



Is less always more? Ceftazidime/avibactam alone or in combination to treat infections caused by *Pseudomonas aeruginosa*: a retrospective observational multi-centre cohort study

Pietro Valsecchi¹ · Marta Colaneri² · Andrea Lombardi^{3,4} · Simona Biscarini³ · Cecilia Azzarà^{3,4} · Alessandra Bandera^{3,4} · Nicolò Corti⁵ · Paolo Bonfanti⁵ · Leonardo Francesco Rezzonico^{6,7} · Giovanna Travi⁶ · Silvia Pontiggia⁸ · Stefania Piconi⁸ · Arianna Lippi^{9,10} · Mario Tumbarello^{9,10} · Matteo Giovanni Lupi¹¹ · Angelo Pan¹¹ · Giacomo Casalini^{2,12} · Spinello Antinori² · Andrea Gori² · Alessandra Calabresi¹³ · Guido Chichino¹³ · Virginia Valeria Ferretti¹⁴ · Giulia Gambini¹⁴ · Catherine Klersy¹⁴ · Raffaele Bruno^{1,7}

Received: 18 July 2025 / Accepted: 9 July 2026

© The Author(s), under exclusive licence to Springer-Verlag GmbH Germany, part of Springer Nature 2026

Abstract

Purpose In vitro and in vivo studies suggest adding a second agent to ceftazidime/avibactam (CZA) to enhance bactericidal activity against *Pseudomonas aeruginosa* (PA). Nevertheless, evidence supporting this practice is scarce.

Methods We conducted a retrospective cohort study of patients admitted to 9 Italian hospitals with microbiologically confirmed PA infection and treated with CZA alone or in combination from 01/01/2015 to 31/12/2021. The primary endpoint was 14-day clinical cure. Secondary endpoints were 28-day all-cause mortality, 56-day microbiological failure, 56-day infection relapse, and emergence of CZA resistance. A generalized linear model extended to the binomial family, with identity link, weighted for IPTW (inverse probability of treatment weighting), was fitted to compare the two groups for 14-day clinical cure. Results were reported in terms of risk difference (RD) and 90% Confidence Interval (90%CI).

Results Eighty-seven patients with infection due to PA were included; 17 were treated with CZA monotherapy, and 70 with combination therapy with CZA plus another anti-pseudomonal agent. Sequential organ failure assessment was significantly higher (median 6 [3–9]) in the combination group compared to monotherapy (median 3 [1–5]; $p=0.049$). After weighting patients according to the IPTW, CZA monotherapy did not reach the clinical equipoise to combination therapy in terms of 14-day clinical cure (RD -0.28, 90% CI -0.41 to -0.14). No significant risk difference between groups was found for any of the secondary endpoints.

Conclusion Our findings suggest that combination therapy with CZA for treating PA infections may provide some clinical benefit over monotherapy despite the absence of a definitive survival advantage.

Keywords *Pseudomonas aeruginosa* · Ceftazidime/avibactam · Combination therapy

Background

Pseudomonas aeruginosa (PA) is a frequent cause of healthcare-associated infections, with a mortality rate exceeding 40% in severe cases such as bloodstream infections (BSIs) and ventilator-associated pneumonia (VAP) [1, 2]. Antimicrobial resistance is a significant concern, as more than 17.5% of the isolates are resistant to at least two antibiotics [3, 4]. Due to its resistance profile, the World Health

Organization included PA among the high-priority pathogens in terms of antimicrobial resistance [5].

Multi-drug resistant (MDR) PA is defined as being non-susceptible to at least one antimicrobial in at least three antimicrobial classes to which it is usually susceptible, while extensively drug-resistant (XDR) isolates are resistant to all but two or fewer classes [6]. In 2018, the concept of difficult-to-treat resistance (DTR) was introduced, describing PA resistant to key-antibiotics such as ceftazidime, cefepime,

Extended author information available on the last page of the article

piperacillin/tazobactam, imipenem-cilastatin, meropenem, ciprofloxacin, levofloxacin, and aztreonam [7].

Ceftazidime/avibactam (CZA) is a β -lactam/ β -lactamase inhibitor combination of the third-generation cephalosporin ceftazidime and the β -lactamase inhibitor avibactam. The combination with avibactam restores ceftazidime activity against a broad range of Gram-negative bacteria, including ESBL-, AmpC-, KPC and OXA 48-producing *Enterobacterales* and PA, but not against metallo- β -lactamase (MBL) producers [8].

CZA represents a valuable therapeutic option against PA isolates, as susceptibility rates range from 85% to 95% according to several epidemiological studies [9, 10]. Nevertheless, the emergence of clinical isolates resistant to CZA has been increasingly described since the introduction of the drug in 2015 [11, 12].

Emerging resistance to CZA in PA is mediated by several mechanisms, including OXA-2 and OXA-10 mutations and modifications at the catalytic site of AmpC enzymes, leading to enhanced ceftazidime hydrolysis and increased resistance to avibactam [13–16]. Other mechanisms of resistance include target site modifications involving penicillin-binding proteins (PBPs), porin mutations (OprD), and efflux pumps [11, 16].

One proposed strategy to counteract resistance is the combination of two active antibiotics. However, while this approach has shown promising results in vitro and in vivo animal studies, translation of this data in clinical practice requires further evidence [17, 18].

Whether combining a second antipseudomonal agent with CZA would improve patients' outcomes and reduce the emergence of CZA-resistant PA strains is still unclear [19, 20].

Due to insufficient clinical data, the 2022 guidelines from the European Society of Clinical Microbiology and Infectious Diseases do not recommend for or against combination therapy with CZA for carbapenem-resistant PA infections, while the Infectious Diseases Society of America 2023 guidance document advises against routine combination therapy for DTR PA [21, 22].

We hypothesize that CZA monotherapy would provide similar clinical and microbiological outcomes compared to CZA combined with other active antimicrobials for infections caused by PA. To test this hypothesis, we retrospectively collected data from multiple centres in Northern Italy and compared outcomes between the two treatment strategies.

Methods

Study design and participants

This was an explorative multicentre retrospective cohort study. Data from adult patients with microbiologically

confirmed PA infection admitted to the participating centres between the 1st of January 2015 and the 31st of December 2021 and treated with CZA for at least 72 h were retrospectively collected.

Patients were excluded if <18 years of age, if antibiotic treatment was empirically driven by colonization without an available bacterial isolate and antimicrobial susceptibility testing, if isolates were obtained from cultures of drainage sites after 48 h from drainage placement or swabs from non-sterile sites (e.g. chronic ulcers, wounds or intact skin), or if there was no assessment for the primary endpoint.

Objectives of the study

The primary endpoint of the study was the 14-day clinical cure, defined as resolution of signs and symptoms of infection. Specific infection types were defined according to CDC/NHSN Surveillance Definitions for Specific Types of Infections [23]. Clinical cure definitions were developed a priori for each infection and are presented in the [Supplementary materials](#).

