag. Care Spec. Pharm.* **2023**, *29*, 216–221. [\[CrossRef\]](https://doi.org/10.18553/jmcp.2023.29.2.216)
- 102. Paganoni, S.; Hendrix, S.; Dickson, S.P.; Knowlton, N.; Macklin, E.A.; Berry, J.D.; Elliott, M.A.; Maiser, S.; Karam, C.; Caress, J.B.; et al. Long-term survival of participants in the CENTAUR trial of sodium phenylbutyrate-taurursodiol in amyotrophic lateral sclerosis. *Muscle Nerve* **2021**, *63*, 31–39. [\[CrossRef\]](https://doi.org/10.1002/mus.27091)
- 103. Oliveira Santos, M.; de Carvalho, M. Profiling tofersen as a treatment of superoxide dismutase 1 amyotrophic lateral sclerosis. *Expert. Rev. Neurother.* **2024**, *24*, 549–553. [\[CrossRef\]](https://doi.org/10.1080/14737175.2024.2355983) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38758193)
- 104. Saini, A.; Chawla, P.A. Breaking barriers with tofersen: Enhancing therapeutic opportunities in amyotrophic lateral sclerosis. *Eur. J. Neurol.* **2024**, *31*, e16140. [\[CrossRef\]](https://doi.org/10.1111/ene.16140)
- 105. Miller, T.M.; Cudkowicz, M.E.; Genge, A.; Shaw, P.J.; Sobue, G.; Bucelli, R.C.; Chiò, A.; Van Damme, P.; Ludolph, A.C.; Glass, J.D.; et al. Trial of Antisense Oligonucleotide Tofersen for SOD1 ALS. *N. Engl. J. Med.* **2022**, *387*, 1099–1110. [\[CrossRef\]](https://doi.org/10.1056/NEJMoa2204705) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36129998)
- 106. Gillon, A.; Nielsen, K.; Steel, C.; Cornwall, J.; Sheard, P. Exercise attenuates age-associated changes in motoneuron number, nucleocytoplasmic transport proteins and neuromuscular health. *Geroscience* **2018**, *40*, 177–192. [\[CrossRef\]](https://doi.org/10.1007/s11357-018-0020-4) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29736782)
- 107. Rygiel, K.A.; Picard, M.; Turnbull, D.M. The ageing neuromuscular system and sarcopenia: A mitochondrial perspective. *J. Physiol.* **2016**, *594*, 4499–4512. [\[CrossRef\]](https://doi.org/10.1113/JP271212)
- 108. Ferri, E.; Marzetti, E.; Calvani, R.; Picca, A.; Cesari, M.; Arosio, B. Role of Age-Related Mitochondrial Dysfunction in Sarcopenia. *Int. J. Mol. Sci.* **2020**, *21*, 5236. [\[CrossRef\]](https://doi.org/10.3390/ijms21155236)
- 109. Sailani, M.R.; Halling, J.F.; Møller, H.D.; Lee, H.; Plomgaard, P.; Pilegaard, H.; Snyder, M.P.; Regenberg, B. Lifelong physical activity is associated with promoter hypomethylation of genes involved in metabolism, myogenesis, contractile properties and oxidative stress resistance in aged human skeletal muscle. *Sci. Rep.* **2019**, *9*, 3272. [\[CrossRef\]](https://doi.org/10.1038/s41598-018-37895-8)
- 110. Gallo, V.; Vanacore, N.; Bueno-de-Mesquita, H.B.; Vermeulen, R.; Brayne, C.; Pearce, N.; Wark, P.A.; Ward, H.A.; Ferrari, P.; Jenab, M.; et al. Physical activity and risk of Amyotrophic Lateral Sclerosis in a prospective cohort study. *Eur. J. Epidemiol.* **2016**, *31*, 255–266. [\[CrossRef\]](https://doi.org/10.1007/s10654-016-0119-9)
- 111. Carreras, I.; Yuruker, S.; Aytan, N.; Hossain, L.; Choi, J.K.; Jenkins, B.G.; Kowall, N.W.; Dedeoglu, A. Moderate exercise delays the motor performance decline in a transgenic model of ALS. *Brain Res.* **2010**, *1313*, 192–201. [\[CrossRef\]](https://doi.org/10.1016/j.brainres.2009.11.051)
- 112. Tseng, C.; Sinha, K.; Pan, H.; Cui, Y.; Guo, P.; Lin, C.Y.; Yang, F.; Deng, Z.; Eltzschig, H.K.; Lu, A.; et al. Markers of Accelerated Skeletal Muscle Regenerative Response in Murphy Roths Large Mice: Characteristics of Muscle Progenitor Cells and Circulating Factors. *Stem Cells* **2019**, *37*, 357–367. [\[CrossRef\]](https://doi.org/10.1002/stem.2957)
- 113. Bennett, E.J.; Mead, R.J.; Azzouz, M.; Shaw, P.J.; Grierson, A.J. Early detection of motor dysfunction in the SOD1G93A mouse model of Amyotrophic Lateral Sclerosis (ALS) using home cage running wheels. *PLoS ONE* **2014**, *9*, e107918. [\[CrossRef\]](https://doi.org/10.1371/journal.pone.0107918) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25268710)
- 114. Pradhan, J.; Noakes, P.G.; Bellingham, M.C. The Role of Altered BDNF/TrkB Signaling in Amyotrophic Lateral Sclerosis. *Front. Cell. Neurosci.* **2019**, *13*, 368. [\[CrossRef\]](https://doi.org/10.3389/fncel.2019.00368) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31456666)
- 115. Wu, Y.; Rakotoarisoa, M.; Angelov, B.; Deng, Y.; Angelova, A. Self-Assembled Nanoscale Materials for Neuronal Regeneration: A Focus on BDNF Protein and Nucleic Acid Biotherapeutic Delivery. *Nanomaterials* **2022**, *12*, 2267. [\[CrossRef\]](https://doi.org/10.3390/nano12132267) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35808102)
- 116. Park, D.; Kwak, S.G.; Park, J.S.; Choo, Y.J.; Chang, M.C. Can Therapeutic Exercise Slow Down Progressive Functional Decline in Patients With Amyotrophic Lateral Sclerosis? A Meta-Analysis. *Front. Neurol.* **2020**, *11*, 853. [\[CrossRef\]](https://doi.org/10.3389/fneur.2020.00853) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32903629)
- 117. Musarò, A.; Dobrowolny, G.; Cambieri, C.; Onesti, E.; Ceccanti, M.; Frasca, V.; Pisano, A.; Cerbelli, B.; Lepore, E.; Ruffolo, G.; et al. Neuromuscular magnetic stimulation counteracts muscle decline in ALS patients: Results of a randomized, double-blind, controlled study. *Sci. Rep.* **2019**, *9*, 2837. [\[CrossRef\]](https://doi.org/10.1038/s41598-019-39313-z)
- 118. Crous-Bou, M.; Minguillón, C.; Gramunt, N.; Molinuevo, J.L. Alzheimer's disease prevention: From risk factors to early intervention. *Alzheimers Res. Ther.* **2017**, *9*, 71. [\[CrossRef\]](https://doi.org/10.1186/s13195-017-0297-z)
- 119. Anand, R.; Gill, K.D.; Mahdi, A.A. Therapeutics of Alzheimer's disease: Past, present and future. *Neuropharmacology* **2014**, *76 Pt A*, 27–50. [\[CrossRef\]](https://doi.org/10.1016/j.neuropharm.2013.07.004)
- 120. DeTure, M.A.; Dickson, D.W. The neuropathological diagnosis of Alzheimer's disease. *Mol. Neurodegener.* **2019**, *14*, 32. [\[CrossRef\]](https://doi.org/10.1186/s13024-019-0333-5)
- 121. Villa, C. Biomarkers for Alzheimer's Disease: Where Do We Stand and Where Are We Going? *J. Pers. Med.* **2020**, *10*, 238. [\[CrossRef\]](https://doi.org/10.3390/jpm10040238)
- 122. Ratan, Y.; Rajput, A.; Maleysm, S.; Pareek, A.; Jain, V.; Kaur, R.; Singh, G. An Insight into Cellular and Molecular Mechanisms Underlying the Pathogenesis of Neurodegeneration in Alzheimer's Disease. *Biomedicines* **2023**, *11*, 1398. [\[CrossRef\]](https://doi.org/10.3390/biomedicines11051398)
- 123. Lane, C.A.; Hardy, J.; Schott, J.M. Alzheimer's disease. *Eur. J. Neurol.* **2018**, *25*, 59–70. [\[CrossRef\]](https://doi.org/10.1111/ene.13439) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28872215)
