Contents lists available at ScienceDirect

# Brain Research Bulletin

journal homepage: www.elsevier.com/locate/brainresbull

# Gait analysis in chemotherapy-induced peripheral neurotoxicity rodent models

Maria Lopez-Garzon<sup>a, b, c, d</sup>, Annalisa Canta<sup>e, f</sup>, Alessia Chiorazzi<sup>e, f</sup>, Paola Alberti<sup>e, f, g, \*</sup>

<sup>a</sup> Biomedical Group (BIO277), Department of Physiotherapy, Faculty of Health Sciences, University of Granada, Granada, Spain

<sup>b</sup> A02-Cuídate, Instituto de Investigación Biosanitaria Ibs, GRANADA, Granada, Spain

<sup>c</sup> Unit of Excellence On Exercise and Health (UCEES), University of Granada, Granada, Spain

<sup>d</sup> Sport and Health Research Center (IMUDs), Granada, Spain

<sup>e</sup> Experimental Neurology Unit, School of Medicine and Surgery, Monza, Italy

<sup>f</sup> NeuroMI (Milan Center for neuroscience), Milan, Italy

<sup>g</sup> Fondazione IRCCS San Gerardo dei Tintori, Monza, Italy

### ARTICLE INFO

Keywords: Chemotherapy-induced peripheral neuropathy Chemotherapy-induced peripheral neurotoxicity Gait analysis Animal models Sensory ataxia Physical therapy Cat Walk<sup>TM</sup> Neuropathy

# ABSTRACT

Gait analysis could be used in animal models as an indicator of sensory ataxia due to chemotherapy-induced peripheral neurotoxicity (CIPN). Over the years, gait analysis in in vivo studies has evolved from simple observations carried out by a trained operator to computerised systems with machine learning that allow the quantification of any variable of interest and the establishment of algorithms for behavioural classification. However, there is not a consensus on gait analysis use in CIPN animal models; therefore, we carried out a systematic review. Of 987 potentially relevant studies, 14 were included, in which different methods were analysed (observation, footprint and CatWalk<sup>™</sup>). We presented the *state-of-the-art* of possible approaches to analyse sensory ataxia in rodent models, addressing advantages and disadvantages of different methods available. Semi-automated methods may be of interest when preventive or therapeutic strategies are evaluated, also considering their methodological simplicity and automaticity; up to now, only CatWalk<sup>™</sup> analysis has been tested. Future studies should expect that CIPN-affected animals tend to reduce hind paw support due to pain, allodynia or loss of sensation, and an increase in swing phase could or should be observed. Few available studies documented these impairments at the last time point, and only appeared later on respect to other earlier signs of CIPN (such as altered neurophysiological findings). For that reason, gait impairment could be interpreted as late repercussions of loss of sensory.

# 1. Introduction

Chemotherapy-induced peripheral neurotoxicity (CIPN) is a common late toxicity of the most commonly used anticancer drugs: platinum-drugs, taxanes, vinca alkaloids, epothilones, proteasome inhibitors and thalidomide (Alberti et al., 2022). CIPN can be long-lasting or even permanent altering the quality of life of cancer survivors (Alberti et al., 2022; Briani et al., 2014). CIPN key features are related mostly to sensory disturbances affecting peripheral nerves: positive and negative signs/symptoms are present. Positive symptoms consist of abnormal sensations such as paraesthesia/dysesthesia and neuropathic pain, whereas sensory loss for the different modalities equals to the so-called negative signs/symptoms. Motor and autonomic changes are possible but usually quite limited (Alberti et al., 2022), with some variations depending on the drug administered. If large fiber sensory modality is impaired, changes in gait and balance are expected, and they are not usually related to motor alterations which can be very mild or even not present (Alberti et al., 2022; Cavaletti et al., 2019). If loss of proprioception is relevant, in fact, a condition known as sensory ataxia is developed (Cavaletti et al., 2019). Sensory ataxia due to CIPN is associated with increased patients' fear of falling (Zahiri et al., 2019) and actual increased risk of falls (Kolb et al., 2016), and with a deterioration in quality of life after chemotherapy treatment completion (Mols et al., 2014). Gait analysis could be used as an indicator of CIPN-related sensory ataxia (Chen et al., 2021), being a tool that can be implemented in in vivo studies (Bruna et al., 2020) that are warranted to pave the way to novel CIPN treatments (Cavaletti et al., 2008); in fact, so far CIPN treatment still lacks robust evidence both for pharmacological (Loprinzi

\* Correspondence to: School of Medicine and Surgery, University of Milano-Bicocca, U8 Building, 1st floor, Room 1034, Via Cadore, 48, 20900 Monza, Italy. *E-mail address:* paola.alberti@unimib.it (P. Alberti).

https://doi.org/10.1016/j.brainresbull.2023.110769

Received 1 August 2023; Received in revised form 5 September 2023; Accepted 22 September 2023 Available online 24 September 2023

0361-9230/© 2023 The Author(s). Published by Elsevier Inc. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).



Review



et al., 2020) and non-pharmacological (Tamburin et al., 2022) treatment strategies (e.g., physical therapy), and, thus, preclinical studies searching for a sound biological rationale are still warranted: translational outcome measures are a key requirement to promptly transfer data from bench to bedside. Gait analysis may be useful in preclinical studies aiming at identifying compounds and/or non-pharmacological strategies to modify sensory ataxia due to CIPN (Cavaletti et al., 2008).

# 2. Gait analysis: an overview of possible approaches

Over the years, gait analysis in in vivo studies has evolved from simple observations carried out by a trained operator to computerised systems with machine learning that allow the quantification of any variable of interest and the establishment of algorithms for behavioural classification (Abbas and Masip Rodo, 2019) (Table 1).

A major breakthrough was the introduction of the Sciatic Functional Index (SFI), especially for the assessment of recovery improvement in the sciatic injury model (Inserra et al., 1998). This index varies its score according to the severity of damage, from 0 for nearly normal function to -100 for severe injury. It has been used mainly in unilateral peripheral nerve injuries, as the healthy contralateral side of the animal is required to apply the formula proposed by Inserra et al., 1998 (Inserra et al., 1998). However, SFI major limitation is related to the scarce number of variables that collects: paw lengths, and the paw widths, between the 1st and 5th digits and the 2nd to 4th digits. This index can be calculated in any gait analysis method, although initially, the animals' paws were dyed with a non-toxic dye and then allowed to walk on a blank sheet of paper. Technical issues might arise in analysing walking tracks, due to the development of flexion contractures, auto-mutilation, smearing of the print, dragging of the tail, or contamination with footprints.

Therefore, to obtain other gait parameters, more robust technologies were needed. Gait analysis recordings while animals walk on a transparent walkway or treadmill and subsequent analysis on the concomitant recording were then widely applied (Heinzel et al., 2020a). Two different approaches were mostly used: in one case the animal walks freely through a narrow walkway (the so called active walk, such as the CatWalk<sup>™</sup> device) towards a nest-like station, whereas in the other approach, the animal is filmed while moving over a treadmill at a pre-set speed (the so called *passive walk*, such as the DigiGait<sup>™</sup> device) (Xu et al., 2019). It is known that gait parameters could be differentially affected by the walking speed (Xu et al., 2019). Therefore, in the active method, in which the animal walks freely, the running speed should always be monitored and even considered in the analysis. Methods in which this speed is pre-selected may be less biased in this regard. However, it is possible that during the trial, animals (or a group of animals) may not be able to run at the same pre-selected speed as a consequence of neuropathy; therefore, this aspect should be carefully weighted too. The advantages of CatWalk<sup>™</sup> include additional intensity data and corresponding 3D images, respect to DigiGait<sup>TM</sup>, due to the Illuminated Footprints<sup>™</sup> technique (Xu et al., 2019). CatWalk<sup>™</sup> allows, in fact, transferring paw's pressure into fluorescence signals.

Another option in rodents is to analyse gait while they are obliged to keep walking for 2 min in the acrylic wheel (GAIT<sup>TM</sup> system) at a predetermined revolutions per minute (rpm) (Matsuda et al., 2016). Authors claim this allows animals to walk more naturally, although the hind paw would bear the dynamic weight, and could only be studied in asymmetric and hind paw models.

In addition to measuring gait in rodents, it could be of interest analysing composite functions, to evaluate distinct aspects of motor function and to determine even subtle loss of movement capacity such as walking on a ledge (Gyengesi et al., 2019) or ladder (Metz and Whishaw, 2009), eventually applying and adapting tests that are known to be used not for sensory but central ataxia and central nervous system disturbances. The Ledge test is a fair example of this; this test is mainly used in cerebellar ataxia and other neurodegenerative disease models. The examiner scores the ability of the animal to walk on the edge, using its tail as a counterbalance and balance to descend gracefully back to the cage or table using its paws. This test is scored on a scale of 0–3, with 0 representing the normal movement and 3 representing the most severe expression of ataxia. This scoring is subjective and based on the analysis of foot slips, rigidity or efficiency of the tail, or the inability to perform the test. Impaired motor function, in ataxic rodents could also be measured by the hindlimb clasping test (Chou et al., 2008; Guyenet et al., 2010; Gyengesi et al., 2019) which is performed to assess whether the mouse clasps its hindlimbs into its body or sprays its limbs when suspended by its tail. However, it should be noted that in mice the hindlimb clasping test can show a flexural response that is characteristic of a central neurological disorder (Guyenet et al., 2010).