Secondary endpoints included:

- Microbiologic failure within 56 days, defined as persistent positive cultures for the index pathogen from the same infection site. (endpoint 2.1)
- Infection relapse within 56 days, defined as the recurrence of the same type of PA infection after achieving clinical cure. (endpoint 2.2)
- Emergence of CZA resistance in follow-up isolates, defined as a new isolate with CZA minimum inhibitory concentrations (MIC) above the breakpoint. (endpoint 2.3)
- 28-day all-cause mortality. (endpoint 2.4)

Both the primary and secondary endpoints were calculated from the index culture collection.

Variables and definitions

Data were retrospectively retrieved from electronic health records by a dedicated infectious disease specialist at each participating centre.

For each patient, demographic data (age, gender), Charlson comorbidity index (CCI) [24] and presence of comorbidities, including those related to an increased risk for PA infection (cystic fibrosis, bronchiectasis, chronic obstructive pulmonary disease, active immunosuppression, neutrophil blood count < 500 cells/mm), were recorded [24–26]. Other relevant variables were intensive care unit (ICU) stay, estimated glomerular filtration rate (eGFR), need for renal replacement therapy, and any prior treatment with antipseudomonal antibiotics within the past 12 months.

Severity of infection onset was assessed using the Sequential Organ Failure Assessment (SOFA) score. The presence of septic shock was defined according to Sepsis-3 criteria as sepsis with persistent hypotension despite adequate fluid replacement and/or blood lactates $> 1,6$ mmol/L [27]. Presence of adequate source control was defined as all the acts aimed to remove the putative source of infection or to reduce the burden of infection, such as central line or urinary catheter removal, abscess drainage, and surgical debridement or removal of infected devices.

CZA was administered either by continuous infusion (over the entire dosing interval) or extended infusion (lasting at least 3 h).

Microbiological methods

Clinical isolates were identified in each centre's Microbiology Unit from biological samples, and antimicrobial susceptibility testing was performed in line with local standard procedures. Results were interpreted according to EUCAST clinical breakpoints [28]. MDR, XDR, and DTR isolates were defined as described above [6]. Combination therapy was considered effective when it was interpreted as susceptible or dose-dependent, according to EUCAST clinical breakpoints [28]; in the absence of clinical breakpoints for aminoglycosides and fosfomycin, their use in combination was interpreted as conforming to respective EUCAST rationale documents [29, 30].

Statistical methods

All statistical analyses were performed using Stata 18 (StataCorp. 2023. Stata Statistical Software: Release 18. College Station, TX: StataCorp LLC.). Two-sided type I error was set at 0.1.

Qualitative variables were described as counts and percentages of each category. Quantitative variables have been summarized as median and Interquartile Range (IQR). The association between 2 qualitative variables was evaluated via Fisher's exact test. Quantitative variables were compared between two groups using the Mann-Whitney test.

Propensity score-based methods were used to compare outcomes between CZA monotherapy and CZA combination therapy for the primary endpoint only, to control for confounding and reduce imbalances between the two treatment groups. Adjustment was not extended to secondary endpoints to preserve statistical power. After trimming the extreme 1% of observations, the data were weighted using the Inverse Probability of Treatment Weighting (IPTW) method.

Data were captured for patients treated with CZA alone versus those treated with CZA in combination. The following covariates were selected for propensity score calculation due to their clinical relevance: age at treatment, CCI, active immunosuppression, treatment with an anti-pseudomonal drug in the last 12 months, ICU admission, hospital-acquired pneumonia (HAP) /VAP, BSI, MDR, XDR, and DTR.

We included MDR, XDR, and DTR in the model since the study focused on targeted treatment, and this information was available to the treating physician when selecting antimicrobial therapy.

Multiple imputation was not performed due to the negligible amount of missing data for these variables.

A generalized linear model extended to the binomial family, with identity link (weighted for IPTW), was fitted to compare CZA alone and CZA combined for 14-day clinical cure (primary endpoint). Results of this model were reported in terms of risk difference (RD) with 90% Confidence Interval (90% CI).

Due to the exploratory nature of the study and the area of uncertainty over the effect of combination therapy on clinical cure, we sought to evaluate clinical equipoise between the two treatment arms. A lower limit of the confidence interval not exceeding -10% was set to ensure that the two treatment groups were comparable.

The same analysis (without weighting for IPTW) was performed to compare CZA alone and CZA combined for microbiologic failure, infection relapse, and emergence of CZA resistance (secondary endpoints 2.1, 2.2, 2.3).

Kaplan-Meier product limit method and Cox regression model were used to estimate 28-day all-cause mortality and to compare them between groups. Results from Cox models were reported in terms of Hazard Ratio (HR) with 90%CI.

Results

Study population

Data from 87 patients treated with CZA for *P. aeruginosa* infection in the participating centres were retrieved, 17 treated with CZA monotherapy and 70 with a combination of CZA and one or more antimicrobials. Detailed demographic and clinical representation of the groups is shown in Table 1.

While comorbidities were similar between the two groups, patients receiving CZA monotherapy were significantly older compared to those treated with CZA in combination.

Table 1 Demographic and clinical characteristics of the study population according to treatment group. Data are presented as median [IQR] for continuous measures, and *n* (%) for categorical measures, and *p* values refer to Wilcoxon rank sum test for continuous variables and to Fisher exact test for categorical variables

	CZA Monotherapy	CZA in Combination therapy	<i>p</i>
	<i>N</i> =17	<i>N</i> =70	
Age, median (IQR)	72 [62–75]	62 [52–69]	0.018
Female, <i>n</i> (%)	4 (23.5)	17 (24.3)	>0.9
Charlson comorbidity index median (IQR)	4 [3–5]	3 [2–5]	0.088
Missing=2			
DM II, <i>n</i> (%)	2 (12.5)	16 (22.9)	0.50
Missing=3			
COPD, <i>n</i> (%)	2 (11.8)	5 (7.2)	0.62
Missing=3			
Solid tumor, <i>n</i> (%)	1 (5.9)	9 (12.9)	0.68
Missing=2			
Hematological malignancy, <i>n</i> (%)	4 (23.5)	9 (12.9)	0.27
Missing=2			
Active immunosuppression, <i>n</i> (%)	3 (17.6)	16 (23.5)	0.75
Missing=4			
Cystic fibrosis, <i>n</i> (%)	1 (5.9)	7 (10.0)	>0.9
Missing=2			
Treatment with anti-pseudomonal drug in the last 12 months, <i>n</i> (%)	10 (58.8)	48 (71.6)	0.38
Missing=6			
eGFR (ml/min), <i>n</i> (%)			0.048
Missing=11			
=> 120	1 (7.1)	13 (20.0)	
50<=X<120	8 (57.1)	40 (61.5)	
30<=X<50	4 (28.6)	4 (6.2)	
15<=X<30	1 (7.1)	1 (1.5)	
X<15	0 (0.0)	7 (10.8)	
Renal replacement, <i>n</i> (%)	4 (23.5)	22 (31.4)	0.77
Missing=2			
CRRT, <i>n</i> (%)	4 (100.0)	21 (95.5)	>0.9
Sequential Organ Failure Assessment (SOFA) Score, median (IQR)	3 [1–5]	6 [3–9]	0.049
Missing=30			
ICU admission, <i>n</i> (%)	8 (47.1)	48 (68.6)	0.16
Missing=2			
Septic shock, <i>n</i> (%)	2 (11.8)	22 (31.9)	0.13
Missing=3			
Bloodstream infection, <i>n</i> (%)	8 (47.1)	20 (29.0)	0.16
Missing=3			
Source of infection, <i>n</i> (%)			0.19
HAP	4 (26.7)	12 (18.5)	
VAP	5 (33.3)	37 (56.9)	
cIAI	0 (0.0)	2 (3.1)	
UTI	1 (6.7)	2 (3.1)	
cUTI	2 (13.3)	1 (1.5)	
ABSSI	1 (6.7)	6 (9.2)	
Blood catheter	2 (13.3)	5 (7.7)	
Length of treatment, median (IQR)	12 [8–15]	14 [10–18]	0.12
Adequate source control, <i>n</i> (%)	7 (43.8)	28 (45.9)	>0.9
Renal adjustment for CZA, <i>n</i> (%)	7 (46.7)	24 (34.8)	0.39
CZA extended infusion, <i>n</i> (%)	8 (53.3)	38 (58.5)	0.78