- 124. D'Argenio, V.; Sarnataro, D. New Insights into the Molecular Bases of Familial Alzheimer's Disease. *J. Pers. Med.* **2020**, *10*, 26. [\[CrossRef\]](https://doi.org/10.3390/jpm10020026) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32325882)
- 125. Ryman, D.C.; Acosta-Baena, N.; Aisen, P.S.; Bird, T.; Danek, A.; Fox, N.C.; Goate, A.; Frommelt, P.; Ghetti, B.; Langbaum, J.B.; et al. Symptom onset in autosomal dominant Alzheimer disease: A systematic review and meta-analysis. *Neurology* **2014**, *83*, 253–260. [\[CrossRef\]](https://doi.org/10.1212/WNL.0000000000000596) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24928124)
- 126. Kawai, M.; Kalaria, R.N.; Cras, P.; Siedlak, S.L.; Velasco, M.E.; Shelton, E.R.; Chan, H.W.; Greenberg, B.D.; Perry, G. Degeneration of vascular muscle cells in cerebral amyloid angiopathy of Alzheimer disease. *Brain Res.* **1993**, *623*, 142–146. [\[CrossRef\]](https://doi.org/10.1016/0006-8993(93)90021-E)
- 127. Ogawa, Y.; Kaneko, Y.; Sato, T.; Shimizu, S.; Kanetaka, H.; Hanyu, H. Sarcopenia and Muscle Functions at Various Stages of Alzheimer Disease. *Front. Neurol.* **2018**, *9*, 710. [\[CrossRef\]](https://doi.org/10.3389/fneur.2018.00710)
- 128. Boyle, P.A.; Buchman, A.S.; Wilson, R.S.; Leurgans, S.E.; Bennett, D.A. Association of muscle strength with the risk of Alzheimer disease and the rate of cognitive decline in community-dwelling older persons. *Arch. Neurol.* **2009**, *66*, 1339–1344. [\[CrossRef\]](https://doi.org/10.1001/archneurol.2009.240)
- 129. Mani, S.; Dubey, R.; Lai, I.C.; Babu, M.A.; Tyagi, S.; Swargiary, G.; Mody, D.; Singh, M.; Agarwal, S.; Iqbal, D.; et al. Oxidative Stress and Natural Antioxidants: Back and Forth in the Neurological Mechanisms of Alzheimer's Disease. *J. Alzheimers Dis.* **2023**, *96*, 877–912. [\[CrossRef\]](https://doi.org/10.3233/JAD-220700)
- 130. Misrani, A.; Tabassum, S.; Yang, L. Mitochondrial Dysfunction and Oxidative Stress in Alzheimer's Disease. *Front. Aging Neurosci.* **2021**, *13*, 617588. [\[CrossRef\]](https://doi.org/10.3389/fnagi.2021.617588)
- 131. Picone, P.; Nuzzo, D.; Caruana, L.; Scafidi, V.; Di Carlo, M. Mitochondrial dysfunction: Different routes to Alzheimer's disease therapy. *Oxid. Med. Cell. Longev.* **2014**, *2014*, 780179. [\[CrossRef\]](https://doi.org/10.1155/2014/780179)
- 132. Bell, S.M.; Barnes, K.; De Marco, M.; Shaw, P.J.; Ferraiuolo, L.; Blackburn, D.J.; Venneri, A.; Mortiboys, H. Mitochondrial Dysfunction in Alzheimer's Disease: A Biomarker of the Future? *Biomedicines* **2021**, *9*, 63. [\[CrossRef\]](https://doi.org/10.3390/biomedicines9010063)
- 133. Tian, Q.; Bilgel, M.; Walker, K.A.; Moghekar, A.R.; Fishbein, K.W.; Spencer, R.G.; Resnick, S.M.; Ferrucci, L. Skeletal muscle mitochondrial function predicts cognitive impairment and is associated with biomarkers of Alzheimer's disease and neurodegeneration. *Alzheimers Dement.* **2023**, *19*, 4436–4445. [\[CrossRef\]](https://doi.org/10.1002/alz.13388) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37530130)
- 134. Turkseven, C.H.; Buyukakilli, B.; Balli, E.; Yetkin, D.; Erdal, M.E.; Yilmaz, S.G.; Sahin, L. Effects of Huperzin-A on the Beta-amyloid accumulation in the brain and skeletal muscle cells of a rat model for Alzheimer's disease. *Life Sci.* **2017**, *184*, 47–57. [\[CrossRef\]](https://doi.org/10.1016/j.lfs.2017.07.012)
- 135. Yamaguchi, H.; Yamazaki, T.; Lemere, C.A.; Frosch, M.P.; Selkoe, D.J. Beta amyloid is focally deposited within the outer basement membrane in the amyloid angiopathy of Alzheimer's disease. An immunoelectron microscopic study. *Am. J. Pathol.* **1992**, *141*, 249–259. [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/1632466)
- 136. Sehar, U.; Rawat, P.; Reddy, A.P.; Kopel, J.; Reddy, P.H. Amyloid Beta in Aging and Alzheimer's Disease. *Int. J. Mol. Sci.* **2022**, *23*, 12924. [\[CrossRef\]](https://doi.org/10.3390/ijms232112924)
- 137. Roher, A.E.; Esh, C.L.; Kokjohn, T.A.; Castaño, E.M.; Van Vickle, G.D.; Kalback, W.M.; Patton, R.L.; Luehrs, D.C.; Daugs, I.D.; Kuo, Y.M.; et al. Amyloid beta peptides in human plasma and tissues and their significance for Alzheimer's disease. *Alzheimers Dement.* **2009**, *5*, 18–29. [\[CrossRef\]](https://doi.org/10.1016/j.jalz.2008.10.004)
- 138. Thakur, S.; Dhapola, R.; Sarma, P.; Medhi, B.; Reddy, D.H. Neuroinflammation in Alzheimer's Disease: Current Progress in Molecular Signaling and Therapeutics. *Inflammation* **2023**, *46*, 1–17. [\[CrossRef\]](https://doi.org/10.1007/s10753-022-01721-1)
- 139. Marino, M.; Scuderi, F.; Provenzano, C.; Bartoccioni, E. Skeletal muscle cells: From local inflammatory response to active immunity. *Gene Ther.* **2011**, *18*, 109–116. [\[CrossRef\]](https://doi.org/10.1038/gt.2010.124) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/20927136)
- 140. Oudbier, S.J.; Goh, J.; Looijaard, S.; Reijnierse, E.M.; Meskers, C.G.M.; Maier, A.B. Pathophysiological Mechanisms Explaining the Association Between Low Skeletal Muscle Mass and Cognitive Function. *J. Gerontol. A Biol. Sci. Med. Sci.* **2022**, *77*, 1959–1968. [\[CrossRef\]](https://doi.org/10.1093/gerona/glac121)
- 141. Moylan, J.S.; Reid, M.B. Oxidative stress, chronic disease, and muscle wasting. *Muscle Nerve* **2007**, *35*, 411–429. [\[CrossRef\]](https://doi.org/10.1002/mus.20743)
- 142. S˛edzikowska, A.; Szablewski, L. Insulin and Insulin Resistance in Alzheimer's Disease. *Int. J. Mol. Sci.* **2021**, *22*, 9987. [\[CrossRef\]](https://doi.org/10.3390/ijms22189987)
- 143. Akhtar, A.; Sah, S.P. Insulin signaling pathway and related molecules: Role in neurodegeneration and Alzheimer's disease. *Neurochem. Int.* **2020**, *135*, 104707. [\[CrossRef\]](https://doi.org/10.1016/j.neuint.2020.104707) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32092326)
- 144. Bosco, D.; Fava, A.; Plastino, M.; Montalcini, T.; Pujia, A. Possible implications of insulin resistance and glucose metabolism in Alzheimer's disease pathogenesis. *J. Cell. Mol. Med.* **2011**, *15*, 1807–1821. [\[CrossRef\]](https://doi.org/10.1111/j.1582-4934.2011.01318.x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21435176)
- 145. Dewanjee, S.; Chakraborty, P.; Bhattacharya, H.; Chacko, L.; Singh, B.; Chaudhary, A.; Javvaji, K.; Pradhan, S.R.; Vallamkondu, J.; Dey, A.; et al. Altered glucose metabolism in Alzheimer's disease: Role of mitochondrial dysfunction and oxidative stress. *Free Radic. Biol. Med.* **2022**, *193*, 134–157. [\[CrossRef\]](https://doi.org/10.1016/j.freeradbiomed.2022.09.032)