It is possible that the movement or position of different joints (kinematics) may vary in these murine models. One form of measurement involves analysis of foot (Varejão et al., 2003) or ankle (Lee et al., 2013) placement while walking, although this may be due to contracture formation. Another analysis could be the observation of dorsal kyphosis (Guyenet et al., 2010; Gyengesi et al., 2019; Thomas et al., 2006), which could be a test to monitor ataxia and muscle strength. The experimenter positioned the mouse on a flat surface and observed the spine while the animal walked. Kyphosis could be an indicator of ataxia (Guyenet et al., 2010).

Another aspect to be considered is the robustness of CIPN animal models when devising preclinical studies to test gait analysis in rodent models. As in patients, preclinical studies assess CIPN using various variables: histopathology, neurophysiology, behavioural test (Bruna et al., 2020; Pozzi et al., 2020). It should be carefully checked if the selected model is fully reproducing CIPN features. On another note, a careful planning of experiments should be verified: gait analysis can be conditioned when nerve conduction studies are performed inserting subdermal stainless steel electrodes in paws, potentially altering animals' gait (Boehmerle et al., 2014; Monza et al., 2021a; b); therefore, nerve conduction studies are to be performed after gait analysis assessment and not the reverse. In addition, in case a relevant disease severity is induced, animals might be hesitant in moving across the walkway (Liu et al., 2018); a possible solution proposed to counterbalance this was to introduce a conditioning phase with sugar pellets (Santos, 2000). Rats were conditioned to walk through the central tunnel into two clear boxes at either end of the tunnel. Each end box has a sugar pellet reward window. The animals are placed on a food deprivation schedule before training to ensure they are receptive to a food reward. This method is repeated daily until the rat is conditioned to walk from end to end for a sugar pellet reward (Santos, 2000).

# 3. Gait analysis in CIPN models: literature review

For this review, only studies published up to January 24th, 2023, were considered. No restrictions were placed on year, but publications were limited by English, Spanish or Italian language. Based on the Population, Intervention, Comparator, Outcomes, and Study Designs strategy studies where gait was measured in rodents' model of CIPN were included (https://www.york.ac.uk/crd/SysRev/!SSL!/WebHelp/SysRev3.htm, accessed on 20th January 2023).

A literature search was conducted using relevant subject headings, keywords, and modifications made according to the three databases searched: Medline (Table 2), Web of Science and Scopus; modifications were made to fit each database. All articles were retrieved and exported to Rayyan (Ouzzani et al., 2016), where duplicates were removed and studies were identified and selected according to the inclusion criteria. All articles identified in the first screening process were included in the following one, in which selected articles were thoroughly read and screened for the inclusion criteria. Articles considered eligible after full-text view were included in the final analysis. Reasons for exclusion were recorded.

The following data were extracted from each article (1) general study

Table 1

ω

Common methods to analyse gait impairment in peripheral neuropathies other than CIPN in rodent models. PNI: Peripheral nerves injuries; SFI: Sciatic Functional Index; \*SFI: can be measured in any of the above methods.

Setting	Target of	Technique	Tool	Parameters Outcom		rs Pros Cons	
	evaluation						
Active walks	Footprints	Illuminated	CatWalk™ (Heinzel et al., 2020a) Semi-Automated methods	<ul> <li>-Print Length (distance unit): Length of the paw print</li> <li>-Print Width (distance unit): Width of the paw print</li> <li>-Print Area (distance unit): Area of the paw print</li> <li>-Base of Support (BoS) (distance unit):</li> <li>Distance between the two hind-or front paws</li> <li>-Mean Paw Print Intensity (arbitrary units): Mean intensity of the paw</li> <li>-Swing Phase (seconds): Swing Time of the paw</li> <li>-Stance Phase (seconds): Stand Time of the paw</li> <li>-Duty Cycle (%): Stand Time divided by Stand Time plus Swing Time</li> <li>-Swing Speed (centimetres / second): Swing Speed of the paw</li> <li>-Regularity Index (RI) (%): Quantification of interlimb coordination</li> <li>-Run Duration (seconds) Duration of the walkway croseing</li> </ul>	General parameters of gait Pain-related Coordination-related Other	Possibility to simultaneously monitor motor and sensory reinnervation in various models of PNI with only one device and setup. The illumination of only those areas which are interesting for the investigator and the correlation of paw pressure and footprint intensity. Compared to DigiGait <sup>™</sup> , CatWalk <sup>™</sup> excels at intrinsic velocity, intensity data (acquired by Illuminated Footprints <sup>™</sup> ), and high-quality 3D images.	Requires meticulous training efforts to accustom the animals to the device. CatWalk™ is limited to animals of the size of rodents and ferrets Limited sensitivity to detect functional recovery after more severe types of nerve injury. It does not function in the situation that rats hardly walk. The animal could decrease velocity and compensate for gait changes that would be different at a higher speed (Piesla et al., 2009).
		Video techniques	Rat's head movement (Santos,	-Gait time (milliseconds)	Gait speeds	If changes in gait velocity occurred, variability	Need to compare both paws, useful in
		techniques	Gait-stance duration (Walker et al., 1994)	-Distances between the front and hind paws, diagonal distances between front and hind paws and height of the iliac crest -Ratios of the basic distances of the injured versus uninjured side including "injured left hind paw to the right front paw" versus "right hind paw to left front paw" (LH-RF/ RH-LF) -The time of floor contact is defined as gait- stance duration. The ratio of the injured/ uninjured hind feet stance duration was	Locomotor ability Coordination Functional recovery	duration of each step with the average gait-stance duration from the 2 steps of the alternate foot immediately preceding and following it. Only a video camera and frame-by-frame playback recorders are required. In animals sustained partial toe loss, the gait- stance duration may be more reliable than any method using toe spreads (such as SFI).	Dependent operator. The cost is greater than ink and paper walking tracks.
		Ink techniqı	technique (Araújo-Filho et al., 2020)	calculated from paired consecutive steps when measured during steady walking -Print length (PL) -Distance between the 1st and 5th toes (TS) -Distance between the 2nd to 4th toes (IT)	Sciatic Functional Index (SFI) (Kim et al., 2007)*	Widely used metric for the pathology and potential treatment of nerve injury. The SFI is determined by comparing the geometric representation of the affected hind paw from an injured mouse or rat, and comparing it to the contralateral paw	Need to compare both paws, useful in asymmetric (not representative of CIPN). The walking track analysis costs are relatively high, and results are frequently unreproducible because of the smearing of the footprints
	Kinematics		Ankle Angle Measurements (Lee et al., 2013)	-The print lengths of both the affected side (EPL) and the untouched side (NPL) PLF= (EPL-NPL)/NPL -% Functional Recovery: (%FR)= (1-PLF) x 100 -Ankle angle (degrees): measured between the leg segment and the foot segment. -Ankle contracture angle	Print Length Factor (PLF) (Ozkan et al., 2005) Functional Recovery Gait kinematic information	Evaluate functional recovery. Ankle contracture to be a useful measure of recovery (the ankle angle in toe-off phase measured during video gait analysis correlates with isometric tetanic force and as such is a useful	The darkened walking alley makes it impossible to evaluate qualitative walking patterns the calculation of SFI is not easy and is time consuming. Walking speed is not identified. Need to compare both paws, useful in asymmetric (not representative of CIPN).

parameter for evaluating functional recovery).

# Table 1 (continued)

4

Setting	etting Target of Techni evaluation		Tool	Parameters	Outcomes	Pros	Cons	
			Toe out angle (TOA) (Varejão et al., 2003)	-TOA (the angle in degrees between the direction of progression and a reference line on the sole of the foot	Abnormal foot rotation (functional recovery after sciatic nerve injury)	Excellent correlation between SFI and TOA. Assess the foot motion in the transverse plane of walking and its underlying biomechanical consequences.		
			Kyphosis (Castillo-Mariqueo and Giménez-Llort, 2022;Guyenet et al., 2010;Gyengesi et al., 2019;Thomas et al., 2006)	-Score 0: mouse walks with a straight spine. -Score 1: a mild kyphosis but the mouse is still able to straighten its back. -Score 2: the mouse is unable to fully straighten its spine and maintained persistent mild kyphosis during locomotion. -Score 3: the mouse maintains a constant pronounced kyphosis	Loss of muscle tone or strength in the spinal muscles.	Kyphosis is a characteristic dorsal curvature of the spine that is a common manifestation of neurodegenerative disease in mouse models. A measurement of ataxia.	Kyphosis is sensitive to an age effect.	
	Tasks		Ladder rung walking tests (LRWT) (Fey et al., 2010)	-Average time needed to cross the entire length of the ladder. -Digit score in each correct placement (full flexion, flexion 45°, flexion 90°) -The spread length (in millimeters) -Ladder spread index (ratio of the spread of	Sensory-motor ability Tibial and peroneal nerve functions	The five categories of paw placement are affected differently depending on the severity of the root lesion. LRWT is a complex skilled motor tasks, which are more sensitive in detecting small impairments in limb use such as skilled paw placement. LRWT requires the integrity of all sensory-motor feedback systems, such as proprioceptive and mechanoreceptive systems.	It was also observed that forepaw placement of the contralateral (uninjured) limb was affected, though no formal assessment of this was done.	
			Ledge test (Guyenet et al., 2010; Gyengesi et al., 2019)	digits 2–4 measured in two successive video frames) -Ladder stance angle -Score 0: Mouse typically walks along the ledge without losing its balance, and lowers itself back into the cage gracefully, using its paws. -Score 1: mouse foot slips while walking along the ledge; the tail may present rigidity and could not be able to counterbalance well but the mouse is still able to walk and descend using its paws. -Score 2: the mouse is not effectively using its hind limbs or lands on its head rather than its paws when descending the cage. -Score 3: the mouse fells off the ledge, or nearly so, when attempting to descend	Tibial nerve (plantar flexion) function Balance and coordination	It is a direct measure of coordination. Sensitive and easily evaluation of disease severity. A measurement of ataxia (mostly central).	Mainly used in mouse models of cerebellar ataxia and other neurodegenerative disease, including Huntington's disease and spinobulbar muscular atrophy. Ledge test is sensitive to an age effect.	
			Hindlimb clasping test (Guyenet et al., 2010;Gyengesi et al., 2019)	back to the cage -Score 0: If the hindlimbs are splayed outward away from the abdomen consistently -Score 1: If one of the hindlimbs is retracted toward the abdomen for more than 50% of the observation time (10 s). -Score 2: If both of the hindlimbs are partially retracted toward the abdomen for more than 50%. -Score 3: If both hindlimbs are entirely retracted for more than 50% of the time.	Motor functions	It is a marker of disease progression in a number of mouse models of neurodegeneration, including certain cerebellar ataxias. May show flexural response characteristic of lesions in the cerebellum, basal ganglia, neocortex or spinal cord pathologies. A measurement of ataxia.		