DM Diabetes mellitus, COPD chronic obstructive pulmonary disease, eGFR estimated glomerular filtration rate, CRRT continuous renal replacement treatment, HAP healthcare acquired pneumonia, VAP ventilator associated pneumonia, cIAI complicated intra-abdominal infection, UTI urinary tract infection, cUTI complicated urinary tract infection, ABSSI acute bacterial skin and soft tissue infection, CZA ceftazidime/avibactam

Table 2 Comparison of characteristics used to create IPTW. Both raw and weighted descriptives are reported

	Unweighted analyses Hotelling t test: $p < 0.001$		Weighted analyses Hotelling t test: $p = 0.999$	
	Mono-therapy ($n = 17$)	Combination ($n = 70$)	Mono-therapy ($n = 11$)	Combination ($n = 33$)
Age, year (Mean, SD)	66 (17)	59 (15)	61 (20)	61 (11)
CCI, (Mean, SD)	4.2 (2.0)	3.3 (2.1)	3.2 (2.2)	3.9 (2.2)
Active immunosuppression Yes, (%)	18%	24%	0%	20%
Treatment with anti-pseudomonal drug in the last 12 months, (%)	59%	72%	56%	62%
ICU admission, (%)	47%	69%	81%	62%
HAP/VAP, (%)	29%	55%	42%	37%
Bloodstream infection, (%)	47%	29%	46%	44%
MDR, (%)	59%	48%	51%	59%
XDR, (%)	18%	13%	5%	8%
DTR, (%)	12%	19%	17%	12%

CCI Charlson comorbidity index, ICU Intensive Care Unit, VAP ventilator-associated pneumonia, MDR multidrug resistant, XDR extremely drug resistant, DTR difficult to treat

With regards to clinical presentation, the median SOFA score was significantly higher in patients receiving combination treatment compared to those receiving CZA monotherapy. Combination treatment was prescribed in 73% of the cases for hospital-acquired pneumonia HAP or VAP.

While patients in the monotherapy group presented more frequently a reduction in creatinine clearance, dose adjustment according to renal function was not significantly different in the two groups. On the other hand, patients in the combination treatment group presented more frequently with an augmented renal clearance. Detailed information regarding the antimicrobials used in combination is presented in the [Supplementary Materials](#).

Table 2 shows the comparison of variables used to create IPTW, before and after weighting. After weighting, there were no significant differences between the two treatment groups using Hotelling's t test ($p = 0.999$).

Microbiological results

Susceptibility results for the tested antimicrobials and proportions of MDR, XDR, and DTR isolates are shown in Table 3. For one single isolate, CZA MIC was not available, but it was included in the analysis since it was susceptible to ceftazidime alone.

Table 3 Antimicrobial resistance according to EUCAST clinical breakpoints between treatment groups. Results are expressed as count and percentages. p-values refer to Fisher exact test. MIC= minimum inhibitory concentration, MDR= multidrug resistant, XDR=extremely drug resistant, DTR= difficult to treat. MIC values are expressed as mg/L

	CZA Monotherapy $N = 17$	CZA in Combination therapy $N = 70$	p
Amikacin R, n (%)	5 (29.4)	9 (13.0)	0.19
Aztreonam R, n (%)	4 (23.5)	13 (18.8)	0.91
Not performed= 51			
Cefepime R, n (%)	6 (35.3)	36 (52.2)	0.20
Not performed= 16			
Ceftazidime R, n (%)	6 (35.3)	34 (49.3)	0.28
Not performed= 7			
Piperacillin/tazobactam R, n (%)	11 (64.7)	45 (65.2)	0.84
Ciprofloxacin R, n (%)	8 (50.0)	32 (47.1)	1.00
Colistin R, n (%)	0 (0.0)	1 (1.5)	0.10
Not performed= 8			
Tobramycin R, n (%)	1 (5.9)	10 (14.5)	0.078
Not performed= 30			
Imipenem R, n (%)	8 (47.1)	44 (64.7)	0.35
Not performed= 8			
Meropenem, n (%)			0.44
MIC ≤ 2	5 (31.2)	16 (23.2)	
$2 < \text{MIC} \leq 8$	1 (6.2)	14 (20.3)	
MIC > 8	10 (62.5)	39 (56.5)	
Ceftolozane/tazobactam R, n (%)	1 (5.9)	8 (11.6)	0.20
Not performed= 13			
Ceftazidime/avibactam, MIC, n (%)			0.36
0.5	0 (0)	4 (5.7)	
1	3 (18.7)	1 (1.4)	
2	6 (37.5)	2 (2.9)	
4	3 (18.7)	15 (21.4)	
8	4 (25)	24 (34.3)	
MDR, n (%)	10 (58.8%)	33 (47.8%)	0.59
XDR, n (%)	3 (17.6%)	9 (13.2%)	0.70
DTR, n (%)	2 (11.8%)	13 (19.1%)	0.72

Primary endpoint: 14-day clinical cure

After 14 days, clinical cure was reached in 40/69 (58%) patients treated with the combination of CZA and in 8/17 (47%) patients treated with CZA monotherapy (RD=-0.11, 90%CI: -0.32 to 0.10).

After weighting patients according to the IPTW, CZA monotherapy did not reach the clinical equipoise to combination therapy for 14-day clinical cure (36.2% vs. 63.8%, RD= -0.28, 90%CI: -0.41 to -0.14).

Table 4 Risk differences for secondary outcomes according to treatment groups. Outcomes are presented as counts and percentage. 90%CI

Outcome	Monotherapy	Combination	Risk difference	90% CI for risk difference
56 days microbiological failure	7 (43.7%)	21 (30.4%)	0.13	-0.09 to 0.36
56 days infection relapse	2 (13.3%)	12 (17.07%)	-0.05	-0.15 to 0.06
CZA resistance	1 (5.9%)	5 (7.5%)	-0.02	-0.10 to 0.07

Secondary outcomes

At univariable analysis, two groups were comparable for 56 days microbiological failure (RD=0.13, 90%CI: -0.09 to 0.36), 56 days infection relapse (RD= -0.05, 90%CI: -0.15 to 0.06), and occurrence of CZA resistance (RD= -0.02, 90%CI: -0.10 to 0.07), as described in Table 4.