- 146. Ezkurdia, A.; Ramírez, M.J.; Solas, M. Metabolic Syndrome as a Risk Factor for Alzheimer's Disease: A Focus on Insulin Resistance. *Int. J. Mol. Sci.* **2023**, *24*, 4354. [\[CrossRef\]](https://doi.org/10.3390/ijms24054354)
- 147. Arosio, B.; Calvani, R.; Ferri, E.; Coelho-Junior, H.J.; Carandina, A.; Campanelli, F.; Ghiglieri, V.; Marzetti, E.; Picca, A. Sarcopenia and Cognitive Decline in Older Adults: Targeting the Muscle-Brain Axis. *Nutrients* **2023**, *15*, 1853. [\[CrossRef\]](https://doi.org/10.3390/nu15081853) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37111070)
- 148. Filardi, M.; Barone, R.; Bramato, G.; Nigro, S.; Tafuri, B.; Frisullo, M.E.; Zecca, C.; Tortelli, R.; Logroscino, G. The Relationship between Muscle Strength and Cognitive Performance across Alzheimer's Disease Clinical Continuum. *Front. Neurol.* **2022**, *13*, 833087. [\[CrossRef\]](https://doi.org/10.3389/fneur.2022.833087)
- 149. Sugimoto, T.; Ono, R.; Murata, S.; Saji, N.; Matsui, Y.; Niida, S.; Toba, K.; Sakurai, T. Prevalence and associated factors of sarcopenia in elderly subjects with amnestic mild cognitive impairment or Alzheimer disease. *Curr. Alzheimer Res.* **2016**, *13*, 718–726. [\[CrossRef\]](https://doi.org/10.2174/1567205013666160211124828)
- 150. Beckett, T.L.; Niedowicz, D.M.; Studzinski, C.M.; Weidner, A.M.; Webb, R.L.; Holler, C.J.; Ahmed, R.R.; LeVine, H., 3rd; Murphy, M.P. Effects of nonsteroidal anti-inflammatory drugs on amyloid-beta pathology in mouse skeletal muscle. *Neurobiol. Dis.* **2010**, *39*, 449–456. [\[CrossRef\]](https://doi.org/10.1016/j.nbd.2010.05.018)
- 151. Alturki, M.; Beyer, I.; Mets, T.; Bautmans, I. Impact of drugs with anti-inflammatory effects on skeletal muscle and inflammation: A systematic literature review. *Exp. Gerontol.* **2018**, *114*, 33–49. [\[CrossRef\]](https://doi.org/10.1016/j.exger.2018.10.011)
- 152. Mantle, D.; Heaton, R.A.; Hargreaves, I.P. Coenzyme Q10, Ageing and the Nervous System: An Overview. *Antioxidants* **2021**, *11*, 2. [\[CrossRef\]](https://doi.org/10.3390/antiox11010002)
- 153. Gutierrez-Mariscal, F.M.; Arenas-de Larriva, A.P.; Limia-Perez, L.; Romero-Cabrera, J.L.; Yubero-Serrano, E.M.; López-Miranda, J. Coenzyme Q(10) Supplementation for the Reduction of Oxidative Stress: Clinical Implications in the Treatment of Chronic Diseases. *Int. J. Mol. Sci.* **2020**, *21*, 7870. [\[CrossRef\]](https://doi.org/10.3390/ijms21217870) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33114148)
- 154. Vints, W.A.J.; Levin, O.; van Griensven, M.; Vlaeyen, J.W.S.; Masiulis, N.; Verbunt, J.; van Laake-Geelen, C.C.M. Neuromuscular electrical stimulation to combat cognitive aging in people with spinal cord injury: Protocol for a single case experimental design study. *BMC Neurol.* **2024**, *24*, 197. [\[CrossRef\]](https://doi.org/10.1186/s12883-024-03699-9)
- 155. Ahmad, S.S.; Ahmad, K.; Lee, E.J.; Lee, Y.H.; Choi, I. Implications of Insulin-Like Growth Factor-1 in Skeletal Muscle and Various Diseases. *Cells* **2020**, *9*, 1773. [\[CrossRef\]](https://doi.org/10.3390/cells9081773)
- 156. Zemva, J.; Schubert, M. The role of neuronal insulin/insulin-like growth factor-1 signaling for the pathogenesis of Alzheimer's disease: Possible therapeutic implications. *CNS Neurol. Disord. Drug Targets* **2014**, *13*, 322–337. [\[CrossRef\]](https://doi.org/10.2174/18715273113126660141) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24059318)
- 157. Van Broeck, B.; Van Broeckhoven, C.; Kumar-Singh, S. Current insights into molecular mechanisms of Alzheimer disease and their implications for therapeutic approaches. *Neurodegener. Dis.* **2007**, *4*, 349–365. [\[CrossRef\]](https://doi.org/10.1159/000105156)
- 158. Franco-Bocanegra, D.K.; McAuley, C.; Nicoll, J.A.R.; Boche, D. Molecular Mechanisms of Microglial Motility: Changes in Ageing and Alzheimer's Disease. *Cells* **2019**, *8*, 639. [\[CrossRef\]](https://doi.org/10.3390/cells8060639)
- 159. Cioffi, F.; Adam, R.H.I.; Broersen, K. Molecular Mechanisms and Genetics of Oxidative Stress in Alzheimer's Disease. *J. Alzheimers Dis.* **2019**, *72*, 981–1017. [\[CrossRef\]](https://doi.org/10.3233/JAD-190863) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31744008)
- 160. Tanganelli, F.; Meinke, P.; Hofmeister, F.; Jarmusch, S.; Baber, L.; Mehaffey, S.; Hintze, S.; Ferrari, U.; Neuerburg, C.; Kammerlander, C.; et al. Type-2 muscle fiber atrophy is associated with sarcopenia in elderly men with hip fracture. *Exp. Gerontol.* **2021**, *144*, 111171. [\[CrossRef\]](https://doi.org/10.1016/j.exger.2020.111171)
- 161. Moon, Y.; Choi, Y.J.; Kim, J.O.; Han, S.H. Muscle profile and cognition in patients with Alzheimer's disease dementia. *Neurol. Sci.* **2018**, *39*, 1861–1866. [\[CrossRef\]](https://doi.org/10.1007/s10072-018-3505-0)
- 162. Veronese, N.; Solmi, M.; Basso, C.; Smith, L.; Soysal, P. Role of physical activity in ameliorating neuropsychiatric symptoms in Alzheimer disease: A narrative review. *Int. J. Geriatr. Psychiatry* **2019**, *34*, 1316–1325. [\[CrossRef\]](https://doi.org/10.1002/gps.4962)
- 163. Santos-Lozano, A.; Pareja-Galeano, H.; Sanchis-Gomar, F.; Quindós-Rubial, M.; Fiuza-Luces, C.; Cristi-Montero, C.; Emanuele, E.; Garatachea, N.; Lucia, A. Physical Activity and Alzheimer Disease: A Protective Association. *Mayo Clin. Proc.* **2016**, *91*, 999–1020. [\[CrossRef\]](https://doi.org/10.1016/j.mayocp.2016.04.024) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27492909)
- 164. Brasure, M.; Desai, P.; Davila, H.; Nelson, V.A.; Calvert, C.; Jutkowitz, E.; Butler, M.; Fink, H.A.; Ratner, E.; Hemmy, L.S.; et al. Physical Activity Interventions in Preventing Cognitive Decline and Alzheimer-Type Dementia: A Systematic Review. *Ann. Intern. Med.* **2018**, *168*, 30–38. [\[CrossRef\]](https://doi.org/10.7326/M17-1528) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29255839)
- 165. Wang, R.; Holsinger, R.M.D. Exercise-induced brain-derived neurotrophic factor expression: Therapeutic implications for Alzheimer's dementia. *Ageing Res. Rev.* **2018**, *48*, 109–121. [\[CrossRef\]](https://doi.org/10.1016/j.arr.2018.10.002) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30326283)
- 166. Pereira, T.; Cipriano, I.; Costa, T.; Saraiva, M.; Martins, A. Exercise, ageing and cognitive function—Effects of a personalized physical exercise program in the cognitive function of older adults. *Physiol. Behav.* **2019**, *202*, 8–13. [\[CrossRef\]](https://doi.org/10.1016/j.physbeh.2019.01.018)
- 167. Polito, R.; Di Meo, I.; Barbieri, M.; Daniele, A.; Paolisso, G.; Rizzo, M.R. Adiponectin Role in Neurodegenerative Diseases: Focus on Nutrition Review. *Int. J. Mol. Sci.* **2020**, *21*, 9255. [\[CrossRef\]](https://doi.org/10.3390/ijms21239255)