M. Lopez-Garzon et al.

(continued on next page)

ed
п
Ϊ
5
9
-
le,

Table 1 (continu	(pai						
Setting	Target of T evaluation	l'echnique	Tool	Parameters	Outcomes	Pros	Cons
Passive walks	Treadmill		DigiGait <sup>TM</sup> (Umansky et al., 2022; Xu et al., 2019) Semi-Automated methods	-Swing phase (manually calculated) -Duty cycle (manually calculated) -Droiected area (recoonized from direct	Coordination Area data	Has advantages in fixed speed and dynamic SFI calculation. Possibility to use various software DiotGait <sup>IM</sup>	In models that warrant neither intensity data nor SFI (e.g., sciatic nerve injury), CatWalk <sup>TM</sup> is slichtly sumerior to DioiGair <sup>TM</sup> owinor to its
				recordings of the walking/running rats from the ventral direction (it was not		Visual Gait Lab (VGL) software.	higher-quality images to explicitly illustrate hindpaw abnormalities.
				merely the actual print of paw-floor contact)			Speed pre-selected is needed for gait analysis.
	Wheel		GAIT® (Matsuda et al., 2016) Semi-Automated methods	-Swing time ratio= (swing time of the normal hind limb) / (swing time of the	Abnormal step cycles (neuropathic pain)	Allow collect data of each step cycle from rats with serious injury, pain, and paralysis because of	It cannot differentiate types of neuronal f abnormalities.
				painful hind limb)		sub-spontaneous walking in the automatically	Need to compare both paws, useful in
				-Number of step cycles		rotated round wheel.	asymmetric (not representative of CIPN).
						Enables a natural walk based on the rodents'	Used only in models of hind paws.
						behavioural characteristic	
						GAIT® system might provide a more sensitive	
						parameter of dynamic weight bearing for	
						evaluating chronic neuropathic pain in rats in	

addition to conventional tests.

# Table 2 Search strategy in MEDLINE database.

PICOS Con	nponents of Search Strategy and filter applied
p	("Peripheral Nervous System Diseases/chemically induced"[Mesh] OR "Peripheral Nervous System Diseases"[Mesh terms] OR Peripheral Nervous System Diseases"[Mesh terms] OR Neuropath* Peripheral[tiab] OR Nerve Disease* Peripheral[tiab] OI Peripheral Nervous System Disorder*[tiab] OR "Small Fiber Neuropath* Peripheral[tiab] OR Nerve Disease* Peripheral[tiab] OI Peripheral Nervous System Disorder*[tiab] OR "Small Fiber Neuropaths"[Mesh] OR Small Fiber Neuropathy[tiab] OR Neuropath* Small Fiber[tiab] OR "Polyneuropathies"[Mesh] OR Polyneuropath*[tiab] OR Polyneuropath* Motor[tiab] OR "Neurotoxicity Syndromes"[Mesh] OR Neurotoxicity syndrome* [tiab] OR Neurotoxin Disorder*[tiab] OR Neurotoxic disorder*[tiab] OR Neurotoxin disease*[tiab] OR Chemotherapy induced peripheral neuropath*[tiab] OR CIPN[tiab] OR Chemotherapy Induced Polyneuropath*[tiab] OR Chemotherapy induced Neuropath [tiab] OR Platinum induced peripheral neurotoxicit*[tiab] OR Bortezomib induced peripheral neurotoxicit*[tiab] OR Gancer treatment induced neurotoxicit*[tiab] OR BIPN[tiab] OR Cancer treatment induced neurotoxicit*[tiab] OR Platinum drugs induced peripheral neuropath*[tiab] OR Chemotherapy induced painful peripheral neuropath*[tiab] OR Chemotherapy induced peripheral neuropath*[tiab] OR Chemotherapy induced peripheral neuropath*[tiab] OR Chemotherapy induced peripheral neuropath*[tiab] OR Detezomib Induced Neuropathic Pain[tiab] OR Chemotherapy induced neuropath*[tiab] OR platinum induced peripheral neuropath*[tiab] OR neuropath* [tiab] OR platinum induced peripheral neuropath*[tiab] OR neuropathy induced by bortezomib[tiab] OR Bortezomib Induced neuropathic Pain[tiab] OR Taxane induced neurotoxic* [tiab] OR bortezomib induced neurotoxic*[tiab] OR taxane induced neuropath*[tiab] OR Taxane induced peripheral neuropath*[tiab] OR bortezomib induced neurotoxic*[tiab] OR taxane induced neuropath*[tiab] OR Therapy related peripheral neuropath*[tiab] OR
I	[tiab] OR cancer neuropath*[tiab]) -
С	
0	("Gait Analysis"[Mesh] OR Analysis Gait[tiab] OR Gait Analyses[tiab OR "Gait/drug effects"[Mesh] OR "Gait Apraxia"[Mesh] OR Apraxia Gait[tiab] OR Apraxias Gait[tiab] OR Gait Apraxias[tiab] OR Dyspraxia of Gait[tiab] OR Gait Dyspraxia[tiab] OR Gait Dyspraxia [tiab] OR Apraxia of Gait[tiab] OR "Walking Speed"[Mesh] OR Speed Walking[tiab] OR Speeds Walking[tiab] OR Walking Speeds[tiab] OR Gait Speed[tiab] OR Gait Speeds[tiab] OR Speed Gait[tiab] OR Speeds Gait[tiab] OR Walking Pace[tiab] OR Pace Walking[tiab] OR Paces Walking[tiab] OR Walking Paces[tiab] OR "Lameness Animal"[Mesh] OR Animal Lameness[tiab] OR Animal Lamenesses [tiab] OR Lamenesses Animal[tiab] OR Gait Disorders Animal[tiab] OR Animal Gait Disorder[tiab] OR Disorders Animal Gait[tiab] OR Disorder Animal Gait[tiab] OR Disorders Animal Gait[tiab] OR Gait Disorder Animal[tiab]]
s	-
Filter	Species: Other Animals

applied

details: title, authors, year of publication and type of study; (2) study eligibility: model, including number of animals, age, sex, model-induced CIPN (cumulative dose, single dose and schedule), gait analysis outcomes and other outcomes to characterize the model, pros y cons of the gait analysis evaluation. The data extraction was documented in a Microsoft Excel spreadsheet.

The initial search found 987 eligible studies, 63 of which were removed during duplicate detection. Of the remaining 924 studies, 65 studies met the inclusion criteria. No paper was added from the reference list, or found with automatic alerts. A total of 14 papers were based on CIPN rodent models and were therefore assessed for the scope of this review (Fig. 1). Details of the literature search and paper selection are shown in Table 3. Considering all 12 experimental studies, a total of 140 rodents per chemotherapy group were included in the narrative synthesis (Table 3). Of the 14 paper included one was a systematic review (Heinzel et al., 2020a) and one was a methodological paper (Bruna et al., 2020), whose characteristics data are, thus, not reported in Table 3. The predominant sex tested was male (75%), and the average age was 7.68  $\pm$  4.65 weeks in the rat studies and 8.17  $\pm$  1.44 weeks in the mice studies. The most predominant rat strains were Dark Agouti and Sprague-Dawley (both 25%), followed by Wistar (16.66%). Four studies



Fig. 1. Flowchart according to the Preferred Reporting Items for Systematic Reviews and Meta-Analyses statement.

used C57BL/6 mice (33.33%) and only one CD-1 (8.33%) mice. With respect to drug-induced models, all of them were potentially neurotoxic (Velasco and Bruna, 2010). The most common model was vincristine (VCR)-induced peripheral neuropathy (6 studies, 50%), but cumulative doses varied greatly between studies, ranging from 200 µg/kg to 34 mg/kg. Five studies used cisplatin (CIS) ranging from 6 mg/kg up to 32 mg/kg and four of them use paclitaxel (PTX) with dosages ranging from 4 mg/kg up to 240 mg/kg. One study used compared carboplatin (CBDCA, 20 mg/kg) and bortezomib (BTZ, 4.8 mg/kg). Regarding the type of studies, despite the fact that all were experimental studies, only five of them randomised animals (Boehmerle et al., 2014; Huehnchen et al., 2013; Sahranavard et al., 2022; Shahid et al., 2017, 2019). In terms of the main scope, three of them aimed to characterise CIPN model with different outcomes (including gait analysis), while the others aimed at demonstrating efficacy neuroprotectant by medicating or applying the study drugs during the period of chemotherapy (secondary prevention) (Gewandter et al., 2018).