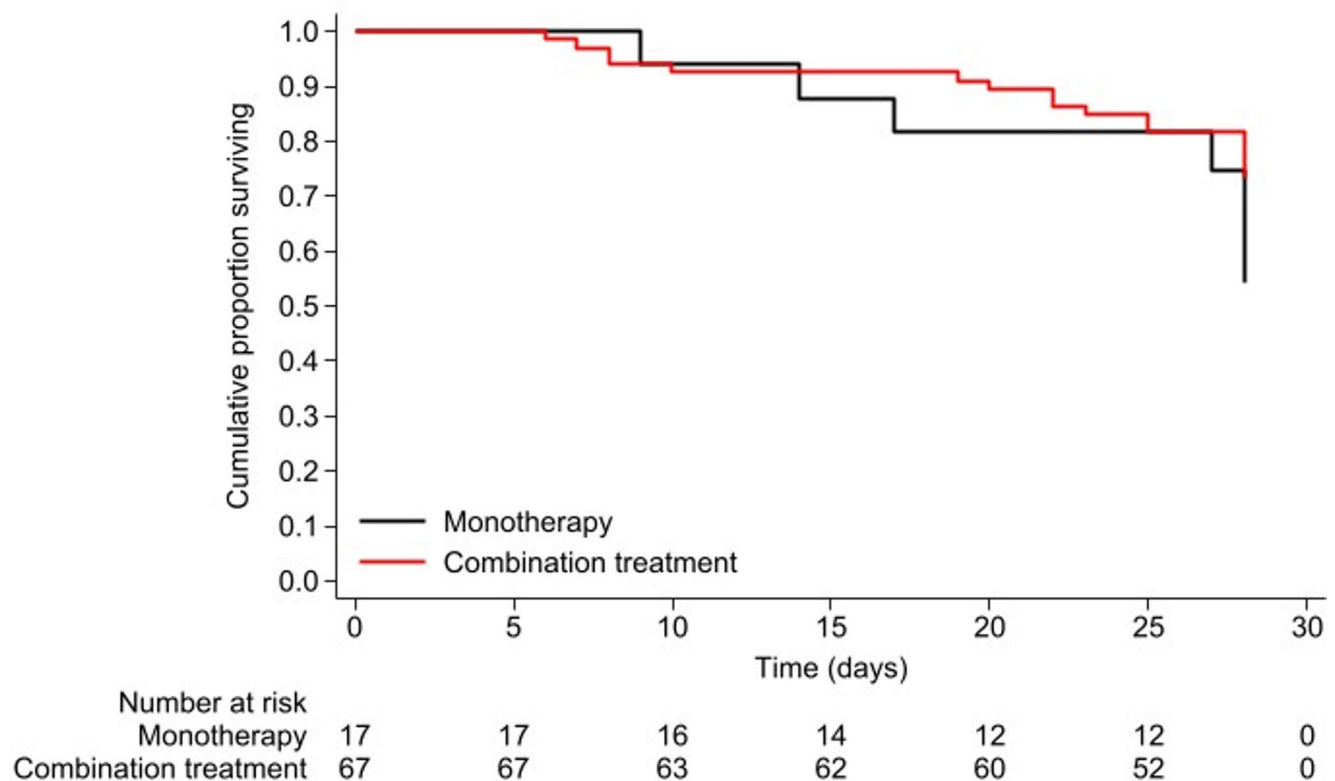
After 28 days, 24 patients had died: 7 (41.2%) treated with CZA monotherapy and 17 (24.3%) treated with CZA in combination therapy. Again, 28-day all-cause mortality was comparable between groups (Fig. 1, HR=0.56, 90%CI: 0.23, 1.35).

Discussion

We found that CZA monotherapy was associated with lower 14-day clinical cure in patients with PA infections when compared to CZA combination with other active antimicrobials. We did not find significant differences between the two regimens for 28-day mortality, 56-day microbiological failure, 56-day infection relapse, and emergence of CZA-resistant isolates. In our cohort, combination therapy was prescribed frequently in patients with pneumonia, higher SOFA scores, and increased renal clearance. On the other hand, patients receiving CZA monotherapy were older and presented with lower renal clearance.

PA represents a significant threat due to the rising incidence of antimicrobial resistance and the associated high mortality in systemic infections [1, 2]. CZA is a key therapeutic option, especially against MDR strains, similar to ceftolozane/tazobactam (C/T) [9]. However, comparative studies between these agents have provided mixed results. While most research shows comparable clinical outcomes, one study reported that C/T resulted in lower recurrence rates and greater cost-effectiveness in patients with PA pneumonia compared to CZA [31–33].

Results from a multicentric cohort study by Shields et al. comparing the two agents showed a significantly higher

**Fig. 1** Kaplan-Meier survival curve according to CZA in monotherapy or combination treatment

odds of clinical cure in patients treated with C/T compared to CZA for MDR PA infections, and may provide good evidence to support the role of C/T as the first-line agent for these infections [33]. Nevertheless, CZA remains a good option, and it is preferred for isolates resistant to C/T, as it was for 9% of the isolates in this cohort.

Although several studies have demonstrated no significant survival benefit from combination therapy in serious PA infections, including VAP and BSI [34–36], we found that most patients in our cohort—particularly those with lower respiratory tract infections and higher SOFA scores—received combination therapy. Several factors may explain clinicians' preference for combination therapy in these patients. Although monotherapy is recommended for definitive treatment to avoid toxicity in the absence of clear benefits [21], one retrospective study suggested that empiric combination therapy could be associated with earlier appropriate treatment in patients with VAP [37]. This is important, as inappropriate empiric treatment in severe infections is linked to increased mortality and clinical failure [35]. However, it is worth noting that these results predate the introduction of new β -lactam/ β -lactamase inhibitors, which might now reduce the impact of combination therapy on empiric coverage.

Furthermore, β -lactam concentrations in the epithelial lining fluid are lower than in plasma, leading to potential sub-optimal antimicrobial concentrations at the infection site, especially in HAP and VAP [38]. Combining two active antibacterial agents can theoretically reduce the risk of clinical failure and selection for antimicrobial resistance due to sub-optimal antimicrobial concentrations. This concern is even greater in patients with augmented renal clearance, who are at increased risk of antimicrobial underexposure, as reflected in our cohort, where patients receiving combination treatment had more frequent augmented renal clearance [39].

More than half of the patients in our cohort received CZA as a prolonged or continuous infusion. Prolonged infusion of time-dependent antibiotics such as CZA has been shown to reduce mortality in septic patients [40]. Furthermore, it has been proposed as a strategy to overcome resistance development by targeting aggressive pharmacokinetic/pharmacodynamic (PK/PD) targets, such as a free fraction 100% time above 4 times the MIC [41].

PK/PD optimization is particularly appealing for CZA, which is prone to subtherapeutic exposure due to an unfavourable beta-lactam/beta-lactamase ratio of 1:4 [42]. This strategy has been somewhat alternative to combination therapy, as a study evaluating a pharmacy-driven stewardship intervention to improve target attainment for KPC producing *K. pneumoniae* infection has shown an improvement in clinical and microbiological cure with a reduction in the use of combination therapy [43]. Nevertheless, limited

access to therapeutic drug monitoring restricts widespread implementation.