- 168. Dvorak, R.V.; Poehlman, E.T. Appendicular skeletal muscle mass, physical activity, and cognitive status in patients with Alzheimer's disease. *Neurology* **1998**, *51*, 1386–1390. [\[CrossRef\]](https://doi.org/10.1212/WNL.51.5.1386)
- 169. Wang, Y.; Liu, Q.; Quan, H.; Kang, S.G.; Huang, K.; Tong, T. Nutraceuticals in the Prevention and Treatment of the Muscle Atrophy. *Nutrients* **2021**, *13*, 1914. [\[CrossRef\]](https://doi.org/10.3390/nu13061914) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34199575)
- 170. Thomas, J.; Thomas, C.J.; Radcliffe, J.; Itsiopoulos, C. Omega-3 Fatty Acids in Early Prevention of Inflammatory Neurodegenerative Disease: A Focus on Alzheimer's Disease. *Biomed. Res. Int.* **2015**, *2015*, 172801. [\[CrossRef\]](https://doi.org/10.1155/2015/172801)
- 171. Nolan, J.M.; Power, R.; Howard, A.N.; Bergin, P.; Roche, W.; Prado-Cabrero, A.; Pope, G.; Cooke, J.; Power, T.; Mulcahy, R. Supplementation With Carotenoids, Omega-3 Fatty Acids, and Vitamin E Has a Positive Effect on the Symptoms and Progression of Alzheimer's Disease. *J. Alzheimers Dis.* **2022**, *90*, 233–249. [\[CrossRef\]](https://doi.org/10.3233/JAD-220556)
- 172. De Sousa, O.V.; Mendes, J.; Amaral, T.F. Nutritional and Functional Indicators and Their Association With Mortality Among Older Adults With Alzheimer's Disease. *Am. J. Alzheimers Dis. Other Dement.* **2020**, *35*, 1533317520907168. [\[CrossRef\]](https://doi.org/10.1177/1533317520907168)
- 173. Feraco, P.; Gagliardo, C.; La Tona, G.; Bruno, E.; D'Angelo, C.; Marrale, M.; Del Poggio, A.; Malaguti, M.C.; Geraci, L.; Baschi, R.; et al. Imaging of Substantia Nigra in Parkinson's Disease: A Narrative Review. *Brain Sci.* **2021**, *11*, 769. [\[CrossRef\]](https://doi.org/10.3390/brainsci11060769) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34207681)
- 174. Vázquez-Vélez, G.E.; Zoghbi, H.Y. Parkinson's Disease Genetics and Pathophysiology. *Annu. Rev. Neurosci.* **2021**, *44*, 87–108. [\[CrossRef\]](https://doi.org/10.1146/annurev-neuro-100720-034518) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34236893)
- 175. Kumar, S.; Goyal, L.; Singh, S. Tremor and Rigidity in Patients with Parkinson's Disease: Emphasis on Epidemiology, Pathophysiology and Contributing Factors. *CNS Neurol. Disord. Drug Targets* **2022**, *21*, 596–609. [\[CrossRef\]](https://doi.org/10.2174/1871527320666211006142100)
- 176. Martin, I.; Dawson, V.L.; Dawson, T.M. Recent advances in the genetics of Parkinson's disease. *Annu. Rev. Genom. Hum. Genet.* **2011**, *12*, 301–325. [\[CrossRef\]](https://doi.org/10.1146/annurev-genom-082410-101440)
- 177. Angelopoulou, E.; Bougea, A.; Papageorgiou, S.G.; Villa, C. Psychosis in Parkinson's Disease: A Lesson from Genetics. *Genes* **2022**, *13*, 1099. [\[CrossRef\]](https://doi.org/10.3390/genes13061099) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35741861)
- 178. Cherian, A.; Divya, K.P. Genetics of Parkinson's disease. *Acta Neurol. Belg.* **2020**, *120*, 1297–1305. [\[CrossRef\]](https://doi.org/10.1007/s13760-020-01473-5)
- 179. Ajith, C.; Divya, K.P.; Asish, V. Parkinson's disease—Genetic cause. *Curr. Opin. Neurol.* **2023**, *36*, 292–301. [\[CrossRef\]](https://doi.org/10.1097/wco.0000000000001167)
- 180. Menšíková, K.; Matěj, R.; Colosimo, C.; Rosales, R.; Tučková, L.; Ehrmann, J.; Hraboš, D.; Kolaříková, K.; Vodička, R.; Vrtěl, R.; et al. Lewy body disease or diseases with Lewy bodies? *npj Park. Dis.* **2022**, *8*, 3. [\[CrossRef\]](https://doi.org/10.1038/s41531-021-00273-9)
- 181. Schulz-Schaeffer, W.J. The synaptic pathology of alpha-synuclein aggregation in dementia with Lewy bodies, Parkinson's disease and Parkinson's disease dementia. *Acta Neuropathol.* **2010**, *120*, 131–143. [\[CrossRef\]](https://doi.org/10.1007/s00401-010-0711-0)
- 182. Jęśko, H.; Lenkiewicz, A.M.; Wilkaniec, A.; Adamczyk, A. The interplay between parkin and alpha-synuclein; possible implications for the pathogenesis of Parkinson's disease. *Acta Neurobiol. Exp.* **2019**, *79*, 276–289. [\[CrossRef\]](https://doi.org/10.21307/ane-2019-026)
- 183. Zhang, G.; Xia, Y.; Wan, F.; Ma, K.; Guo, X.; Kou, L.; Yin, S.; Han, C.; Liu, L.; Huang, J.; et al. New Perspectives on Roles of Alpha-Synuclein in Parkinson's Disease. *Front. Aging Neurosci.* **2018**, *10*, 370. [\[CrossRef\]](https://doi.org/10.3389/fnagi.2018.00370) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30524265)
- 184. Moradi Vastegani, S.; Nasrolahi, A.; Ghaderi, S.; Belali, R.; Rashno, M.; Farzaneh, M.; Khoshnam, S.E. Mitochondrial Dysfunction and Parkinson's Disease: Pathogenesis and Therapeutic Strategies. *Neurochem. Res.* **2023**, *48*, 2285–2308. [\[CrossRef\]](https://doi.org/10.1007/s11064-023-03904-0) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36943668)
- 185. Alqahtani, T.; Deore, S.L.; Kide, A.A.; Shende, B.A.; Sharma, R.; Dadarao Chakole, R.; Nemade, L.S.; Kishor Kale, N.; Borah, S.; Shrikant Deokar, S.; et al. Mitochondrial dysfunction and oxidative stress in Alzheimer's disease, and Parkinson's disease, Huntington's disease and Amyotrophic Lateral Sclerosis—An updated review. *Mitochondrion* **2023**, *71*, 83–92. [\[CrossRef\]](https://doi.org/10.1016/j.mito.2023.05.007) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37269968)
- 186. Weng, M.; Xie, X.; Liu, C.; Lim, K.L.; Zhang, C.W.; Li, L. The Sources of Reactive Oxygen Species and Its Possible Role in the Pathogenesis of Parkinson's Disease. *Park. Dis.* **2018**, *2018*, 9163040. [\[CrossRef\]](https://doi.org/10.1155/2018/9163040)
- 187. Zuo, L.; Motherwell, M.S. The impact of reactive oxygen species and genetic mitochondrial mutations in Parkinson's disease. *Gene* **2013**, *532*, 18–23. [\[CrossRef\]](https://doi.org/10.1016/j.gene.2013.07.085)
- 188. Oczkowska, A.; Kozubski, W.; Lianeri, M.; Dorszewska, J. Mutations in PRKN and SNCA Genes Important for the Progress of Parkinson's Disease. *Curr. Genom.* **2013**, *14*, 502–517. [\[CrossRef\]](https://doi.org/10.2174/1389202914666131210205839)
- 189. Lotankar, S.; Prabhavalkar, K.S.; Bhatt, L.K. Biomarkers for Parkinson's Disease: Recent Advancement. *Neurosci. Bull.* **2017**, *33*, 585–597. [\[CrossRef\]](https://doi.org/10.1007/s12264-017-0183-5)
- 190. Karabiyik, C.; Lee, M.J.; Rubinsztein, D.C. Autophagy impairment in Parkinson's disease. *Essays Biochem.* **2017**, *61*, 711–720. [\[CrossRef\]](https://doi.org/10.1042/ebc20170023)