# 3.1. Types of gait analysis

# 3.1.1. Observation

Three studies carried out by the same group analysed gait via simple observation (Boyle et al., 2001; Boyle et al., 1996, 1999). Gait disturbance was assessed independently by two observers, who were blinded

to the assigned treatment. The first definitive change consisted of toe-walking with an arched hind paw, which was scored as a positive result. This progressed to a general paucity of motor activity and ultimately to severe hind limb weakness, which interfered with standing and grooming behaviour. This method could be classified as quick and easy but has some disadvantages. For example, it is operator-dependent and only classifies positive and negative results once CIPN occurs; therefore, although it could classify mild CIPN or severe CIPN, it is not sensible to early onset symptoms/signs.

The results with this method can be interpreted as the number of animals developing gait disturbances in each timepoint (highlighting the percentage per group). In the first study, on day 8, only 24% of the VCR-treated animals developed gait disturbances, but by day 15 all animals were observed to be toe-walking with an arched back (Boyle et al., 1996). Subsequently, the same group tested the same approach with different CIPN models; in the CIS model, first gait impairment was observed at week 8 (15% of animals), increasing to 32% at week 7% and 100% at week 8. Onset of toe-walking gait was observed in PTX-treated rats from week 2 (16%), and by week 3 affected 100% of rats receiving PTX (Boyle et al., 1999). Finally, in the last study, only PTX resulted in gait disturbances from day 9 (17%) and 33% on day 14. The other treatments (CIS or CBDCA) did not show any alterations in gait (Boyle et al., 2001). These inconsistent findings in the different proportion of animals developing toe-walking with an arched hind-paw between two

# Table 3

Characteristics of the 12 studies on gait analysis in chemotherapy-induced peripheral neurotoxicity-related sensory ataxia. #: BTZ: Affected forepaws more than hind paws; 1: decrease; 1: increase; Bold font means that the authors found a significant difference (time\*group).

Study	Model	Model induced	Instrument	Outcomes	Proc	Cons	Other outcomes
Study	Model	dose, schedule)	instrument	outcomes	PIOS	cons	Other outcomes
Patricia M. Whitaker-Azmitia et al. (1995) ( Whitaker-Azmitia et al., 1995)	CIPN (Sprague- Dawley pups rats)	Cisplatin 8 mg/ kg (1 mg/kg, twice weekly for 4 weeks)	Footprint patterns	Degree of toeing-in ↓ (by subtracting the interheel distance from the intertoe distance, 5 means per animals, and reported group mean)	It assesses a new marker, the convergence of the hind footprint, which could also indicate kinematics of the leg during gait.	The footprints can be erased by the mouse itself; it is necessary that the mouse is not missing any toes.	Tail-flick latency test (analgesic response) Immunocytochemistry (calcitonin gene-related peptide).
Frances M.Boyle et al. (1996) ( Boyle et al., 1996)	CIPN (Dark Agouti rats)	Vincristine 1.8 mg/kg (0.15 mg/kg, five consecutive days the first 2 weeks, and 2 consecutive days in the 3rd week)	Observation	Toe-walking with an arched back General paucity of motor activity Severe hind limb weakness, which interfered with standing and	Simple and fast. It classifies on a severity scale after CIPN onset.	Dependent operator It does not detect small variations, only + or - when CIPN exists.	Tail-flick test (thermal nociceptive thresholds) Rotarod performance (in light and dark environment).
Frances M.Boyle et al. (1999) ( Boyle et al., 1999)		Cisplatin 32 mg/kg (2 mg/ kg, twice weekly until 20% weight loss occurred) Paclitaxel 36 mg/kg (9 mg/ kg, twice weekly until 20% weight loss occurred)		grooming behaviour.			
Frances M.Boyle et al. (2001) ( Boyle et al., 2001)		Paclitaxel 30 mg/kg (5 mg/ kg, for 6 consecutive days) Cisplatin 6 mg/ kg (3 mg/kg, twice in a week) Carboplatin 20 mg/kg (10 mg/ kg, twice in a week)					
Patricia C Contreras et al. (1997) ( Contreras et al., 1997)	CIPN (CD-1 mice)	Vincristine 34 mg/kg (1.7 mg/ kg, twice weekly for 10 weeks)	Footprint patterns	Stride Length ↓ Gait support ↓ Toe spread Internal toe spread	No specific software is needed to analyse. It is also likely that changes in gait were not the result of vincristine-induced myopathy as the changes in caudal conduction velocity are indicative of a neuropathy and there was little consistent evidence of a myopathy histologically in the soleus and gastrocnemius muscles	It is more time- consuming to dye the animals' feet beforehand. The footprints can be erased by the mouse itself; it is necessary that the mouse is not missing any toes.	Hot-plate test (analgesic) Electrophysiological testing (caudal nerve conduction velocity) Histological analysis (percentage of degenerating fibers and total axon and myelin areas).
Petra Huehnchen et al. (2013) ( Huehnchen et al., 2013)	CIPN (C57BL/6 J mice)	Paclitaxel 240 mg/kg (20 mg/ kg, three per week for 4 weeks)	CatWalk™	Swing phase ↑ Stance phase ↓ Duty cycle ↓ The print area ↓ of the hind paws	Objective tool for evaluating sensory and motor neuropathy	Requires the inclusion of a non- neuropathic control group. Mice are much lighter than rats and the equipment therefore operates closer to its detection limit.	Rotarod test (motor coordination). Von Frey Hair test (mechanical allodynia). Electrophysiological testing (caudal nerve conduction velocity and sensory nerve action potential amplitudes).

Another contributing

Study	Model	Model-induced Cumulative dose (single dose, schedule)	Instrument	Outcomes	Pros	Cons	Other outcomes
						factor is the limited sensitivity of intensity measurements: Previous studies found that at least 40% weight variation is required for a statistically significant	
Wolfgang Boehmerle et al. (2014) ( Boehmerle et al., 2014)	CIPN (C57BL/6 J mice)	Vincristine 200 µg/kg (200 µg/kg, single dose) Paclitaxel 4 mg/kg (1 mg/ kg, 4 alternative days) Cisplatin 23 mg/kg (2.3 mg/ kg, five for 3 weeks x 2 cycles) Bortezomib 4.8 mg/kg (400 µg/kg, three per week for 4 weeks)	CatWalk™	Stance phase of the hindpaws ↓ (PTX, CIS, VIN, BTZ#) Duty cycle ↓ (PTX, CIS, VIN, BTZ#)	Gait analysis is not based on behaviour evoked by an artificial stimulus and is less influenced by the researcher, as well as by methodological simplicity.	result. Gait alterations appeared later than electrophysiological changes or mechanical allodynia.	Open fields (wellbeing of mice after injection). Rotarod test (motor coordination). Von Frey Hair test (mechanical allodynia). Electrophysiology (caudal nerve conduction velocity and sensory nerve action potential amplitudes). Histology (myelin and axon areas, changes in myelination were assessed with G-ratios which are determined by calculating the ratio of (ideal) axon diameter to (ideal) total fibre diameter)
Chang-Ning Liu et al. (2017) (Liu et al., 2018)	CIPN Wistar rats C57BL/6 J mice	for 4 weeks) Vincristine 0.75 mg/kg (75 μg/kg, once per day for 10 days) 1.1 mg/kg (100 μg/kg, once per day for 11 days)	CatWalk™	Swing phase Stance phase Stride length <b>Print area</b> ↑ (only mice both front and hind paws) Print intensity	As the time needed for an animal to cross the walkway is on the order of several seconds, relatively large groups of animals can be tested in a short time span. Because the analyses are based on captured images and stored by the system, the raw data can be retrieved and quality controlled retrospectively as	Footprints could be affected by weight. CatWalk™ gait analysis is more suitable for testing in mice than rats, which often showed a slower or more interrupted run on the walkway, although they were more cooperative in the pre- dose or early dosing phase.	First flick response of the tail (cold allodynia in mice). <i>Ex vivo</i> nerves mechanical testing (maximal load and the load/extension ratio). Histology (Dorsal root ganglia (DRG) and sciatic nerve).
Muhammad Shahida et al. (2017) (Shahid et al., 2017)	CIPN (Sprague- Dawley rats)	Cisplatin 12 mg/kg (3 mg/ kg, once per week for 4 weeks)	Footprint patterns	Stride length (average distance of forward movement between each stride) Overlap between forepaw and hind paw placement (distance between the front and hind footprints on each eide)	No specific software is needed to analyse	It is more time- consuming to dye the animals' feet beforehand. Only two trials were made per animal. It only reports two variables. It requires the inclusion of a control group.	Von Frey filaments (mechanical allodynia). Hot-plate test (thermal hypoalgesia). Rotarod test (motor performance).
Muhammad Shahid et al. (2019) ( Shahid et al., 2019)	CIPN (Sprague Dawley rats)	Cisplatin 12 mg/kg (3 mg/ kg, once per week for 4 weeks)	Footprint patterns	overlap (cm between forepaw and hind paw)			Von Frey filaments (mechanical allodynia). Hot-plate test (thermal hypoalgesia). Rotarod test (motor performance).
Crystal T.Bluette et al. (2021) ( Bluette et al., 2021)	CIPN (C57BL/6 J mice)	Vincristine Nano-VCR (NP- 684) 1.05 mg/ kg (0.15 mg/kg, per 7 alternatives days) Solution-based VCR 1.05 mg/ kg (0.15 mg/kg.	CatWalk™	Swing Stance ↑ (VCR in front) Cycle ↑ (VCR in front and hind paws) Stride length Print area Print intensity	Examined the 4 paws separately but concluded that the model is symmetrical.	It is possible that other non morphological factors also play a role in animal's pain behavioural manifestation, including gait behaviours.	Histology (microscopic lesions were graded as minimal, mild, moderate, marked, or severe in L4 and L5 DRG and sciatic nerve). miRNA quantification (miR-124, miR-338, and miR-183).