Conversely, patients receiving monotherapy were older and more likely to have reduced renal clearance, reflecting a concern about potential toxicity, particularly nephrotoxicity, in elderly patients. Aminoglycosides and colistin, which were frequently part of the combination regimen, are associated with significant renal toxicity [44, 45].

Interestingly, the proportion of patients with active immunosuppression was similar between the two treatment arms. This subgroup is prone to developing serious PA infections and to being exposed to multiple antibiotic courses [26]; nonetheless, these concerns were not associated with an inclination to choose combination over CZA monotherapy.

We found a notable difference in 14-day clinical cure rates favouring combination therapy. This contrasts with several observational studies comparing monotherapy and combination therapy in serious PA infections, which mainly focused on mortality as the primary outcome [34–36]. Mortality is an objective and highly relevant endpoint, especially given the high mortality rates associated with these infections. In contrast, clinical cure—while more subjective—may reveal smaller differences between treatment groups. To minimize bias, we used rigorous and standardized definitions of clinical cure.

Our findings are at odds with some previous studies on CZA monotherapy versus combination therapy for PA [19, 20]. For instance, Xu et al. reported no significant differences in clinical cure between the two approaches in patients with HAP and VAP [19]. However, the lack of pre-specified time points for evaluating clinical cure makes the comparison difficult. It is possible that combination treatment could offer benefits in the initial phase of treatment, without impacting overall cure rates. Another observational study by Corbella et al. found lower 14-day clinical cure rates with CZA-containing combination regimens, which the authors attributed to residual confounding factors and increased toxicity, although only 3 patients in their cohort experienced antimicrobial-related adverse events [20].

In our cohort, 28-day all-cause mortality was higher than described in the aforementioned studies [19, 20]. This can be explained by the higher proportion of critically ill patients, who accounted for more than half of our sample. Similarly, we did not find a significant difference in mortality between patients treated with CZA monotherapy and patients treated with CZA in combination with other active antimicrobials.

While suppression of further resistance is a theoretical benefit of combination therapy, the proportion of MDR, XDR, and DTR strains at baseline was similar between the two treatment arms. In particular, more than half of the isolates were characterized as MDR, and 14% were XDR. Similarly, 17% of the isolates were characterized as DTR.

The emergence of resistance after treatment with CZA varies from 0% to 40% across studies. Epidemiological differences and differences in resistance definitions could explain this variability [11, 46, 47]. In our cohort, almost 7% of patients developed subsequent resistance, defined as MIC values above the EUCAST clinical breakpoint, without differences between the two groups [28].

Emerging resistance to CZA is mainly mediated by mutations in *ampC*, combined with the expression of efflux pumps [12, 14, 46–49]. These mutations may lead to increased MIC to other cephalosporins, such as C/T and cefiderocol [50, 51]. A recent comparative study showed lower rates of emerging resistance following treatment with C/T compared to CZA, possibly due to the higher stability of ceftolozane to cephalosporinases [46]. The low rate of resistance in our study can be explained by the smaller number of MDR isolates compared to other cohorts, as several co-resistance mechanisms occur in these isolates and make them more prone to develop further resistance [46, 47, 52].

Our study has some limitations. Firstly, the small sample size limits the generalizability of our findings. Despite including patients from multiple centres with diverse infection types, the overall sample is consistent with previous studies of CZA for PA. Secondly, a shortage of C/T during the study period may have led to increased CZA use, complicating future studies on this topic. Thirdly, the retrospective and observational nature of the study also introduces the possibility of missing data and residual confounding.

Although the presence of missing data was overall limited, the relevant proportion of missing values for the SOFA score precluded the inclusion of an important variable of acute illness severity in the propensity score.

Despite the efforts to reduce confounding through the application of IPTW, we cannot exclude the presence of residual confounding, especially due to the limited sample and unbalanced groups.

The inclusion of several different types of infection could introduce substantial heterogeneity, given the limited sample size. Since the majority of patients presented with HAP/VAP, the results may be driven mainly by this subgroup, although distribution within treatment groups was balanced using propensity score.

Since this study was pathogen-directed, microbiological identification of PA was required to define clinical relapse, thus possibly excluding patients presenting with recurrent signs/symptoms of infection that do not receive adequate microbiological work-up.

Furthermore, the impact of COVID-19 cannot be overlooked, as it complicates the evaluation of clinical cure in patients with lower respiratory tract infections and viral pneumonia. Finally, cystic fibrosis was present in 10% of patients in our cohort, further complicating cure evaluation in these cases.

Our findings suggest that the debate between monotherapy and combination therapy for PA infections, particularly with CZA, remains unresolved despite the lack of a clear survival benefit for combination therapy. Due to the study design and the aforementioned limitations, these results should be considered hypothesis-generating.

While monotherapy offers the benefit of lower toxicity, especially in elderly patients, combination therapy—at least in the early phase—might be associated with higher clinical cure rates by improving empirical coverage or reducing bacterial burden. Although randomized controlled trials on this topic are unlikely, large cohorts of prospectively collected data, particularly focusing on lower respiratory tract infections, may help clarify the optimal treatment approach.

Supplementary Information The online version contains supplementary material available at <https://doi.org/10.1007/s10096-026-05599-x>.

Acknowledgements None.

Author contributions All authors contributed to the study conception and design. Data collection was performed by Pietro Valsecchi, Simona Biscarini, Cecilia Azzarà, Nicolò Corti, Leonardo Francesco Rezzonico, Silvia Pontiggia, Arianna Lippi, Matteo Giovanni Lupi, Giacomo Casalini and Alessandra Calabresi. Analysis was performed by Virginia Valeria Ferretti, Giulia Gambini and Catherine Klersy. The first draft of the manuscript was written by Pietro Valsecchi and Marta Colaneri. All authors commented on previous versions of the manuscript. All authors read and approved the final manuscript.

Funding None declared.

Data availability The datasets generated during and/or analysed during the current study are available from the corresponding author on reasonable request.

Declarations

Ethical approval The study was approved by the Internal Review Board of all the participating centres (N° 0025169/22). Results were reported following the STROBE (Strengthening the Reporting of Observational Studies in Epidemiology) guidelines. Informed consent was waived due to the retrospective nature of the study.

Competing interests The authors declare no competing interests.

Transparency The authors declare no conflicts of interest.

Sequence information Not applicable.