- 191. Hou, X.; Watzlawik, J.O.; Fiesel, F.C.; Springer, W. Autophagy in Parkinson's Disease. *J. Mol. Biol.* **2020**, *432*, 2651–2672. [\[CrossRef\]](https://doi.org/10.1016/j.jmb.2020.01.037)
- 192. Behl, T.; Kumar, S.; Althafar, Z.M.; Sehgal, A.; Singh, S.; Sharma, N.; Badavath, V.N.; Yadav, S.; Bhatia, S.; Al-Harrasi, A.; et al. Exploring the Role of Ubiquitin-Proteasome System in Parkinson's Disease. *Mol. Neurobiol.* **2022**, *59*, 4257–4273. [\[CrossRef\]](https://doi.org/10.1007/s12035-022-02851-1)
- 193. Liang, Y.; Zhong, G.; Ren, M.; Sun, T.; Li, Y.; Ye, M.; Ma, C.; Guo, Y.; Liu, C. The Role of Ubiquitin-Proteasome System and Mitophagy in the Pathogenesis of Parkinson's Disease. *Neuromol. Med.* **2023**, *25*, 471–488. [\[CrossRef\]](https://doi.org/10.1007/s12017-023-08755-0) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37698835)
- 194. Troncoso-Escudero, P.; Parra, A.; Nassif, M.; Vidal, R.L. Outside in: Unraveling the Role of Neuroinflammation in the Progression of Parkinson's Disease. *Front. Neurol.* **2018**, *9*, 860. [\[CrossRef\]](https://doi.org/10.3389/fneur.2018.00860) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30459700)
- 195. Lee, Y.; Lee, S.; Chang, S.C.; Lee, J. Significant roles of neuroinflammation in Parkinson's disease: Therapeutic targets for PD prevention. *Arch. Pharm. Res.* **2019**, *42*, 416–425. [\[CrossRef\]](https://doi.org/10.1007/s12272-019-01133-0) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30830660)
- 196. Collins, L.M.; Toulouse, A.; Connor, T.J.; Nolan, Y.M. Contributions of central and systemic inflammation to the pathophysiology of Parkinson's disease. *Neuropharmacology* **2012**, *62*, 2154–2168. [\[CrossRef\]](https://doi.org/10.1016/j.neuropharm.2012.01.028)
- 197. Surendranathan, A.; Su, L.; Mak, E.; Passamonti, L.; Hong, Y.T.; Arnold, R.; Vázquez Rodríguez, P.; Bevan-Jones, W.R.; Brain, S.A.E.; Fryer, T.D.; et al. Early microglial activation and peripheral inflammation in dementia with Lewy bodies. *Brain* **2018**, *141*, 3415–3427. [\[CrossRef\]](https://doi.org/10.1093/brain/awy265)
- 198. Rui, Q.; Ni, H.; Li, D.; Gao, R.; Chen, G. The Role of LRRK2 in Neurodegeneration of Parkinson Disease. *Curr. Neuropharmacol.* **2018**, *16*, 1348–1357. [\[CrossRef\]](https://doi.org/10.2174/1570159X16666180222165418) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29473513)
- 199. Kluss, J.H.; Mamais, A.; Cookson, M.R. LRRK2 links genetic and sporadic Parkinson's disease. *Biochem. Soc. Trans.* **2019**, *47*, 651–661. [\[CrossRef\]](https://doi.org/10.1042/BST20180462)
- 200. Tolosa, E.; Vila, M.; Klein, C.; Rascol, O. LRRK2 in Parkinson disease: Challenges of clinical trials. *Nat. Rev. Neurol.* **2020**, *16*, 97–107. [\[CrossRef\]](https://doi.org/10.1038/s41582-019-0301-2)
- 201. Lavin, K.M.; Ge, Y.; Sealfon, S.C.; Nair, V.D.; Wilk, K.; McAdam, J.S.; Windham, S.T.; Kumar, P.L.; McDonald, M.N.; Bamman, M.M. Rehabilitative Impact of Exercise Training on Human Skeletal Muscle Transcriptional Programs in Parkinson's Disease. *Front. Physiol.* **2020**, *11*, 653. [\[CrossRef\]](https://doi.org/10.3389/fphys.2020.00653)
- 202. Kaufmann, H.; Goldstein, D.S. Autonomic dysfunction in Parkinson disease. *Handb. Clin. Neurol.* **2013**, *117*, 259–278. [\[CrossRef\]](https://doi.org/10.1016/b978-0-444-53491-0.00021-3)
- 203. Gdynia, H.J.; Sperfeld, A.D.; Unrath, A.; Ludolph, A.C.; Sabolek, M.; Storch, A.; Kassubek, J. Histopathological analysis of skeletal muscle in patients with Parkinson's disease and 'dropped head'/'bent spine' syndrome. *Park. Relat. Disord.* **2009**, *15*, 633–639. [\[CrossRef\]](https://doi.org/10.1016/j.parkreldis.2009.06.003) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/19574084)
- 204. Gamborg, M.; Hvid, L.G.; Thrue, C.; Johansson, S.; Franzén, E.; Dalgas, U.; Langeskov-Christensen, M. Muscle Strength and Power in People With Parkinson Disease: A Systematic Review and Meta-analysis. *J. Neurol. Phys. Ther.* **2023**, *47*, 3–15. [\[CrossRef\]](https://doi.org/10.1097/NPT.0000000000000421) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36318503)
- 205. Bose, A.; Beal, M.F. Mitochondrial dysfunction in Parkinson's disease. *J. Neurochem.* **2016**, *139* (Suppl. S1), 216–231. [\[CrossRef\]](https://doi.org/10.1111/jnc.13731) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27546335)
- 206. Penn, A.M.; Roberts, T.; Hodder, J.; Allen, P.S.; Zhu, G.; Martin, W.R. Generalized mitochondrial dysfunction in Parkinson's disease detected by magnetic resonance spectroscopy of muscle. *Neurology* **1995**, *45*, 2097–2099. [\[CrossRef\]](https://doi.org/10.1212/WNL.45.11.2097)
- 207. Banerjee, R.; Starkov, A.A.; Beal, M.F.; Thomas, B. Mitochondrial dysfunction in the limelight of Parkinson's disease pathogenesis. *Biochim. Biophys. Acta* **2009**, *1792*, 651–663. [\[CrossRef\]](https://doi.org/10.1016/j.bbadis.2008.11.007)
- 208. Schulz, J.B.; Beal, M.F. Mitochondrial dysfunction in movement disorders. *Curr. Opin. Neurol.* **1994**, *7*, 333–339. [\[CrossRef\]](https://doi.org/10.1097/00019052-199408000-00010)
- 209. Yang, Q.; Wang, Y.; Zhao, C.; Pang, S.; Lu, J.; Chan, P. α-Synuclein aggregation causes muscle atrophy through neuromuscular junction degeneration. *J. Cachexia Sarcopenia Muscle* **2023**, *14*, 226–242. [\[CrossRef\]](https://doi.org/10.1002/jcsm.13123)
- 210. Huang, Z.; Chen, Y.; Zhang, Y. Mitochondrial reactive oxygen species cause major oxidative mitochondrial DNA damages and repair pathways. *J. Biosci.* **2020**, *45*, 84. [\[CrossRef\]](https://doi.org/10.1007/s12038-020-00055-0)
- 211. Islam, M.T. Oxidative stress and mitochondrial dysfunction-linked neurodegenerative disorders. *Neurol. Res.* **2017**, *39*, 73–82. [\[CrossRef\]](https://doi.org/10.1080/01616412.2016.1251711)
- 212. Truban, D.; Hou, X.; Caulfield, T.R.; Fiesel, F.C.; Springer, W. PINK1, Parkin, and Mitochondrial Quality Control: What can we Learn about Parkinson's Disease Pathobiology? *J. Park. Dis.* **2017**, *7*, 13–29. [\[CrossRef\]](https://doi.org/10.3233/JPD-160989)
- 213. Van Humbeeck, C.; Cornelissen, T.; Hofkens, H.; Mandemakers, W.; Gevaert, K.; De Strooper, B.; Vandenberghe, W. Parkin interacts with Ambra1 to induce mitophagy. *J. Neurosci.* **2011**, *31*, 10249–10261. [\[CrossRef\]](https://doi.org/10.1523/JNEUROSCI.1917-11.2011) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21753002)
- 214. Li, Y.Y.; Zhou, T.T.; Zhang, Y.; Chen, N.H.; Yuan, Y.H. Distribution of α-Synuclein Aggregation in the Peripheral Tissues. *Neurochem. Res.* **2022**, *47*, 3627–3634. [\[CrossRef\]](https://doi.org/10.1007/s11064-022-03586-0) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35348944)