# Table 3 (continued)

(continued on next page)

Table 3 (continued)

Tuble 9 (continued)							
Study	Model	Model-induced Cumulative dose (single dose, schedule)	Instrument	Outcomes	Pros	Cons	Other outcomes
S.Sahranavard et al. (2022) ( Sahranavard et al., 2022)	CIPN (Wistar rats)	per 7 alternatives days) Vincristine 1.4 mg/kg (0.1 mg/ kg, daily for 2 weeks)	Footprint test (hind limb was dipped in ink and they were permitted to walk on a white paper placed on the surface of the track. Analysis with ImageJ)	Pressure of the foot ↓ by the measurement of pixel values (VCR)	Simple and fast	Operator dependent. It reports tinted area on paper as leg pressure which can lead to bias.	Open field test (general locomotor activity). Hot-plate test (thermal hyperalgesia). Von Frey hair test (mechanical allodynia). Grip strength (muscular strength).

studies (Boyle et al., 2001; Boyle et al., 1999), could be due to the different amount of CIS cumulative dose used (32 mg/kg vs 6 mg/kg).

# 3.1.2. Footprint

In total, five studies measured gait using footprint patterns by dyeing the animals' paws (Contreras et al., 1997; Sahranavard et al., 2022; Shahid et al., 2017, 2019; Whitaker-Azmitia et al., 1995). Generally for analysis, all (Shahid et al., 2017, 2019) or only the hind paws(Contreras et al., 1997; Sahranavard et al., 2022; Whitaker-Azmitia et al., 1995) were dyed and the animal was placed in a lighted corridor covered with white absorbent paper. Usually at least two footprint patterns were analysed. However, the variables studied differs greatly between the studies.

In 1995, Whitaker-Azmitia et al. (Whitaker-Azmitia et al., 1995) analysed the degree of gait abnormality as the degree of toeing-in. This parameter is determined by subtracting the inter-heel distance from the inter-toe distance: a reduction of the toe-heel distance (cm) is assigned as sign of peripheral neuropathy. In this study, they showed that animals treated with CIS had a significant (p < 0.02) decrease in the toe-heel measure respect to the control group ( $1.15 \pm 0.17$  cm. vs. 1.61  $\pm$  0.17 cm, respectively). However, this study used rat puppies and, therefore, gait abnormality could be due to central nervous system (CNS) toxicity: the blood-brain barrier is not intact in puppies having the same age as the ones used in this experiment, thus allowing cisplatin to reach CNS. Therefore, differ from what might be found in a CIPN exclusive model. This way of analysing gait would be more in line with kinematics or foot positioning during gait (see Table 1).

Another study (Contreras et al., 1997) measured stride length, gait support, toe spread and internal toe spread, using a similar method which represents the ability of the animal to control placement of its hind paws when walking. Administration of VCR resulted in significant reductions only in stride length and gait support of 17.3 and 10.1 mm, respectively compared to the control group (which had no change) at 10 weeks. Other works, in which all animal's paws were dyed, (Shahid et al., 2017, 2019), allowed to analyse the stride length (average distance of forward movement between each stride) on the right side and the overlap (distance between the front and hind footprints on each side) on both sides. However, they did not find any difference in CIS groups compared to the control group.

Another outcome of interest was defined as the area of footprint of the paw measured in pixels (Sahranavard et al., 2022). The group treated with VCR decrease the pressure of the foot by the measurement of pixel values in comparison with the control group (1248 pixels vs 9272 pixels); according to the Authors, this decrease would be due to a weakening of the muscles of the paws.

# 3.1.3. CatWalk™

Four studies used Semi-Automated methods for gait analysis (Boehmerle et al., 2014; Huehnchen et al., 2013; Liu et al., 2018; Shahid et al., 2019). This method does not rely on behaviour evoked by an artificial stimulus and is less influenced by the investigator, and it is also characterised by methodological simplicity and automaticity.

For a reliable comparison (both inter-studies or inter-groups) the gait parameters of groups it is of utmost importance that there is no difference in speed of gait among groups of animals; as already stated, in fact, the speed of gait is known to strongly affect gait parameters analysis with this technique (Deumens et al., 2007). In these four studies, the trial were regarded as successful if rodents did not stop on the runway and the cut-off time for running was 15 and 5 s for rats and mice, respectively (Liu et al., 2018; Shahid et al., 2019), or if the animal did not show a maximum speed variation greater than 60% (Boehmerle et al., 2014; Huehnchen et al., 2013) or did not exceed a walking speed of 400 mm/s (Boehmerle et al., 2014; Huehnchen et al., 2013).

As mentioned above, CatWalk<sup>TM</sup> can analyse pain-related outcomes through duration of the swing phase (no paw contact), duration of the stance phase (paw contact), and duty cycle (%) which is stand time divided by stand time plus swing time. Nevertheless, as mentioned by other Authors (Heinzel et al., 2020a) the term "pain-related" does not imply that the parameters subsumed under it are necessarily exclusively related to pain, but can for example also be influenced by impaired weight-loading due to muscle weakness or altered sensation, other than pain (e.g. numbness (Deumens et al., 2007; Li et al., 2014).

Literature data show some inconsistencies. Stance phase in hind paws were recorded as decreasing from day 10 until day 30 after the start of chemotherapy in two similar studies which used control animals to normalise the data from treated animals (Boehmerle et al., 2014; Huehnchen et al., 2013). Specifically, the reductions were by 90% after 14 days, 82% after 30 days at high doses of PTX (240 mg/kg) (Huehnchen et al., 2013), and more slightly reduced at minimal doses by 11% in PTX after 13 days, 17% in CIS after 20 days and 16% in VCR after 9 days. However, mice receiving BTZ reduced stance phase only in fore paws by 12% after 26 days (Boehmerle et al., 2014). These findings were only observed at a later time point, and thus later than the development of electrophysiological measurements and mechanical allodynia. Another study, found the opposite finding, they suggested that stance phase is increased in forepaws 10 days after VCR. It should be noted that this opposite findings, should be because of the induction of pain instead of CIPN in VCR administration in Bluette et al. study, as reported previously (Authier et al., 2003) or by the fact that 2 out of 7 mice did not show axonal degeneration in microscopy analysis (Bluette et al., 2021).

Parallel to these changes, the same findings were found in duty cycle

(as expected, because this outcome is calculated through stance phase and swing phase). Higher doses of PTX reduced duty cycle in the hind paws by 94% after 14 days, and by 92% after 30 days (Huehnchen et al., 2013). Comparing different treatments, at lower doses, Boehmerle et al. found a reduction at late points by 7% in PTX, by 12% by CIS, by 6% by VCR in hind paws. As before, BTZ induced a reduction in duty cycle only in the forepaws by 4% at the late time point (26 days after the start of chemotherapy). In contrast with these findings, a significant increase in duty cycle was documented in fore and hind paws (Bluette et al., 2021); however, a true CIPN model may not have been achieved in their study.

Another outcome reported was the print area, which is considered as a general gait parameter and it is the most reported outcome in peripheral nerve injury (Heinzel et al., 2020a). Two out of 4 studies reported significant differences in this outcome. On one hand, the first one, reported a reduction in day 30 after the start of PTX (20 mg/kg cumulative doses) by 68% in hind paws compared to baseline (Huehnchen et al., 2013). On the other hand, Lui et al. (Liu et al., 2018) reported (only in mice) an increase in print area both in fore and hind paws after 5, 12 and 15 days after first doses of VCR. This last finding should be interpreted cautiously, due to the fact that they did not find any microscopic findings in sciatic nerve or the dorsal root ganglion (Liu et al., 2018), therefore, this model might have not induced CIPN; however, sciatic nerve could be a too much proximal site to look for PTX-induced CIPN as well as DRG: it cannot be ruled out that a mild distal axonopathy was present (Pozzi et al., 2023; Wozniak et al., 2018).

# 4. Discussion

CIPN-related sensory ataxia is, potentially, a late neurological toxicity that impairs quality of life of cancer survivors (Cavaletti et al., 2015; Park et al., 2022; Tamburin et al., 2022). This condition is still little known and quite often confounded with a motor impairment which is not actually present (Cavaletti et al., 2019). The diminished manual dexterity and the impaired gait/balance can be impactful in patients' daily life, also decreasing their working ability, making this condition impactful both for the individual and the society (Cavaletti et al., 2023; Pike et al., 2012). Clinical trials addressing this condition, based mostly on physical treatments, are increasing in number but there is not definite evidence on the best approach for treatment and prevention of CIPN-related sensory ataxia (Park et al., 2022; Tamburin et al., 2022); this is due, mostly, to a lack of consensus on the ideal study design and outcome measures to evaluate CIPN per se in a clinical trial (Argyriou et al., 2019; Cavaletti et al., 2010), even if some indications were given in the last few years by international large study groups and experts' opinion: a combination of robust physician based scales, such as the Total Neuropathy Score (TNS®), with patients' reported outcomes is strongly encouraged (Alberti et al., 2021; Alberti et al., 2014; Cavaletti et al., 2013; Dorsey et al., 2019; Gewandter et al., 2018). However, for what regards the specific case of sensory ataxia, a formal evaluation such as gait analysis could be a relevant and powerful tool that still waits to be implemented in this setting, specifically for interventional clinical trials (Jiang et al., 2022; Wang et al., 2022). This is even truer if the preclinical setting is considered too: in absence of a defined CIPN treatment (Loprinzi et al., 2020), in fact, preclinical research is need to devise mechanisms of damage and test novel neuroprotectant drugs based on a sound biological rationale (Alberti, 2017; Bruna et al., 2020). In case of sensory ataxia, if a transition from bench to bed side is to be suggested, for sure gait analysis is a fair option to obtain a more objective evaluation in order to test novel hypotheses and then go back to the bed side. However, gait analysis in animal models can be performed with several different approaches and there is not a guideline specifically addressing its use in CIPN. Therefore, we presented a detailed description of possible strategies to analyse sensory ataxia in CIPN rodent models, addressing advantages and disadvantages of different methods available.

semi-automated methods - which do not rely on behaviour evoked by an artificial stimulus and are less influenced by the investigator - may be of interest when preventive or therapeutic strategies are evaluated, also considering their methodological simplicity and automaticity; up to now, only CatWalk<sup>TM</sup> analysis has been tested in CIPN setting.