References

1. Reynolds D, Kollef M (2021) The Epidemiology and Pathogenesis and Treatment of *Pseudomonas aeruginosa* Infections: An Update. *Drugs* 81:2117–2131. <https://doi.org/10.1007/s40265-021-01635-6>
2. Suetens C, Latour K, Kärki T, Ricchizzi E, Kinross P, Moro ML et al (2018) Prevalence of healthcare-associated infections,

- estimated incidence and composite antimicrobial resistance index in acute care hospitals and long-term care facilities: results from two European point prevalence surveys, 2016 to 2017. *Eurosurveillance* 23. <https://doi.org/10.2807/1560-7917.ES.2018.23.46.1800516>
3. WHO Regional Office for Europe/European Centre for Disease Prevention and Control (2022) Antimicrobial resistance surveillance in Europe 2022–2020 data. WHO Regional Office for Europe, Copenhagen
 4. Karruli A, Catalini C, D'Amore C, Foglia F, Mari F, Harxhi A et al (2023) Evidence-based treatment of *Pseudomonas aeruginosa* infections: a critical reappraisal. *Antibiotics* 12. <https://doi.org/10.3390/antibiotics12020399>
 5. WHO Bacterial Priority Pathogens List (2024) 2024: bacterial pathogens of public health importance to guide research, development and strategies to prevent and control antimicrobial resistance. Geneva: World Health Organization. Licence: CC BY-NC-SA 3.0 IGO
 6. Magiorakos AP, Srinivasan A, Carey RB, Carmeli Y, Falagas ME, Giske CG et al (2012) Multidrug-resistant, extensively drug-resistant and pandrug-resistant bacteria: An international expert proposal for interim standard definitions for acquired resistance. *Clin Microbiol Infect* 18:268–281. <https://doi.org/10.1111/j.1469-0691.2011.03570.x>
 7. Kadri SS, Adjemian J, Lai YL, Spaulding AB, Ricotta E, Rebecca Prevots D et al (2018) Difficult-to-treat resistance in gram-negative bacteremia at 173 US hospitals: Retrospective cohort analysis of prevalence, predictors, and outcome of resistance to all first-line agents. *Clin Infect Dis* 67:1803–1814. <https://doi.org/10.1093/cid/ciy378>
 8. Zhanel GG, Lawson CD, Adam H, Schweizer F, Zelenitsky S, Lagacé-Wiens PRS et al (2013) Ceftazidime-Avibactam: a Novel Cephalosporin/ β -lactamase Inhibitor Combination. *Drugs* 73:159–177. <https://doi.org/10.1007/s40265-013-0013-7>
 9. Karlowsky JA, Lob SH, Bauer KA, Esterly J, Siddiqui F, Young K et al (2024) Activity of ceftolozane/tazobactam, imipenem/relebactam and ceftazidime/avibactam against clinical Gram-negative isolates - SMART United States 2019–21. *JAC Antimicrob Resist* 6. <https://doi.org/10.1093/jacamr/dlad152>
 10. Wise MG, Karlowsky JA, Lemos-Luengas EV, Valdez RR, Sahn DF (2023) Epidemiology and in vitro activity of ceftazidime-avibactam and comparator agents against multidrug-resistant isolates of Enterobacterales and *Pseudomonas aeruginosa* collected in Latin America as part of the ATLAS surveillance program in 2015–2020. *Brazilian J Infect Dis* 27. <https://doi.org/10.1016/j.bjid.2023.102759>
 11. Gaibani P, Giani T, Bovo F, Lombardo D, Amadesi S, Lazzarotto T et al (2022) Resistance to ceftazidime/avibactam, meropenem/vaborbactam and imipenem/relebactam in Gram-Negative MDR Bacilli: molecular mechanisms and susceptibility testing. *Antibiotics* 11. <https://doi.org/10.3390/antibiotics11050628>
 12. Mojica MF, De La Cadena E, García-Betancur JC, Porras J, Novoa-Cacedo I, Páez-Zamora L et al (2023) Molecular mechanisms of resistance to ceftazidime/avibactam in clinical isolates of enterobacterales and *Pseudomonas aeruginosa* in Latin American Hospitals. *MSphere* 8. <https://doi.org/10.1128/msphere.00651-22>
 13. Fraile-Ribot PA, Mulet X, Cabot G, Del Barrio-Tofiño E, Juan C, Pérez JL, Oliver A (2017) *Vivo* Emergence of Resistance to Novel Cephalosporin- β -Lactamase Inhibitor Combinations through the Duplication of Amino Acid D149 from OXA-2 β -Lactamase (OXA-539) in Sequence Type 235 *Pseudomonas aeruginosa*. *Antimicrob Agents Chemother* 61(9):e01117–e01117. <https://doi.org/10.1128/AAC.01117-17> PMID: 28674059; PMCID: PMC5571340
 14. Compain F, Debray A, Adjadj P, Dorcène D, Arthur M (2020) Ceftazidime-avibactam resistance mediated by the N346Y substitution in various AmpC β -lactamases. *Antimicrob Agents Chemother* 64. <https://doi.org/10.1128/AAC.02311-19>
 15. Slater CL, Winogrodzki J, Fraile-Ribot PA, Oliver A, Khajehpour M, Mark BL (2020) Adding insult to injury: mechanistic basis for how AmpC mutations allow *Pseudomonas aeruginosa* to accelerate cephalosporin hydrolysis and evade avibactam. *Antimicrob Agents Chemother* 64. <https://doi.org/10.1128/AAC.00894-20>
 16. Oliver A, Rojo-Molinero E, Arca-Suarez J, Bešli Y, Bogaerts P et al (2024) *Pseudomonas aeruginosa* antimicrobial susceptibility profiles, resistance mechanisms and international clonal lineages: update from ESGARS-ESCMID/ISARPAE Group. *Clin Microbiol Infect* 30(4):469–480. <https://doi.org/10.1016/j.cmi.2023.12.026>
 17. Aslan AT, Ezure Y, Horcajada JP, Harris PNA, Paterson DL (2023) In vitro, in vivo and clinical studies comparing the efficacy of ceftazidime-avibactam monotherapy with ceftazidime-avibactam-containing combination regimens against carbapenem-resistant Enterobacterales and multidrug-resistant *Pseudomonas aeruginosa* isolates or infections: a scoping review. *Front Med (Lausanne)* 10. <https://doi.org/10.3389/fmed.2023.1249030>
 18. Papp-Wallace KM, Zeiser ET, Becka SA, Park S, Wilson BM, Winkler ML et al (2020) Ceftazidime-Avibactam in Combination with Fosfomycin: A Novel Therapeutic Strategy against Multidrug-Resistant *Pseudomonas aeruginosa*. *J Infect Dis* 221:666–676. <https://doi.org/10.1093/infdis/jiz149>
 19. Xu C, Zeng F, Huang Y, Xu Q, Yang Y, Gong W et al (2024) Clinical efficacy of ceftazidime/avibactam combination therapy for severe hospital-acquired pulmonary infections caused by carbapenem-resistant and difficult-to-treat *Pseudomonas aeruginosa*. *Int J Antimicrob Agents* 63. <https://doi.org/10.1016/j.ijantimicag.2023.107021>
 20. Corbella L, Boán J, San-Juan R, Fernández-Ruiz M, Carretero O, Lora D et al (2022) Effectiveness of ceftazidime-avibactam for the treatment of infections due to *Pseudomonas aeruginosa*. *Int J Antimicrob Agents* 59. <https://doi.org/10.1016/j.ijantimicag.2021.106517>
 21. Paul M, Carrara E, Retamar P, Tängdén T, Bitterman R, Bonomo RA et al (2022) European Society of Clinical Microbiology and Infectious Diseases (ESCMID) guidelines for the treatment of infections caused by multidrug-resistant Gram-negative bacilli (endorsed by European society of intensive care medicine). *Clin Microbiol Infect* 28:521–547. <https://doi.org/10.1016/j.cmi.2021.11.025>
 22. Tamma PD, Aitken SL, Bonomo RA, Mathers AJ, van Duin D, Clancy CJ (2022) Infectious Diseases Society of America 2022 Guidance on the Treatment of Extended-Spectrum β -lactamase Producing Enterobacterales (ESBL-E), Carbapenem-Resistant Enterobacterales (CRE), and *Pseudomonas aeruginosa* with Difficult-to-Treat Resistance (DTR-P. *aeruginosa*). *Clin Infect Dis* 75(2):187–212. <https://doi.org/10.1093/cid/ciac268>. PMID: 35439291; PMCID: PMC9890506
 23. National Center for Emerging and Zoonotic Infectious Diseases (U.S.) (2023) Division of Healthcare Quality Promotion, National Healthcare Safety Network; CDC/NHSN surveillance definitions for specific types of infections Published Date: January 2023 Pages in Document: 30 numbered pages Source : National Healthcare Safety Network (NHSN) patient safety component manual. Chapter 17. <https://stacks.cdc.gov/view/cdc/127235>
 24. Charlson ME, Pompei P, Ales KL, MacKenzie CR (1987) A new method of classifying prognostic comorbidity in longitudinal studies: development and validation. *J Chronic Dis* 40(5):373–83. [https://doi.org/10.1016/0021-9681\(87\)90171-8](https://doi.org/10.1016/0021-9681(87)90171-8)
 25. Sadikot RT, Blackwell TS, Christman JW, Prince AS (2005) Pathogen-host interactions in *Pseudomonas aeruginosa* pneumonia. *Am J Respir Crit Care Med* 171:1209–1223. <https://doi.org/10.1164/rccm.200408-1044SO>