- 215. Sahoo, S.; Padhy, A.A.; Kumari, V.; Mishra, P. Role of Ubiquitin-Proteasome and Autophagy-Lysosome Pathways in α-Synuclein Aggregate Clearance. *Mol. Neurobiol.* **2022**, *59*, 5379–5407. [\[CrossRef\]](https://doi.org/10.1007/s12035-022-02897-1)
- 216. Le Guerroué, F.; Youle, R.J. Ubiquitin signaling in neurodegenerative diseases: An autophagy and proteasome perspective. *Cell Death Differ.* **2021**, *28*, 439–454. [\[CrossRef\]](https://doi.org/10.1038/s41418-020-00667-x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33208890)
- 217. Sala, G.; Marinig, D.; Arosio, A.; Ferrarese, C. Role of Chaperone-Mediated Autophagy Dysfunctions in the Pathogenesis of Parkinson's Disease. *Front. Mol. Neurosci.* **2016**, *9*, 157. [\[CrossRef\]](https://doi.org/10.3389/fnmol.2016.00157)
- 218. Basellini, M.J.; Kothuis, J.M.; Comincini, A.; Pezzoli, G.; Cappelletti, G.; Mazzetti, S. Pathological Pathways and Alpha-Synuclein in Parkinson's Disease: A View from the Periphery. *Front. Biosci.* **2023**, *28*, 33. [\[CrossRef\]](https://doi.org/10.31083/j.fbl2802033)
- 219. Mu, L.; Sobotka, S.; Chen, J.; Su, H.; Sanders, I.; Adler, C.H.; Shill, H.A.; Caviness, J.N.; Samanta, J.E.; Beach, T.G. Alpha-synuclein pathology and axonal degeneration of the peripheral motor nerves innervating pharyngeal muscles in Parkinson disease. *J. Neuropathol. Exp. Neurol.* **2013**, *72*, 119–129. [\[CrossRef\]](https://doi.org/10.1097/NEN.0b013e3182801cde)
- 220. Gelpi, E.; Navarro-Otano, J.; Tolosa, E.; Gaig, C.; Compta, Y.; Rey, M.J.; Martí, M.J.; Hernández, I.; Valldeoriola, F.; Reñé, R.; et al. Multiple organ involvement by alpha-synuclein pathology in Lewy body disorders. *Mov. Disord.* **2014**, *29*, 1010–1018. [\[CrossRef\]](https://doi.org/10.1002/mds.25776)
- 221. Wang, T.; Shi, C.; Luo, H.; Zheng, H.; Fan, L.; Tang, M.; Su, Y.; Yang, J.; Mao, C.; Xu, Y. Neuroinflammation in Parkinson's Disease: Triggers, Mechanisms, and Immunotherapies. *Neuroscientist* **2022**, *28*, 364–381. [\[CrossRef\]](https://doi.org/10.1177/1073858421991066)
- 222. Zhang, W.; Xiao, D.; Mao, Q.; Xia, H. Role of neuroinflammation in neurodegeneration development. *Signal Transduct. Target. Ther.* **2023**, *8*, 267. [\[CrossRef\]](https://doi.org/10.1038/s41392-023-01486-5)
- 223. Zhang, H.; Wu, J.; Shen, F.F.; Yuan, Y.S.; Li, X.; Ji, P.; Zhu, L.; Sun, L.; Ding, J.; Niu, Q.; et al. Activated Schwann cells and increased inflammatory cytokines IL-1β, IL-6, and TNF-α in patients' sural nerve are lack of tight relationship with specific sensory disturbances in Parkinson's disease. *CNS Neurosci. Ther.* **2020**, *26*, 518–526. [\[CrossRef\]](https://doi.org/10.1111/cns.13282) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31828965)
- 224. Brodacki, B.; Staszewski, J.; Toczyłowska, B.; Kozłowska, E.; Drela, N.; Chalimoniuk, M.; Stepien, A. Serum interleukin (IL-2, IL-10, IL-6, IL-4), TNFalpha, and INFgamma concentrations are elevated in patients with atypical and idiopathic parkinsonism. *Neurosci. Lett.* **2008**, *441*, 158–162. [\[CrossRef\]](https://doi.org/10.1016/j.neulet.2008.06.040) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/18582534)
- 225. Thundyil, J.; Lim, K.L. DAMPs and neurodegeneration. *Ageing Res. Rev.* **2015**, *24*, 17–28. [\[CrossRef\]](https://doi.org/10.1016/j.arr.2014.11.003)
- 226. Moehlman, A.T.; Kanfer, G.; Youle, R.J. Loss of STING in parkin mutant flies suppresses muscle defects and mitochondria damage. *PLoS Genet.* **2023**, *19*, e1010828. [\[CrossRef\]](https://doi.org/10.1371/journal.pgen.1010828)
- 227. Ji, Y.; Li, M.; Chang, M.; Liu, R.; Qiu, J.; Wang, K.; Deng, C.; Shen, Y.; Zhu, J.; Wang, W.; et al. Inflammation: Roles in Skeletal Muscle Atrophy. *Antioxidants* **2022**, *11*, 1686. [\[CrossRef\]](https://doi.org/10.3390/antiox11091686)
- 228. Liu, J.; Wang, L.N. Mitochondrial enhancement for neurodegenerative movement disorders: A systematic review of trials involving creatine, coenzyme Q10, idebenone and mitoquinone. *CNS Drugs* **2014**, *28*, 63–68. [\[CrossRef\]](https://doi.org/10.1007/s40263-013-0124-4) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24242074)
- 229. Yang, L.; Calingasan, N.Y.; Wille, E.J.; Cormier, K.; Smith, K.; Ferrante, R.J.; Beal, M.F. Combination therapy with coenzyme Q10 and creatine produces additive neuroprotective effects in models of Parkinson's and Huntington's diseases. *J. Neurochem.* **2009**, *109*, 1427–1439. [\[CrossRef\]](https://doi.org/10.1111/j.1471-4159.2009.06074.x)
- 230. Garrido-Maraver, J.; Cordero, M.D.; Oropesa-Avila, M.; Vega, A.F.; de la Mata, M.; Pavon, A.D.; Alcocer-Gomez, E.; Calero, C.P.; Paz, M.V.; Alanis, M.; et al. Clinical applications of coenzyme Q10. *Front. Biosci.* **2014**, *19*, 619–633. [\[CrossRef\]](https://doi.org/10.2741/4231)
- 231. Arenas-Jal, M.; Suñé-Negre, J.M.; García-Montoya, E. Coenzyme Q10 supplementation: Efficacy, safety, and formulation challenges. *Compr. Rev. Food Sci. Food Saf.* **2020**, *19*, 574–594. [\[CrossRef\]](https://doi.org/10.1111/1541-4337.12539)
- 232. Delgobo, M.; Agnes, J.P.; Gonçalves, R.M.; Dos Santos, V.W.; Parisotto, E.B.; Zamoner, A.; Zanotto-Filho, A. N-acetylcysteine and alpha-lipoic acid improve antioxidant defenses and decrease oxidative stress, inflammation and serum lipid levels in ovariectomized rats via estrogen-independent mechanisms. *J. Nutr. Biochem.* **2019**, *67*, 190–200. [\[CrossRef\]](https://doi.org/10.1016/j.jnutbio.2019.02.012)
- 233. Ciulla, M.; Marinelli, L.; Cacciatore, I.; Stefano, A.D. Role of Dietary Supplements in the Management of Parkinson's Disease. *Biomolecules* **2019**, *9*, 271. [\[CrossRef\]](https://doi.org/10.3390/biom9070271) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31295842)
- 234. Alrouji, M.; Al-Kuraishy, H.M.; Al-Gareeb, A.I.; Saad, H.M.; Batiha, G.E. A story of the potential effect of non-steroidal antiinflammatory drugs (NSAIDs) in Parkinson's disease: Beneficial or detrimental effects. *Inflammopharmacology* **2023**, *31*, 673–688. [\[CrossRef\]](https://doi.org/10.1007/s10787-023-01192-2)
- 235. Klegeris, A.; McGeer, P.L. Non-steroidal anti-inflammatory drugs (NSAIDs) and other anti-inflammatory agents in the treatment of neurodegenerative disease. *Curr. Alzheimer Res.* **2005**, *2*, 355–365. [\[CrossRef\]](https://doi.org/10.2174/1567205054367883)
- 236. Fouré, A.; Bendahan, D. Is Branched-Chain Amino Acids Supplementation an Efficient Nutritional Strategy to Alleviate Skeletal Muscle Damage? A Systematic Review. *Nutrients* **2017**, *9*, 1047. [\[CrossRef\]](https://doi.org/10.3390/nu9101047)