Notably, it should be highlighted that comparison of literature data was complicated by the fact that CIPN models differ greatly among different studies; this is a crucial issue since a different schedule and, even, a different animal strain can determine a more mild/severe CIPN, or even not inducing CIPN at all but just a nocifensive behaviour (Pozzi et al., 2020). The first indication that should be given, in fact, it is that a robust preclinical CIPN model should rely on a multimodal approach exploiting neuropathology, neurophysiology, behavioural tests, to ensure the schedule actually induced nerve damage (Bruna et al., 2020; Monza et al., 2021a; b; Pozzi et al., 2020; Pozzi et al., 2023).

Taking into account these limitations, we can state that the few available studies documented gait disturbances at the last time point, and only appeared later on respect to other earlier signs of CIPN (such as altered neurophysiological findings). For that reason, gait impairment could be interpreted as late repercussions sensory loss. Future studies should expect a decremental effect on stance phase and print area in several agents chemotherapeutic such as PTX, CIS, VCR in hind paws. We hypothesised that chemotherapy-treated rodents tend to reduce hind paw support due to pain, allodynia or loss of sensation, and an increase in swing phase could or should be observed; this theory has been documented by Huehnchen et al. (Huehnchen et al., 2013), but unfortunately did not show significant differences.

Nevertheless, alterations in gait must be interpreted with caution in these in vivo models. These changes can also be related to a possible functional adaptation to maintain an inconspicuous gait, given that rats are a prey species and try to avoid showing pain or disability to potential predators(Graham, 2016). It is, therefore, recommended to use automated gait analysis as a complementary tool(Heinzel et al., 2020b).

Some other functional tests mentioned above might be eventually explored in the future, translating some lessons learnt from central nervous system disease model, taking into account the behaviour as it is in the legde test (Guyenet et al., 2010) and LRWT (Fey et al., 2010); these tests could give an overview of coordination and motor component during gait. So far, though, in preclinical studies they were used for the measurement of cerebral ataxia or other neurodegenerative syndromes, and not for CIPN models.

# 5. Concluding remarks

In conclusion, it can be suggested that further studies are needed to consolidate the use of gait analysis in rodent models. It is crucial that the schedule is adequate to induce CIPN fully (e.g., not just a nocifensive behaviour) and that CIPN onset is carefully assessed with objective methods such as histopathology and neurophysiology (Alberti et al., 2020; Ballarini et al., 2022; Bruna et al., 2020; Monza et al., 2021a; b; Pozzi et al., 2023). Empowering this tool in preclinical models would be of great value since gait analysis is being introduced also in CIPN clinical trials, especially the ones exploring the role of physical therapy treatment to cure CIPN (Lopez-Garzon et al., 2022) (Park et al., 2022): having a translational outcome measure to test sensory ataxia similarly at bench and bed side would allow to promptly translate preclinical evidence to a robust clinical trial.

# Funding

MLG has received funding for its training with the grant FI19/00230 and MV22/00095 by the Fondo de Investigación Sanitaria del Instituto de Salud Carlos III (Spain).

From the literature data presented so far, it could be suggested that

# **Declaration of Competing Interest**

None.

# Data availability

No data was used for the research described in the article.

## References

- Abbas, W., Masip Rodo, D., 2019. Computer methods for automatic locomotion and gesture tracking in mice and small animals for neuroscience applications: a survey. Sens. Basel 19.
- Alberti, P., 2017. Chemotherapy-induced peripheral neurotoxicity outcome measures: the issue. Expert Opin. Drug Metab. Toxicol. 13, 241–243.
- Alberti, P., Rossi, E., Cornblath, D.R., Merkies, I.S., Postma, T.J., Frigeni, B., Bruna, J., Velasco, R., Argyriou, A.A., Kalofonos, H.P., Psimaras, D., Ricard, D., Pace, A., Galiè, E., Briani, C., Dalla Torre, C., Faber, C.G., Lalisang, R.I., Boogerd, W., Brandsma, D., Koeppen, S., Hense, J., Storey, D., Kerrigan, S., Schenone, A., Fabbri, S., Valsecchi, M.G., Cavaletti, G., Group, C.-P., 2014. Physician-assessed and patient-reported outcome measures in chemotherapy-induced sensory peripheral neurotoxicity: two sides of the same coin. Ann. Oncol. 25, 257–264.
- Alberti, P., Canta, A., Chiorazzi, A., Fumagalli, G., Meregalli, C., Monza, L., Pozzi, E., Ballarini, E., Rodriguez-Menendez, V., Oggioni, N., Sancini, G., Marmiroli, P., Cavaletti, G., 2020. Topiramate prevents oxaliplatin-related axonal hyperexcitability and oxaliplatin induced peripheral neurotoxicity. Neuropharmacology 164, 107905.
- Alberti, P., Bernasconi, D.P., Cornblath, D.R., Merkies, I.S.J., Park, S.B., Velasco, R., Bruna, J., Psimaras, D., Koeppen, S., Pace, A., Dorsey, S.G., Argyriou, A.A., Kalofonos, H.P., Briani, C., Schenone, A., Faber, C.G., Mazzeo, A., Grisold, W., Valsecchi, M., Cavaletti, G., group, C.-P., 2021. Prospective evaluation of health care provider and patient assessments in chemotherapy-induced peripheral neurotoxicity. Neurology 97, e660–e672.
- Alberti, P., Salvalaggio, A., Argyriou, A.A., Bruna, J., Visentin, A., Cavaletti, G., Briani, C., 2022. Neurological complications of conventional and novel anticancer treatments. Cancers Basel 14.
- Altmann, C., Vasic, V., Hardt, S., Heidler, J., Häussler, A., Wittig, I., Schmidt, M.H.H., Tegeder, I., 2016. Progranulin promotes peripheral nerve regeneration and reinnervation: role of notch signaling. Mol. Neurodegener. 11, 69.
- Araújo-Filho, H.G., Pereira, E.W.M., Heimfarth, L., Souza Monteiro, B., Santos Passos, F. R., Siqueira-Lima, P., Gandhi, S.R., Viana Dos Santos, M.R., Guedes da Silva Almeida, J.R., Picot, L., Grougnet, R., Almeida, R.S., Douglas Melo Coutinho, H., Quintans-Júnior, L.J., Martins, N., Quintans, J.S.S., 2020. Limonene, a food additive, and its active metabolite perillyl alcohol improve regeneration and attenuate neuropathic pain after peripheral nerve injury: evidence for IL-1β, TNF-α, GAP, NGF and ERK involvement. Int. Immunopharmacol. 86, 106766.
- Argyriou, A.A., Park, S.B., Islam, B., Tamburin, S., Velasco, R., Alberti, P., Bruna, J., Psimaras, D., Cavaletti, G., Cornblath, D.R., (TNC), T.N.C, 2019. Neurophysiological, nerve imaging and other techniques to assess chemotherapy-induced peripheral neurotoxicity in the clinical and research settings. J. Neurol. Neurosurg. Psychiatry.
- Authier, N., Gillet, J.P., Fialip, J., Eschalier, A., Coudore, F., 2003. A new animal model of vincristine-induced nociceptive peripheral neuropathy. Neurotoxicology 24, 797–805.
- Ballarini, E., Malacrida, A., Rodriguez-Menendez, V., Pozzi, E., Canta, A., Chiorazzi, A., Monza, L., Semperboni, S., Meregalli, C., Carozzi, V.A., Hashemi, M., Nicolini, G., Scuteri, A., Housley, S.N., Cavaletti, G., Alberti, P., 2022. Sodium-Calcium Exchanger 2: a pivotal role in oxaliplatin induced peripheral neurotoxicity and axonal damage. Int. J. Mol. Sci. 23.
- Bluette, C.T., Shoieb, A.M., Peng, Q., Manickam, B., Huang, W., Shin, E., Zhang, W., Song, Y.H., Liu, C.N., 2021. Behavioral, histopathologic, and molecular biological responses of nanoparticle- and solution-based formulations of vincristine in mice. Int. J. Toxicol. 40, 40–51.
- Boehmerle, W., Huehnchen, P., Peruzzaro, S., Balkaya, M., Endres, M., 2014. Electrophysiological, behavioral and histological characterization of paclitaxel, cisplatin, vincristine and bortezomib-induced neuropathy in C57Bl/6 mice. Sci. Rep. 4, 6370.
- Boyle, F.M., Wheeler, H.R., Shenfield, G.M., 1996. Glutamate ameliorates experimental vincristine neuropathy. J. Pharmacol. Exp. Ther. 279, 410–415.
- Boyle, F.M., Wheeler, H.R., Shenfield, G.M., 1999. Amelioration of experimental cisplatin and paclitaxel neuropathy with glutamate. J. Neurooncol 41, 107–116.
   Boyle, F.M., Beatson, C., Monk, R., Grant, S.L., Kurek, J.B., 2001. The experimental
- Boyle, F.M., Beatson, C., MORK, A., Grant, S.L., KHER, J.B., 2001. The experimental neuroprotectant leukaemia inhibitory factor (LIF) does not compromise antitumour activity of paclitaxel, cisplatin and carboplatin. Cancer Chemother. Pharmacol. 48, 429–434.
- Briani, C., Argyriou, A.A., Izquierdo, C., Velasco, R., Campagnolo, M., Alberti, P., Frigeni, B., Cacciavillani, M., Bergamo, F., Cortinovis, D., Cazzaniga, M., Bruna, J., Cavaletti, G., Kalofonos, H.P., 2014. Long-term course of oxaliplatin-induced polyneuropathy: a prospective 2-year follow-up study. J. Peripher Nerv. Syst. 19, 299–306.
- Bruna, J., Alberti, P., Calls-Cobos, A., Caillaud, M., Damaj, M.I., Navarro, X., 2020. Methods for in vivo studies in rodents of chemotherapy induced peripheral neuropathy. Exp. Neurol. 325, 113154.