26. Babich T, Naucler P, Valik JK, Giske CG, Benito N, Cardona R et al (2020) Risk factors for mortality among patients with *Pseudomonas aeruginosa* bacteraemia: a retrospective multicentre study. *Int J Antimicrob Agents* 55. <https://doi.org/10.1016/j.ijantimicag.2019.11.004>
27. Singer M, Deutschman CS, Seymour C, Shankar-Hari M, Annane D, Bauer M et al (2016) The third international consensus definitions for sepsis and septic shock (sepsis-3). *JAMA - J Am Med Association* 315:801–810. <https://doi.org/10.1001/jama.2016.0287>
28. The European Committee on Antimicrobial Susceptibility Testing (2023) Breakpoint tables for interpretation of MICs and zone diameters. Version 13.0. <http://www.eucast.org>
29. The European Committee on Antimicrobial Susceptibility Testing (2020) Implementation and use of the 2020 revised aminoglycoside breakpoints. <http://www.eucast.org>
30. The European Committee on Antimicrobial Susceptibility Testing (2023) Fosfomycin intravenous: rationale for EUCAST clinical breakpoints. Version 1.0. <http://www.eucast.org>
31. Lodise TP, Obi EN, Watanabe AH, Yucel E, Min J, Nathanson BH (2024) Comparative evaluation of early treatment with ceftolozane/tazobactam versus ceftazidime/avibactam for non-COVID-19 patients with pneumonia due to multidrug-resistant *Pseudomonas aeruginosa*. *J Antimicrob Chemother* 79(11):2954–964. <https://doi.org/10.1093/jac/dkac313>
32. Almangour TA, Ghonem L, Alassiri D, Aljurbua A, Al Musawa M et al (2023) Ceftolozane-Tazobactam Versus Ceftazidime-Avibactam for the Treatment of Infections Caused by Multidrug-Resistant *Pseudomonas aeruginosa*: a Multicenter Cohort Study. *Antimicrob Agents Chemother* 67:e00405–e00423. <https://doi.org/10.1128/aac.00405-23>
33. Shields RK, Abbo LM, Ackley R, Aitken SL, Albrecht B, Babiker A et al (2024) Effectiveness of ceftazidime–avibactam versus ceftolozane–tazobactam for multidrug-resistant *Pseudomonas aeruginosa* infections in the USA (CACTUS): a multicentre, retrospective, observational study. *Lancet Infect Dis*. [https://doi.org/10.1016/S1473-3099\(24\)00648-0](https://doi.org/10.1016/S1473-3099(24)00648-0)
34. Kim YJ, Jun YH, Kim YR, Park KG, Park YJ, Kang JY, Kim SI (2014) Risk factors for mortality in patients with *Pseudomonas aeruginosa* bacteremia; retrospective study of impact of combination antimicrobial therapy. *BMC Infect Dis* 14:161. <https://doi.org/10.1186/1471-2334-14-161>. PMID: 24655422; PMCID: PMC3994322
35. Peña C, Suarez C, Ocampo-Sosa A, Murillas J, Almirante B, Pomar V et al (2013) Effect of adequate single-drug vs combination antimicrobial therapy on mortality in *Pseudomonas aeruginosa* bloodstream infections: A post hoc analysis of a prospective cohort. *Clin Infect Dis* 57:208–216. <https://doi.org/10.1093/cid/cit223>
36. Fouchier A, Dessalle T, Tuffet S, Federici L, Dahyot-Fizelier C, Barbier F et al (2023) Association between combination antibiotic therapy as opposed as monotherapy and outcomes of ICU patients with *Pseudomonas aeruginosa* ventilator-associated pneumonia: an ancillary study of the iDIAPASON trial. *Crit Care* 27:211. <https://doi.org/10.1186/s13054-023-04457-y>
37. Garnacho-Montero J, Sa-Borges M, Sole-Violan J, Barcenilla F, Escoreca-Ortega A et al (2007) Optimal management therapy for *Pseudomonas aeruginosa* ventilator-associated pneumonia: an observational, multicenter study comparing monotherapy with combination antibiotic therapy. *Crit Care Med* 35(8):1888–95. <https://doi.org/10.1097/01.CCM.0000275389.31974.22>
38. Drwiega EN, Rodvold KA (2022) Penetration of Antibacterial Agents into Pulmonary Epithelial Lining Fluid: An Update. *Clin Pharmacokinet* 61(1):17–46. <https://doi.org/10.1007/s40262-021-01061-7>. Epub 2021 Oct 15. PMID: 34651282; PMCID: PMC8516621
39. Xu Y, Tang J, Yuan B, Luo X, Liang P, Liu N et al (2024) A descriptive pharmacokinetic/pharmacodynamic analysis of ceftazidime-avibactam in a case series of critically ill patients with augmented renal clearance. *Pharmacol Res Perspect* 12. <https://doi.org/10.1002/prp2.1163>
40. Abdul-Aziz MH, Hammond NE, Brett SJ, Cotta MO, De Waele JJ et al (2024) Prolonged vs Intermittent Infusions of β -Lactam Antibiotics in Adults With Sepsis or Septic Shock: A Systematic Review and Meta-Analysis. *JAMA* 332(8):638–648 PMID: 38864162; PMCID: PMC11170459
41. Sumi CD, Heffernan AJ, Lipman J, Roberts JA, Sime FB (2019) What antibiotic exposures are required to suppress the emergence of resistance for gram-negative bacteria? A systematic review. *Clinical Pharmacokinetics* 58(11):1407–1443. <https://doi.org/10.1007/s40262-019-00791-z>
42. Gatti M, Rinaldi M, Bonazzetti C, Gaibani P, Giannella M, Viale P, Pea F (2023) Could an optimized joint pharmacokinetic/pharmacodynamic target attainment of continuous infusion ceftazidime-avibactam be a way to avoid the need for combo therapy in the targeted treatment of deep-seated DTR Gram-negative infections? *Antimicrob Agents Chemother* 67:e00969–e00923. <https://doi.org/10.1128/aac.00969-23>
43. Gatti M, Rinaldi M, Cojutti PG, Bonazzetti C, Siniscalchi A et al (2025) A pre-post quasi-experimental study of antimicrobial stewardship exploring the impact of a multidisciplinary approach aimed at attaining an aggressive joint pharmacokinetic/pharmacodynamic target with ceftazidime/avibactam on treatment outcome of KPC-producing *Klebsiella pneumoniae* infections and on ceftazidime/avibactam resistance development. *Antimicrob Agents Chemother* 69:e00488–e00425. <https://doi.org/10.1128/aac.00488-25>
44. Eljaaly K, Bidell MR, Gandhi RG, Alshehri S, Enani MA, Al-Jedai A et al (2021) Colistin nephrotoxicity: meta-analysis of randomized controlled trials. *Open Forum Infect Dis* 8. <https://doi.org/10.1093/ofid/ofab026>
45. Wargo KA, Edwards JD (2014) Aminoglycoside-induced nephrotoxicity. *J Pharm Pract* 27:573–577. <https://doi.org/10.1177/0897190014546836>
46. Hareza DA, Cosgrove SE, Bonomo RA, Dzintars K, Karaba SM, Hawes AM et al (2024) Clinical outcomes and emergence of resistance of *Pseudomonas aeruginosa* infections treated with ceftolozane-tazobactam versus ceftazidime-avibactam. *Antimicrob Agents Chemother*. <https://doi.org/10.1128/aac.00907-24>
47. Shah S, Kline EG, Haidar G, Squires KM, Pogue JM, McCreary EK et al (2024) Rates of Resistance to Ceftazidime-Avibactam and Ceftolozane-Tazobactam Among Patients Treated for Multidrug-Resistant *Pseudomonas aeruginosa* Bacteremia or Pneumonia. *Clin Infect Dis*. <https://doi.org/10.1093/cid/ciae332>
48. Lopez-Montesinos I, Montero MM, Domene-Ochoa S, López-Causapé C, Echeverria D, Sorli L et al (2022) Suboptimal concentrations of ceftazidime/avibactam (CAZ-AVI) may select for CAZ-AVI resistance in extensively drug-resistant *Pseudomonas aeruginosa*: in vivo and in vitro evidence. *Antibiotics* 11. <https://doi.org/10.3390/antibiotics11111456>
49. Papp-Wallace KM, Mack AR, Taracila MA, Bonomo RA (2020) Resistance to Novel β -Lactam- β -Lactamase Inhibitor Combinations: The Price of Progress. *Infect Dis Clin North Am* 34:773–819. <https://doi.org/10.1016/j.idc.2020.05.001>
50. Shields RK, Kline EG, Squires KM, Van Tyne D, Doi Y (2023) In vitro activity of cefiderocol against *Pseudomonas aeruginosa* demonstrating evolved resistance to novel β -lactam/ β -lactamase inhibitors. *JAC Antimicrob Resist* 5. <https://doi.org/10.1093/jacamr/dlad107>