- 237. Neinast, M.; Murashige, D.; Arany, Z. Branched Chain Amino Acids. *Annu. Rev. Physiol.* **2019**, *81*, 139–164. [\[CrossRef\]](https://doi.org/10.1146/annurev-physiol-020518-114455)
- 238. Shimomura, Y.; Yamamoto, Y.; Bajotto, G.; Sato, J.; Murakami, T.; Shimomura, N.; Kobayashi, H.; Mawatari, K. Nutraceutical effects of branched-chain amino acids on skeletal muscle. *J. Nutr.* **2006**, *136*, 529s–532s. [\[CrossRef\]](https://doi.org/10.1093/jn/136.2.529S) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/16424141)
- 239. Wolfe, R.R. Branched-chain amino acids and muscle protein synthesis in humans: Myth or reality? *J. Int. Soc. Sports Nutr.* **2017**, *14*, 30. [\[CrossRef\]](https://doi.org/10.1186/s12970-017-0184-9)
- 240. Son, S.M.; Park, S.J.; Stamatakou, E.; Vicinanza, M.; Menzies, F.M.; Rubinsztein, D.C. Leucine regulates autophagy via acetylation of the mTORC1 component raptor. *Nat. Commun.* **2020**, *11*, 3148. [\[CrossRef\]](https://doi.org/10.1038/s41467-020-16886-2) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32561715)
- 241. Yan, Z.; Yang, F.; Sun, L.; Yu, J.; Si, Y.; Yao, L. Role of gut microbiota-derived branched-chain amino acids in the pathogenesis of Parkinson's disease: An animal study. *Brain Behav. Immun.* **2022**, *106*, 307–321. [\[CrossRef\]](https://doi.org/10.1016/j.bbi.2022.09.009)
- 242. Tosukhowong, P.; Boonla, C.; Dissayabutra, T.; Kaewwilai, L.; Muensri, S.; Chotipanich, C.; Joutsa, J.; Rinne, J.; Bhidayasiri, R. Biochemical and clinical effects of Whey protein supplementation in Parkinson's disease: A pilot study. *J. Neurol. Sci.* **2016**, *367*, 162–170. [\[CrossRef\]](https://doi.org/10.1016/j.jns.2016.05.056)
- 243. Matthews, V.B.; Aström, M.B.; Chan, M.H.; Bruce, C.R.; Krabbe, K.S.; Prelovsek, O.; Akerström, T.; Yfanti, C.; Broholm, C.; Mortensen, O.H.; et al. Brain-derived neurotrophic factor is produced by skeletal muscle cells in response to contraction and enhances fat oxidation via activation of AMP-activated protein kinase. *Diabetologia* **2009**, *52*, 1409–1418. [\[CrossRef\]](https://doi.org/10.1007/s00125-009-1364-1)
- 244. Krumpolec, P.; Vallova, S.; Slobodova, L.; Tirpakova, V.; Vajda, M.; Schon, M.; Klepochova, R.; Janakova, Z.; Straka, I.; Sutovsky, S.; et al. Aerobic-Strength Exercise Improves Metabolism and Clinical State in Parkinson's Disease Patients. *Front. Neurol.* **2017**, *8*, 698. [\[CrossRef\]](https://doi.org/10.3389/fneur.2017.00698) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29312123)
- 245. Schootemeijer, S.; van der Kolk, N.M.; Bloem, B.R.; de Vries, N.M. Current Perspectives on Aerobic Exercise in People with Parkinson's Disease. *Neurotherapeutics* **2020**, *17*, 1418–1433. [\[CrossRef\]](https://doi.org/10.1007/s13311-020-00904-8) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32808252)
- 246. Lamotte, G.; Rafferty, M.R.; Prodoehl, J.; Kohrt, W.M.; Comella, C.L.; Simuni, T.; Corcos, D.M. Effects of endurance exercise training on the motor and non-motor features of Parkinson's disease: A review. *J. Park. Dis.* **2015**, *5*, 21–41. [\[CrossRef\]](https://doi.org/10.3233/jpd-140425)
- 247. De Oliveira, M.P.B.; Lobato, D.F.M.; Smaili, S.M.; Carvalho, C.; Borges, J.B.C. Effect of aerobic exercise on functional capacity and quality of life in individuals with Parkinson's disease: A systematic review of randomized controlled trials. *Arch. Gerontol. Geriatr.* **2021**, *95*, 104422. [\[CrossRef\]](https://doi.org/10.1016/j.archger.2021.104422) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33932826)
- 248. Supriya, R.; Singh, K.P.; Gao, Y.; Gu, Y.; Baker, J.S. Effect of Exercise on Secondary Sarcopenia: A Comprehensive Literature Review. *Biology* **2021**, *11*, 51. [\[CrossRef\]](https://doi.org/10.3390/biology11010051)
- 249. Silva-Batista, C.; Mattos, E.C.; Corcos, D.M.; Wilson, J.M.; Heckman, C.J.; Kanegusuku, H.; Piemonte, M.E.; Túlio de Mello, M.; Forjaz, C.; Roschel, H.; et al. Resistance training with instability is more effective than resistance training in improving spinal inhibitory mechanisms in Parkinson's disease. *J. Appl. Physiol.* **2017**, *122*, 1–10. [\[CrossRef\]](https://doi.org/10.1152/japplphysiol.00557.2016)
- 250. Silva-Batista, C.; Corcos, D.M.; Kanegusuku, H.; Piemonte, M.E.P.; Gobbi, L.T.B.; de Lima-Pardini, A.C.; de Mello, M.T.; Forjaz, C.L.M.; Ugrinowitsch, C. Balance and fear of falling in subjects with Parkinson's disease is improved after exercises with motor complexity. *Gait Posture* **2018**, *61*, 90–97. [\[CrossRef\]](https://doi.org/10.1016/j.gaitpost.2017.12.027)
- 251. Elangovan, N.; Cheung, C.; Mahnan, A.; Wyman, J.F.; Tuite, P.; Konczak, J. Hatha yoga training improves standing balance but not gait in Parkinson's disease. *Sports Med. Health Sci.* **2020**, *2*, 80–88. [\[CrossRef\]](https://doi.org/10.1016/j.smhs.2020.05.005)
- 252. Feng, Y.S.; Yang, S.D.; Tan, Z.X.; Wang, M.M.; Xing, Y.; Dong, F.; Zhang, F. The benefits and mechanisms of exercise training for Parkinson's disease. *Life Sci.* **2020**, *245*, 117345. [\[CrossRef\]](https://doi.org/10.1016/j.lfs.2020.117345)
- 253. Hackney, M.E.; Wolf, S.L. Impact of Tai Chi Chu'an practice on balance and mobility in older adults: An integrative review of 20 years of research. *J. Geriatr. Phys. Ther.* **2014**, *37*, 127–135. [\[CrossRef\]](https://doi.org/10.1519/JPT.0b013e3182abe784) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24406709)
- 254. Jablonka, S.; Hennlein, L.; Sendtner, M. Therapy development for spinal muscular atrophy: Perspectives for muscular dystrophies and neurodegenerative disorders. *Neurol. Res. Pract.* **2022**, *4*, 2. [\[CrossRef\]](https://doi.org/10.1186/s42466-021-00162-9) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34983696)
- 255. Sacheck, J.M.; Hyatt, J.P.; Raffaello, A.; Jagoe, R.T.; Roy, R.R.; Edgerton, V.R.; Lecker, S.H.; Goldberg, A.L. Rapid disuse and denervation atrophy involve transcriptional changes similar to those of muscle wasting during systemic diseases. *FASEB J.* **2007**, *21*, 140–155. [\[CrossRef\]](https://doi.org/10.1096/fj.06-6604com) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/17116744)
- 256. Tiryaki, E.; Horak, H.A. ALS and other motor neuron diseases. *Continuum* **2014**, *20*, 1185–1207. [\[CrossRef\]](https://doi.org/10.1212/01.CON.0000455886.14298.a4)
- 257. Uitti, R.J.; Berry, K.; Yasuhara, O.; Eisen, A.; Feldman, H.; McGeer, P.L.; Calne, D.B. Neurodegenerative 'overlap' syndrome: Clinical and pathological features of Parkinson's disease, motor neuron disease, and Alzheimer's disease. *Park. Relat. Disord.* **1995**, *1*, 21–34. [\[CrossRef\]](https://doi.org/10.1016/1353-8020(95)00004-P)