Castillo-Mariqueo, L., Giménez-Llort, L., 2022. Clasping, ledge-score coordination and early gait impairments as primary behavioural markers of functional impairment in Alzheimer's disease. Behav. Brain Res. 435, 114054.

Cavaletti, G., Nicolini, G., Marmiroli, P., 2008. Neurotoxic effects of antineoplastic drugs: the lesson of pre-clinical studies. Front. Biosci. (13), 3506–3524.

- Cavaletti, G., Frigeni, B., Lanzani, F., Mattavelli, L., Susani, E., Alberti, P., Cortinovis, D., Bidoli, P., 2010. Chemotherapy-Induced Peripheral Neurotoxicity assessment: a critical revision of the currently available tools. Eur. J. Cancer 46, 479–494.
- Cavaletti, G., Cornblath, D.R., Merkies, I.S., Postma, T.J., Rossi, E., Frigeni, B., Alberti, P., Bruna, J., Velasco, R., Argyriou, A.A., Kalofonos, H.P., Psimaras, D., Ricard, D., Pace, A., Galiè, E., Briani, C., Dalla Torre, C., Faber, C.G., Lalisang, R.I., Boogerd, W., Brandsma, D., Koeppen, S., Hense, J., Storey, D., Kerrigan, S., Schenone, A., Fabbri, S., Valsecchi, M.G., Group, C.-P., 2013. The chemotherapy-induced peripheral neuropathy outcome measures standardization study: from consensus to the first validity and reliability findings. Ann. Oncol. 24, 454–462.
- Cavaletti, G., Alberti, P., Marmiroli, P., 2015. Chemotherapy-induced peripheral neurotoxicity in cancer survivors: an underdiagnosed clinical entity. Am. Soc. Clin. Oncol. Educ. Book e553–e560.
- Cavaletti, G., Cornblath, D.R., Merkies, I.S.J., Postma, T.J., Rossi, E., Alberti, P., Bruna, J., Argyriou, A.A., Briani, C., Velasco, R., Kalofonos, H.P., Psimaras, D., Ricard, D., Pace, A., Faber, C.G., Lalisang, R.I., Brandsma, D., Koeppen, S., Kerrigan, S., Schenone, A., Grisold, W., Mazzeo, A., Padua, L., Dorsey, S.G., Penas-Prado, M., Valsecchi, M.G., Group, C.-P., 2019. Patients' and physicians' interpretation of chemotherapy-induced peripheral neurotoxicity. J. Peripher Nerv. Syst.
- Cavaletti, G., D'Acunti, A., Porcu, A., Masiello, G., Del Campo, L., Traclò, G., De Lorenzo, F., Bernasconi, D.P., 2023. Self-reported assessment of the socio-economic impact of anticancer chemotherapy-related neurotoxicity. Toxics 11.
- Chen, C.S., Kim, J., Garg, N., Guntupalli, H., Jagsi, R., Griggs, J.J., Sabel, M., Dorsch, M. P., Callaghan, B.C., Hertz, D.L., 2021. Chemotherapy-induced peripheral neuropathy detection via a smartphone app: cross-sectional pilot study. JMIR Mhealth Uhealth 9, e27502.
- Chou, A.H., Yeh, T.H., Ouyang, P., Chen, Y.L., Chen, S.Y., Wang, H.L., 2008. Polyglutamine-expanded ataxin-3 causes cerebellar dysfunction of SCA3 transgenic mice by inducing transcriptional dysregulation. Neurobiol. Dis. 31, 89–101.
- Contreras, P.C., Vaught, J.L., Gruner, J.A., Brosnan, C., Steffler, C., Arezzo, J.C., Lewis, M.E., Kessler, J.A., Apfel, S.C., 1997. Insulin-like growth factor-I prevents development of a vincristine neuropathy in mice. Brain Res. 774, 20–26.
- Deumens, R., Jaken, R.J., Marcus, M.A., Joosten, E.A., 2007. The CatWalk gait analysis in assessment of both dynamic and static gait changes after adult rat sciatic nerve resection. J. Neurosci. Methods 164, 120–130.
- Dorsey, S.G., Kleckner, I.R., Barton, D., Mustian, K., O'Mara, A., St Germain, D., Cavaletti, G., Danhauer, S.C., Hershman, D., Hohmann, A.G., Hoke, A., Hopkins, J. O., Kelly, K.P., Loprinzi, C.L., McLeod, H.L., Mohile, S., Paice, J., Rowland, J.H., Salvemini, D., Segal, R.A., Lavoie Smith, E., McCaskill Stevens, W., Janelsins, M.C., 2019. NCI Clinical Trials Planning Meeting for prevention and treatment of chemotherapy-induced peripheral neuropathy. J Natl Cancer Inst.
- Fey, A., Schachner, M., Irintchev, A., 2010. A novel motion analysis approach reveals late recovery in C57BL/6 mice and deficits in NCAM-deficient mice after sciatic nerve crush. J. Neurotrauma 27, 815–828.
- Gewandter, J.S., Brell, J., Cavaletti, G., Dougherty, P.M., Evans, S., Howie, L., McDermott, M.P., O'Mara, A., Smith, A.G., Dastros-Pitei, D., Gauthier, L.R., Haroutounian, S., Jarpe, M., Katz, N.P., Loprinzi, C., Richardson, P., Lavoie-Smith, E. M., Wen, P.Y., Turk, D.C., Dworkin, R.H., Freeman, R., 2018. Trial designs for chemotherapy-induced peripheral neuropathy prevention: acttion recommendations. Neurology 91, 403–413.
- Graham, D.M., 2016. Methods for measuring pain in laboratory animals. Lab Anim. 45, 99–101.
- Guyenet, S.J., Furrer, S.A., Damian, V.M., Baughan, T.D., La Spada, A.R., Garden, G.A., 2010. A simple composite phenotype scoring system for evaluating mouse models of cerebellar ataxia. J. Vis. Exp.
- Gyengesi, E., Rangel, A., Ullah, F., Liang, H., Niedermayer, G., Asgarov, R., Venigalla, M., Gunawardena, D., Karl, T., Münch, G., 2019. Chronic microglial activation in the GFAP-IL6 mouse contributes to age-dependent cerebellar volume loss and impairment in motor function. Front Neurosci. 13, 303.
- Heinzel, J., Längle, G., Oberhauser, V., Hausner, T., Kolbenschlag, J., Prahm, C., Grillari, J., Hercher, D., 2020a. Use of the CatWalk gait analysis system to assess functional recovery in rodent models of peripheral nerve injury - a systematic review. J. Neurosci. Methods 345, 108889.
- Heinzel, J., Swiadek, N., Ashmwe, M., Rührnößl, A., Oberhauser, V., Kolbenschlag, J., Hercher, D., 2020b. Automated gait analysis to assess functional recovery in rodents with peripheral nerve or spinal cord contusion injury. J. Vis. Exp.
- Huehnchen, P., Boehmerle, W., Endres, M., 2013. Assessment of paclitaxel induced sensory polyneuropathy with "Catwalk" automated gait analysis in mice. PLoS One 8, e76772.
- Inserra, M.M., Bloch, D.A., Terris, D.J., 1998. Functional indices for sciatic, peroneal, and posterior tibial nerve lesions in the mouse. Microsurgery 18, 119–124.
- Jiang, X., Deng, F., Rui, S., Ma, Y., Wang, M., Deng, B., Wang, H., Du, C., Chen, B., Yang, X., Boey, J., Armstrong, D.G., Deng, W., Duan, X., 2022. The evaluation of gait and balance for patients with early diabetic peripheral neuropathy: a cross-sectional study. Risk Manag. Health Policy 15, 543–552.
- Kim, S.M., Lee, S.K., Lee, J.H., 2007. Peripheral nerve regeneration using a three dimensionally cultured schwann cell conduit. J. Craniofac Surg. 18, 475–488.
- Kolb, N.A., Smith, A.G., Singleton, J.R., Beck, S.L., Stoddard, G.J., Brown, S., Mooney, K., 2016. The association of chemotherapy-induced peripheral neuropathy symptoms and the risk of falling. JAMA Neurol.

# M. Lopez-Garzon et al.

Lee, J.Y., Giusti, G., Wang, H., Friedrich, P.F., Bishop, A.T., Shin, A.Y., 2013. Functional evaluation in the rat sciatic nerve defect model: a comparison of the sciatic functional index, ankle angles, and isometric tetanic force. Plast. Reconstr. Surg. 132, 1173–1180.