51. Gomis-Font MA, Clari MA, López-Causapé C, Navarro D, Oliver A (2024) Emergence of cefiderocol resistance during ceftazidime/ avibactam treatment caused by a large genomic deletion, including ampD and piuCD genes, in *Pseudomonas aeruginosa*. *Antimicrob Agents Chemother* 68. <https://doi.org/10.1128/aac.01192-23>
52. Appaneal HJ, Caffrey AR, Lopes V, Piehl EC, Puzniak LA, LaPlante KL (2022) Assessing rates of co-resistance and patient outcomes in multidrug-resistant *pseudomonas aeruginosa*. *Microbiol Spectr* 10. <https://doi.org/10.1128/spectrum.02336-22>

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Springer Nature or its licensor (e.g. a society or other partner) holds exclusive rights to this article under a publishing agreement with the author(s) or other rightsholder(s); author self-archiving of the accepted manuscript version of this article is solely governed by the terms of such publishing agreement and applicable law.

Authors and Affiliations

Pietro Valsecchi¹  · Marta Colaneri² · Andrea Lombardi^{3,4} · Simona Biscarini³ · Cecilia Azzarà^{3,4} · Alessandra Bandera^{3,4} · Nicolò Corti⁵ · Paolo Bonfanti⁵ · Leonardo Francesco Rezzonico^{6,7} · Giovanna Travi⁶ · Silvia Pontiggia⁸ · Stefania Piconi⁸ · Arianna Lippi^{9,10} · Mario Tumbarello^{9,10} · Matteo Giovanni Lupi¹¹ · Angelo Pan¹¹ · Giacomo Casalini^{2,12} · Spinello Antinori² · Andrea Gori² · Alessandra Calabresi¹³ · Guido Chichino¹³ · Virginia Valeria Ferretti¹⁴ · Giulia Gambini¹⁴ · Catherine Klersy¹⁴ · Raffaele Bruno^{1,7}

✉ Raffaele Bruno
raffaele.bruno@unipv.it

Pietro Valsecchi
p.valsecchi@smatteo.pv.it

¹ Struttura Complessa Malattie Infettive I, Fondazione I.R.C.C.S. Policlinico San Matteo, Pavia, Italy

² Dipartimento di Scienze Biomediche e Cliniche, Università degli Studi di Milano, Milan, Italy

³ Infectious Diseases Unit, Fondazione IRCCS Ca' Granda Ospedale Maggiore Policlinico, Milan, Italy

⁴ Department of Pathophysiology and Transplantation, University of Milan, Milan, Italy

⁵ University of Milano-Bicocca, Fondazione IRCCS San Gerardo dei Tintori, Milan, Italy

⁶ Division of Infectious Diseases, ASST Grande Ospedale Metropolitano Niguarda, Milan, Italy

⁷ Dipartimento di Scienze Clinico-chirurgiche, Diagnostiche e Pediatriche, Università degli Studi di Pavia, Pavia, Italy

⁸ Infectious Diseases Unit, A. Manzoni Hospital, Via dell'Eremo 9, Lecco, Italy

⁹ Infectious and Tropical Diseases Unit, Department of Medical Sciences, University Hospital of Siena, Siena, Italy

¹⁰ Department of Medical Biotechnologies, University of Siena, Siena, Italy

¹¹ Infectious Disease Unit, ASST Cremona, Cremona, Italy

¹² III Division of Infectious Diseases, Luigi Sacco Hospital, ASST Fatebenefratelli Sacco, Milan, Italy

¹³ SOC Malattie Infettive, AOU SS Antonio e Biagio e C. Arrigo, Alessandria, Italy

¹⁴ SSD Biostatistica e Clinical Trial Center, Fondazione I.R.C.C.S. Policlinico San Matteo, Pavia, Italy