- 258. Johri, A.; Beal, M.F. Mitochondrial dysfunction in neurodegenerative diseases. *J. Pharmacol. Exp. Ther.* **2012**, *342*, 619–630. [\[CrossRef\]](https://doi.org/10.1124/jpet.112.192138) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/22700435)
- 259. Moreira, P.I.; Duarte, A.I.; Santos, M.S.; Rego, A.C.; Oliveira, C.R. An integrative view of the role of oxidative stress, mitochondria and insulin in Alzheimer's disease. *J. Alzheimers Dis.* **2009**, *16*, 741–761. [\[CrossRef\]](https://doi.org/10.3233/JAD-2009-0972)
- 260. Schapira, A.H. Mitochondrial dysfunction in Parkinson's disease. *Cell Death Differ.* **2007**, *14*, 1261–1266. [\[CrossRef\]](https://doi.org/10.1038/sj.cdd.4402160)
- 261. Irvine, G.B.; El-Agnaf, O.M.; Shankar, G.M.; Walsh, D.M. Protein aggregation in the brain: The molecular basis for Alzheimer's and Parkinson's diseases. *Mol. Med.* **2008**, *14*, 451–464. [\[CrossRef\]](https://doi.org/10.2119/2007-00100.Irvine)
- 262. Abyadeh, M.; Gupta, V.; Paulo, J.A.; Mahmoudabad, A.G.; Shadfar, S.; Mirshahvaladi, S.; Nguyen, C.T.O.; Finkelstein, D.I.; You, Y.; Haynes, P.A.; et al. Amyloid-beta and tau protein beyond Alzheimer's disease. *Neural Regen. Res.* **2024**, *19*, 1262–1276. [\[CrossRef\]](https://doi.org/10.4103/1673-5374.386406)
- 263. Khan, M.S.H.; Hegde, V. Obesity and Diabetes Mediated Chronic Inflammation: A Potential Biomarker in Alzheimer's Disease. *J. Pers. Med.* **2020**, *10*, 42. [\[CrossRef\]](https://doi.org/10.3390/jpm10020042) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32455946)
- 264. Nagatsu, T.; Sawada, M. Inflammatory process in Parkinson's disease: Role for cytokines. *Curr. Pharm. Des.* **2005**, *11*, 999–1016. [\[CrossRef\]](https://doi.org/10.2174/1381612053381620) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/15777250)
- 265. Phani, S.; Loike, J.D.; Przedborski, S. Neurodegeneration and inflammation in Parkinson's disease. *Park. Relat. Disord.* **2012**, *18* (Suppl. S1), S207–S209. [\[CrossRef\]](https://doi.org/10.1016/S1353-8020(11)70064-5) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/22166436)
- 266. Choe, M.A.; Koo, B.S.; An, G.J.; Jeon, S. Effects of Treadmill Exercise on the Recovery of Dopaminergic Neuron Loss and Muscle Atrophy in the 6-OHDA Lesioned Parkinson's Disease Rat Model. *Korean J. Physiol. Pharmacol.* **2012**, *16*, 305–312. [\[CrossRef\]](https://doi.org/10.4196/kjpp.2012.16.5.305) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/23129977)
- 267. Buchman, A.S.; Bennett, D.A. Loss of motor function in preclinical Alzheimer's disease. *Expert. Rev. Neurother.* **2011**, *11*, 665–676. [\[CrossRef\]](https://doi.org/10.1586/ern.11.57)
- 268. Martignon, C.; Ruzzante, F.; Giuriato, G.; Laginestra, F.G.; Pedrinolla, A.; Di Vico, I.A.; Saggin, P.; Stefanelli, D.; Tinazzi, M.; Schena, F.; et al. The key role of physical activity against the neuromuscular deterioration in patients with Parkinson's disease. *Acta Physiol.* **2021**, *231*, e13630. [\[CrossRef\]](https://doi.org/10.1111/apha.13630)
- 269. Bettio, L.E.B.; Rajendran, L.; Gil-Mohapel, J. The effects of aging in the hippocampus and cognitive decline. *Neurosci. Biobehav. Rev.* **2017**, *79*, 66–86. [\[CrossRef\]](https://doi.org/10.1016/j.neubiorev.2017.04.030)
- 270. Archer, T. Physical exercise alleviates debilities of normal aging and Alzheimer's disease. *Acta Neurol. Scand.* **2011**, *123*, 221–238. [\[CrossRef\]](https://doi.org/10.1111/j.1600-0404.2010.01412.x)
- 271. Tsitkanou, S.; Della Gatta, P.; Foletta, V.; Russell, A. The Role of Exercise as a Non-pharmacological Therapeutic Approach for Amyotrophic Lateral Sclerosis: Beneficial or Detrimental? *Front. Neurol.* **2019**, *10*, 783. [\[CrossRef\]](https://doi.org/10.3389/fneur.2019.00783)
- 272. Lautenschlager, N.T.; Cox, K.L.; Ellis, K.A. Physical activity for cognitive health: What advice can we give to older adults with subjective cognitive decline and mild cognitive impairment? *Dialogues Clin. Neurosci.* **2019**, *21*, 61–68. [\[CrossRef\]](https://doi.org/10.31887/DCNS.2019.21.1/nlautenschlager)
- 273. Pham, J.; Keon, M.; Brennan, S.; Saksena, N. Connecting RNA-Modifying Similarities of TDP-43, FUS, and SOD1 with MicroRNA Dysregulation Amidst A Renewed Network Perspective of Amyotrophic Lateral Sclerosis Proteinopathy. *Int. J. Mol. Sci.* **2020**, *21*, 3464. [\[CrossRef\]](https://doi.org/10.3390/ijms21103464) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32422969)
- 274. Guo, Y.; Wang, Q.; Chen, S.; Xu, C. Functions of amyloid precursor protein in metabolic diseases. *Metabolism* **2021**, *115*, 154454. [\[CrossRef\]](https://doi.org/10.1016/j.metabol.2020.154454) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33248065)
- 275. Mizushima, N.; Levine, B.; Cuervo, A.M.; Klionsky, D.J. Autophagy fights disease through cellular self-digestion. *Nature* **2008**, *451*, 1069–1075. [\[CrossRef\]](https://doi.org/10.1038/nature06639) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/18305538)
- 276. Kwon, Y.T.; Ciechanover, A. The Ubiquitin Code in the Ubiquitin-Proteasome System and Autophagy. *Trends Biochem. Sci.* **2017**, *42*, 873–886. [\[CrossRef\]](https://doi.org/10.1016/j.tibs.2017.09.002)
- 277. Hong, L.; Huang, H.C.; Jiang, Z.F. Relationship between amyloid-beta and the ubiquitin-proteasome system in Alzheimer's disease. *Neurol. Res.* **2014**, *36*, 276–282. [\[CrossRef\]](https://doi.org/10.1179/1743132813Y.0000000288)
- 278. Wong, E.; Cuervo, A.M. Autophagy gone awry in neurodegenerative diseases. *Nat. Neurosci.* **2010**, *13*, 805–811. [\[CrossRef\]](https://doi.org/10.1038/nn.2575)
- 279. Ciechanover, A.; Kwon, Y.T. Degradation of misfolded proteins in neurodegenerative diseases: Therapeutic targets and strategies. *Exp. Mol. Med.* **2015**, *47*, e147. [\[CrossRef\]](https://doi.org/10.1038/emm.2014.117)
- 280. Xilouri, M.; Stefanis, L. Autophagy in the central nervous system: Implications for neurodegenerative disorders. *CNS Neurol. Disord. Drug Targets* **2010**, *9*, 701–719. [\[CrossRef\]](https://doi.org/10.2174/187152710793237421)
- 281. Tanik, S.A.; Schultheiss, C.E.; Volpicelli-Daley, L.A.; Brunden, K.R.; Lee, V.M. Lewy body-like α-synuclein aggregates resist degradation and impair macroautophagy. *J. Biol. Chem.* **2013**, *288*, 15194–15210. [\[CrossRef\]](https://doi.org/10.1074/jbc.M113.457408)
- 282. Menzies, F.M.; Fleming, A.; Rubinsztein, D.C. Compromised autophagy and neurodegenerative diseases. *Nat. Rev. Neurosci.* **2015**, *16*, 345–357. [\[CrossRef\]](https://doi.org/10.1038/nrn3961)

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.