- Li, M.T., Willett, N.J., Uhrig, B.A., Guldberg, R.E., Warren, G.L., 2014. Functional analysis of limb recovery following autograft treatment of volumetric muscle loss in the quadriceps femoris. J. Biomech. 47, 2013–2021.
- Liu, C.N., Berryman, E., Zakur, D., Shoieb, A.M., Pardo, I.D., Boucher, M., Somps, C.J., Bagi, C.M., Cook, J.C., 2018. A novel endpoint for the assessment of chemotherapyinduced peripheral neuropathy in rodents: biomechanical properties of peripheral nerve. J. Appl. Toxicol. 38, 193–200.
- Lopez-Garzon, M., Cantarero-Villanueva, I., Postigo-Martin, P., González-Santos, Á., Lozano-Lozano, M., Galiano-Castillo, N., 2022. Can physical exercise prevent chemotherapy-induced peripheral neuropathy in patients with cancer? A systematic review and meta-analysis. Arch. Phys. Med. Rehabil. 103, 2197–2208.
- Loprinzi, C.L., Lacchetti, C., Bleeker, J., Cavaletti, G., Chauhan, C., Hertz, D.L., Kelley, M. R., Lavino, A., Lustberg, M.B., Paice, J.A., Schneider, B.P., Lavoie Smith, E.M., Smith, M.L., Smith, T.J., Wagner-Johnston, N., Hershman, D.L., 2020. Prevention and management of chemotherapy-induced peripheral neuropathy in survivors of adult cancers: ASCO guideline update. J. Clin. Oncol. JCO2001399.
- Matsuda, K., Orito, K., Amagai, Y., Jang, H., Matsuda, H., Tanaka, A., 2016. Swing time ratio, a new parameter of gait disturbance, for the evaluation of the severity of neuropathic pain in a rat model of partial sciatic nerve ligation. J. Pharmacol. Toxicol. Methods 79, 7–14.
- Metz, G.A., Whishaw, I.Q., 2009. The ladder rung walking task: a scoring system and its practical application. J. Vis. Exp.
- Mols, F., Beijers, T., Vreugdenhil, G., van de Poll-Franse, L., 2014. Chemotherapyinduced peripheral neuropathy and its association with quality of life: a systematic review. Support Care Cancer 22, 2261–2269.
- Monza, L., Fumagalli, G., Chiorazzi, A., Alberti, P., 2021a. Addressing the need of a translational approach in peripheral neuropathy research: morphology meets function. Brain Sci. 11.
- Monza, L., Fumagalli, G., Chiorazzi, A., Alberti, P., 2021b. Translating morphology from bench side to bed side via neurophysiology: 8-min protocol for peripheral neuropathy research. J. Neurosci. Methods 363, 109323.
- Ouzzani, M., Hammady, H., Fedorowicz, Z., Elmagarmid, A., 2016. Rayyan-a web and mobile app for systematic reviews. Syst. Rev. 5, 210.
- Ozkan, O., Duman, O., Haspolat, S., Ozgentaş, H.E., Dikici, M.B., Gürer, I., Güngör, H.A., Güzide Gökhan, A., 2005. Effect of systemic creatine monohydrate supplementation on denervated muscle during reinnervation: experimental study in the rat. J. Reconstr. Microsurg 21, 573–579.
- Park, S.B., Tamburin, S., Schenone, A., Kleckner, I.R., Velasco, R., Alberti, P., Kanzawa-Lee, G., Lustberg, M., Dorsey, S.G., Mantovani, E., Hamedani, M., Argyriou, A.A., Cavaletti, G., Hoke, A., Consortium, T.N., 2022. Optimal outcome measures for assessing exercise and rehabilitation approaches in chemotherapy-induced peripheral-neurotoxicity: systematic review and consensus expert opinion. Expert Rev. Neurother. 22, 65–76.
- Piesla, M.J., Leventhal, L., Strassle, B.W., Harrison, J.E., Cummons, T.A., Lu, P., Whiteside, G.T., 2009. Abnormal gait, due to inflammation but not nerve injury, reflects enhanced nociception in preclinical pain models. Brain Res. 1295, 89–98.
- Pike, C.T., Birnbaum, H.G., Muehlenbein, C.E., Pohl, G.M., Natale, R.B., 2012. Healthcare costs and workloss burden of patients with chemotherapy-associated peripheral neuropathy in breast, ovarian, head and neck, and nonsmall cell lung cancer. Chemother. Res. Pr. 2012, 913848.
- Pozzi, E., Fumagalli, G., Chiorazzi, A., Canta, A., Meregalli, C., Monza, L., Carozzi, V.A., Oggioni, N., Rodriguez-Menendez, V., Cavaletti, G., Marmiroli, P., 2020. The relevance of multimodal assessment in experimental oxaliplatin-induced peripheral neurotoxicity. Exp. Neurol. 334, 113458.

- Pozzi, E., Monza, L., Ballarini, E., Bossi, M., Rodriguez-Menendez, V., Canta, A., Chiorazzi, A., Carozzi, V.A., Crippa, L., Marmiroli, P., Cavaletti, G., Alberti, P., 2023. Morpho-functional characterisation of the rat ventral caudal nerve in a model of axonal peripheral neuropathy. Int. J. Mol. Sci. 24.
- Sahranavard, S., Khoramjouy, M., Khakpash, M., Askari, S.A., Faizi, M., Mosaddegh, M., 2022. Hydroethanolic extract of. Res. Pharm. Sci. 17, 265–273.
- Santos, P.M., 2000. A functional model system of an hypoxic nerve injury and its evaluation. Laryngoscope 110, 845–853.
- Shahid, M., Subhan, F., Ahmad, N., Sewell, R.D.E., 2017. The flavonoid 6-methoxyflavone allays cisplatin-induced neuropathic allodynia and hypoalgesia. Biomed. Pharm. 95, 1725–1733.
- Shahid, M., Subhan, F., Ahmad, N., Sewell, R.D.E., 2019. Efficacy of a topical gabapentin gel in a cisplatin paradigm of chemotherapy-induced peripheral neuropathy. BMC Pharm. Toxicol. 20, 51.
- Tamburin, S., Park, S.B., Schenone, A., Mantovani, E., Hamedani, M., Alberti, P., Yildiz-Kabak, V., Kleckner, I.R., Kolb, N., Mazzucchelli, M., McNeish, B.L., Argyriou, A.A., Cavaletti, G., Hoke, A., Consortium, T.N., 2022. Rehabilitation, exercise, and related non-pharmacological interventions for chemotherapy-induced peripheral neurotoxicity: systematic review and evidence-based recommendations. Crit. Rev. Oncol. Hematol. 171, 103575.
- Thomas, P.S., Fraley, G.S., Damian, V., Woodke, L.B., Zapata, F., Sopher, B.L., Plymate, S. R., La Spada, A.R., 2006. Loss of endogenous androgen receptor protein accelerates motor neuron degeneration and accentuates androgen insensitivity in a mouse model of X-linked spinal and bulbar muscular atrophy. Hum. Mol. Genet 15, 2225–2238.
- Umansky, D., Hagen, K.M., Chu, T.H., Pathiyil, R.K., Alzahrani, S., Ousman, S.S., Midha, R., 2022. Functional gait assessment using manual, semi-automated and deep learning approaches following standardized models of peripheral nerve injury in mice. Biomolecules 12.
- Varejão, A.S., Cabrita, A.M., Geuna, S., Melo-Pinto, P., Filipe, V.M., Gramsbergen, A., Meek, M.F., 2003. Toe out angle: a functional index for the evaluation of sciatic nerve recovery in the rat model. Exp. Neurol. 183, 695–699.
- Velasco, R., Bruna, J., 2010. Chemotherapy-induced peripheral neuropathy: an unresolved issue. Neurologia 25, 116–131.
- Walker, J.L., Evans, J.M., Meade, P., Resig, P., Sisken, B.F., 1994. Gait-stance duration as a measure of injury and recovery in the rat sciatic nerve model. J. Neurosci. Methods 52, 47–52.
- Wang, Z., Peng, S., Zhang, H., Sun, H., Hu, J., 2022. Gait parameters and peripheral neuropathy in patients with diabetes: a meta-analysis. Front Endocrinol. Lausanne 13, 891356.
- Whitaker-Azmitia, P.M., Raio, M., Raio, D., Borella, A., 1995. A 5-HT3 receptor antagonist fails to prevent cisplatin-induced toxicity in immature rat spinal cord. Eur. J. Pharmacol. 275, 139–143.
- Wozniak, K.M., Vornov, J.J., Wu, Y., Liu, Y., Carozzi, V.A., Rodriguez-Menendez, V., Ballarini, E., Alberti, P., Pozzi, E., Semperboni, S., Cook, B.M., Littlefield, B.A., Nomoto, K., Condon, K., Eckley, S., DesJardins, C., Wilson, L., Jordan, M.A., Feinstein, S.C., Cavaletti, G., Polydefkis, M., Slusher, B.S., 2018. Peripheral neuropathy induced by microtubule-targeted chemotherapies: insights into acute injury and long-term recovery. Cancer Res. 78, 817–829.
- injury and long-term recovery. Cancer Res. 78, 817–829. Xu, Y., Tian, N.X., Bai, Q.Y., Chen, Q., Sun, X.H., Wang, Y., 2019. Gait assessment of pain and analgesics: comparison of the DigiGait<sup>™</sup> and CatWalk<sup>™</sup> Gait Imaging Systems. Neurosci. Bull. 35, 401–418.
- Zahiri, M., Chen, K.M., Zhou, H., Nguyen, H., Workeneh, B.T., Yellapragada, S.V., Sada, Y.H., Schwenk, M., Najafi, B., 2019. Using wearables to screen motor performance deterioration because of cancer and chemotherapy-induced peripheral neuropathy (CIPN) in adults - Toward an early diagnosis of CIPN. J. Geriatr. Oncol. 10, 960–967.